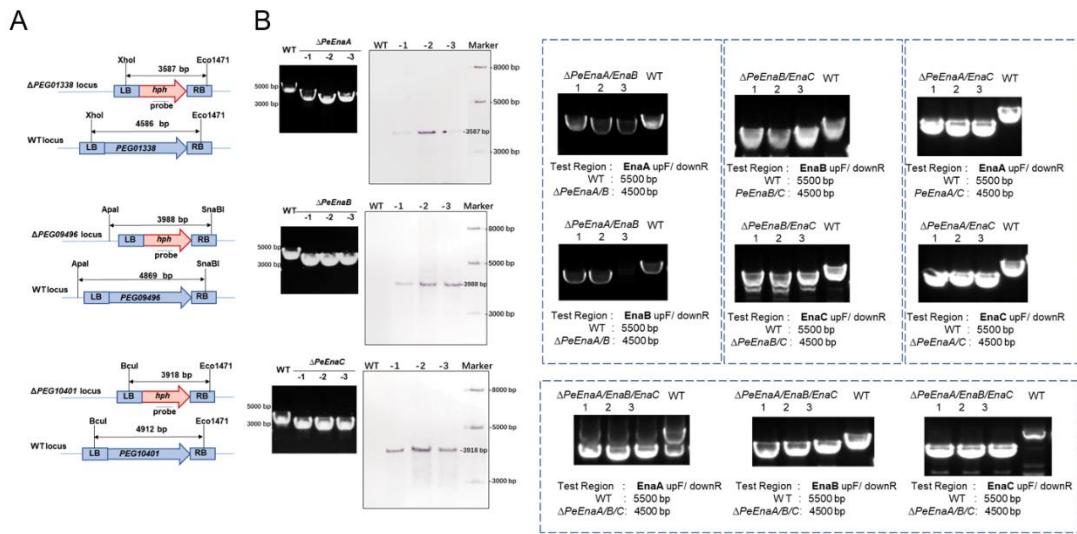
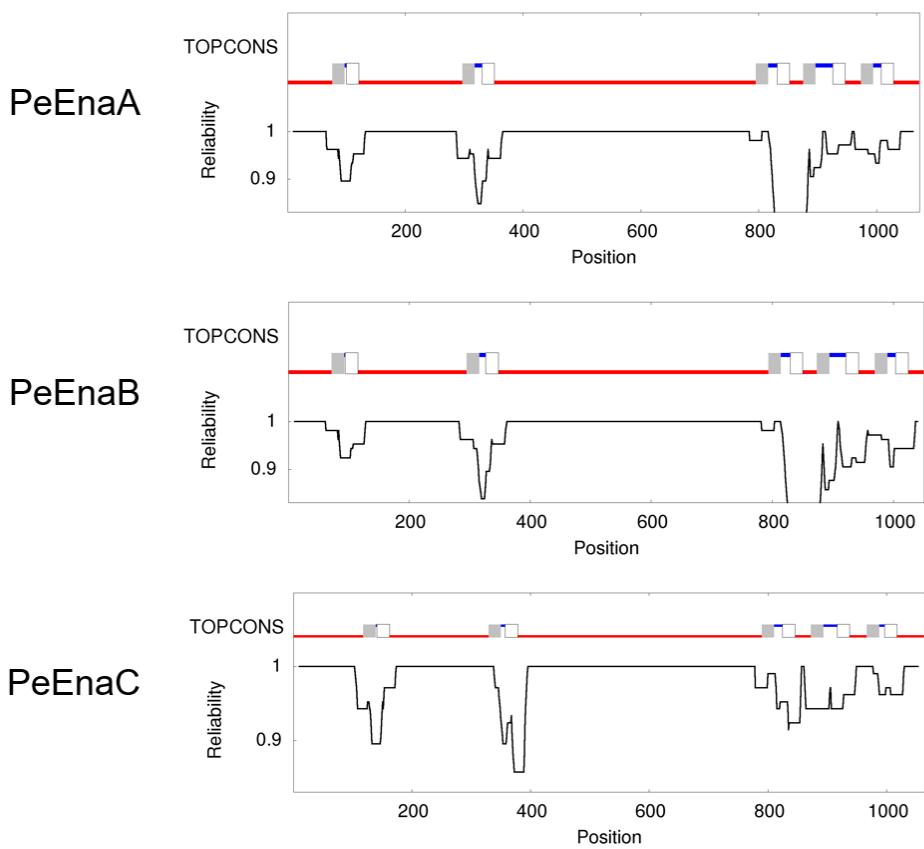


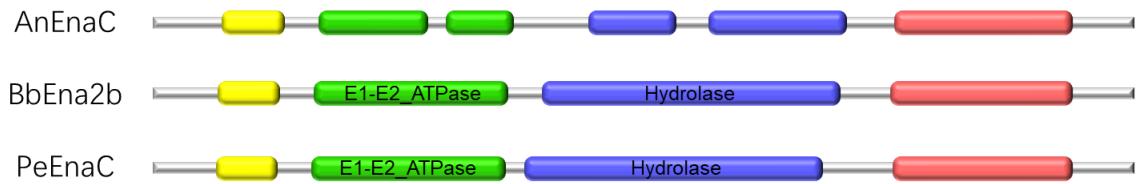
## Supplementary materials



**Figure S1 Knockout of *PeEnaA*, *PeEnaB*, and *PeEnaC* in *P. expansum*. A.** *PeEnaA*, *PeEnaB*, and *PeEnaC* deletion strategy used by homologous recombination. **B.** PCR detection and Southern Blot of  $\Delