

Figure S1. Detection of the GFP-Ade16 fusion proteins.

The expression of the GFP-Ade16 fusion protein in TBL308 was confirmed by Western blotting.

M: protein marker.

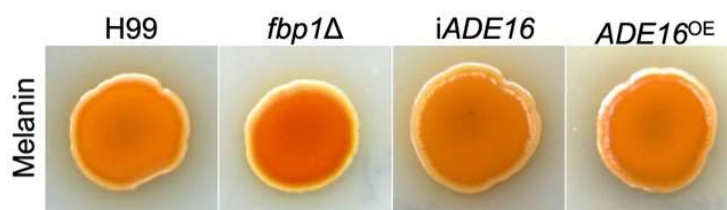


Figure S2. Ade16 is not involved in melanin production of *C. neoformans*.

Melanin production by the *C. neoformans* *ADE16* interference or overexpression strains was induced on Niger seed plates and photographed after incubation at 37°C for two days.

Table S1. Strains and plasmids used in this study

Strains/plasmids	Genotype/properties	Source/reference
<i>E. coli</i>		
DH5α	cloning strain	
<i>C. neoformans</i>		
H99	<i>MATα</i>	Perfect et al., 1993
KN99a	<i>MATa</i>	Nielsen et al., 2003
TBL3	<i>MATα fbp1Δ::NEO</i>	In this study
TBL81	<i>MATα Fbp1-Flag::URA5</i>	In this study
TBL248	<i>MATα Fbp1-Flag::URA5 P<sub>CTR4</sub>-Ade16-HA::NAT</i>	In this study
TBL264	<i>MATα P<sub>CTR4</sub>-Ade16-HA::NAT</i>	In this study
TBL265	<i>MATα fbp1Δ::NEO P<sub>CTR4</sub>-Ade16-HA::NAT</i>	In this study
TBL270	<i>MATα P<sub>CTR4</sub>-Ade16::NAT</i>	In this study
TBL288	<i>MATα P<sub>ACTIN</sub>-Ade16-HA::NAT</i>	In this study

TBL302	<i>MATa P<sub>ACTIN</sub>-Ade16-HA::NAT</i>	In this study
TBL308	<i>MATa P<sub>H3</sub>-GFP-Ade16::NAT</i>	In this study
TBL309	<i>MATa P<sub>H3</sub>-GFP-Ade16::NAT</i>	In this study
TBL310	<i>MATa P<sub>ADE16</sub>-mCherry::NAT</i>	In this study
TBL378	<i>MATa P<sub>ADE16</sub>-mCherry::NAT</i>	In this study
TBL414	<i>MATa iADE16::NAT</i>	In this study
TBL415	<i>MATa iADE16::NAT</i>	In this study
TBL445	<i>MATa ADE16<sup>OE</sup>::NAT Nop1-mCherry::NEO</i>	In this study
TBL446	<i>MATa ADE16<sup>OE</sup>::NAT Nop1-mCherry::NEO</i>	In this study
<i>S. cerevisiae</i>		
YS1	NMY32	In this study
YS2	NMY32::AD-T7 BD-53	In this study
YS3	NMY32:: AD-T7 BD-LAM	In this study
YS5	NMY32:: AD-Fbp1 BD-Ade16	In this study
YS9	NMY32:: AD BD-Fbp1	In this study
YS16	NMY32:: AD BD-Ade16	In this study
YS17	NMY32:: AD-Ade16 BD-Fbp1	In this study
Plasmids		
pCN19	Amp <sup>r</sup> Plasmid harboring <i>GFP</i> under histone H3 promoter	Price et al., 2008
pTBL5	Amp <sup>r</sup> Plasmid harboring mCherry-GPD1 terminator and <i>NAT</i> marker	In this study
pTBL59	amp <sup>r</sup> Vector for <i>P<sub>ACTIN</sub>-Nop1-mCherry-NEO</i> for nuclear positioning	In this study
pTBL100	Kana <sup>r</sup> Vector for pGBKT7-FBP1 for yeast two-hybrid	In this study
pTBL106	Amp <sup>r</sup> Vector for pGADT7-ADE16 for yeast two-hybrid	In this study
pTBL142	Amp <sup>r</sup> Vector for pGADT7-FBP1 for yeast two-hybrid	In this study
pTBL145	Kana <sup>r</sup> Vector for pGBKT7-ADE16 for yeast two-hybrid	In this study
pTBL149	Amp <sup>r</sup> Vector for <i>P<sub>CTR4</sub>-ADE16-HA-NAT</i> for Ade16 stability assay	In this study
pTBL153	Amp <sup>r</sup> Vector for <i>P<sub>ACTIN</sub>-Grp1-HA-NAT</i> for <i>GRP1</i> overexpression	In this study
pTBL174	Amp <sup>r</sup> Vector for <i>P<sub>ACTIN</sub>-Ade16-HA-NAT</i> for Ade16 overexpression	In this study
pTBL186	Amp <sup>r</sup> Vector for <i>P<sub>H3</sub>-GFP-Ade16-NAT</i> for Ade16 localization	In this study
pTBL196	Amp <sup>r</sup> Vector for <i>P<sub>ADE16</sub>-mCherry-NAT</i> for temporal expression assay	In this study
pTBL237	Amp <sup>r</sup> Vector for <i>ADE16</i> RNAi	In this study

Table S2. PCR primers used in this study

Primers	Description	Sequence (5' -3')
TL17	M13 F	GTAAAACGACGGCCAG
TL18	M13 R	CAGGAAACAGCTATGAC
TL19	<i>NEO</i> split F	GGGCGCCCGTTCTTTTGTCA
TL20	<i>NEO</i> split R	TTGGTGGTGAATGGGCAGGTAGC
TL59	<i>NEO</i> R4	TGTGGATGCTGGCGGAGGATA
TL67	<i>STE20A</i> α F	CCAAAAGCTGATGCTGTGGA
TL68	<i>STE20A</i> α R	AGGACATCTATAGCAGAT
TL69	<i>STE20A</i> a F	TCCACTGGCAACCCTGCGAG
TL70	<i>STE20A</i> a R	ATCAGAGACAGAGGAGCAAGAC
TL367	<i>ADE16</i> KO F1	ACATGACGAGCACAGCCAGTAGCA
TL368	<i>ADE16</i> KO R1	CTGGCCGTCGTTTACGGTCGCCCCGCTCTCCAC
TL369	<i>ADE16</i> KO F2	TGCGAAGTGCTGGATAGATGAAGATG
TL370	<i>ADE16</i> KO R2	AGACCCTTCACCCCGCTGTTACG
TL371	<i>ADE16</i> KO F3	AGACCCTTCACCCCGCTGTTACG

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TL372	<i>ADE16</i> KO R3	TCCACTGGGGGCTGCGAGGTAGGT
TL373	<i>ADE16</i> KO F4	ATCTCGCTGCACTCTTCTCAAATA
TL572	Fbp1-BD F	CCGAATTCCCGGGGATCCACATGCCCCGTACGACCGTCAAGAAGC
TL573	Fbp1-BD R	GTTATGCGGCCGCTGCAGTCAACGTCCGTTACCGAATCGTTG
TL588	Ade16-AD F	GGCATCGATACGGGATCCACATGTCTTCCGAGGCTCCTATCGGT
TL589	Ade16-AD R	ATTCATCTGCAGCTCGAGTTAATGGTGGAAGTGGAGATGATT
TL879	Fbp1-AD F	GGCATCGATACGGGATCCACATGCCCCGTACGACCGTCAAGAAGC
TL880	Fbp1-AD R	ATTCATCTGCAGCTCGAGTCAACGTCCGTTACCGAATCGTTG
TL883	Ade16-BD F	CCGAATTCCCGGGGATCCACATGTCTTCCGAGGCTCCTATCGGT
TL884	Ade16-BD R	GTTATGCGGCCGCTGCAGTTAATGGTGGAAGTGGAGATGATT
TL893	<i>P<sub>CTR4</sub></i> -Ade16-HA F1	TCCTGCAGCCCCGGGGGATCCATGTCTTCCGAGGCTCCTATCGGT
TL894	<i>P<sub>CTR4</sub></i> -Ade16-HA R1	ACGTCGTATGGGTAGGATCCATGGTGGAAGTGGAGATGATTG
TL1036	<i>ADE16 P<sub>CTR4</sub></i> REP F1	TATCGGGCAGCAGAGGGCAGAG
TL1037	<i>ADE16 P<sub>CTR4</sub></i> REP R1	CTGGCCGTCGTTTTACGTGAGTGAACCACCACAGTCAGAT
TL1034	<i>ADE16 P<sub>CTR4</sub></i> REP F2	GTCATAGCTGTTTCTTGCCAAATCAAGCTTATCGATGGTC
TL1038	<i>ADE16 P<sub>CTR4</sub></i> REP R2	TCGTCGAGGGGAAGACCGACGG
TL1092	<i>P<sub>ACTIN</sub></i> -Ade16-HA F1	CGCCCAACATGTCTGGATCCATGTCTTCCGAGGCTCCTATCGGT
TL1093	<i>P<sub>ACTIN</sub></i> -Ade16-HA R1	ACGTCGTATGGGTAGGATCCATGGTGGAAGTGGAGATGATTG
TL1164	GFP-Ade16 F	GACGAGCTGTACGGATCCATGTCTTCCGAGGCTCCTATCGGT
TL1165	GFP-Ade16 R	CTGGCGGCCGTTACTAGTTTAATGGTGGAAGTGGAGATGATT
TL1166	<i>P<sub>ADE16</sub></i> -mCherry F	ACGGTATCGATAAGCTTCTGTTCTGTGATGGCGAGAGAACAT
TL1167	<i>P<sub>ADE16</sub></i> -mCherry R	CTAGAACTAGTGGATCCGTTGCTTGATGTGGATGAGGTTG
TL1248	<i>ADE16</i> qRT-PCR F	CAAGAAAGGCACCAAGCG
TL1249	<i>ADE16</i> qRT-PCR R	GGGGAAGGGGAAAAAGG
TL1334	<i>ADE16</i> Intro F	AACTCGCCCAACATGTCTGGATCCGTGAGTTGTTTCTCCCGC
TL1335	<i>ADE16</i> Intro R	CTACTGCTACTGTAACCCTTAATTAACTAGTCTGTTTGATGTCAAGAGG
TL1336	<i>ADE16</i> (RNAi) 5'-3' F	AACTCGCCCAACATGTCTGGATCCGGATGTTTCCAACATCACCA
TL1337	<i>ADE16</i> (RNAi) 5'-3' R	GCGGGAGAAACAACCTACGGATCCGAAGTAATCACTGATAGCCTCG
TL1340	<i>ADE16</i> (RNAi) R	GGATAACGGAGAAAGAGGAAATCA
TL1467	<i>ADE16</i> RNAi reverse F1	CCTTAATTAAGGATGTTTCCAACATCACCA
TL1468	<i>ADE16</i> RNAi reverse R1	GGACTAGTTTAGAAGTAATCACTGATAGCCTCG

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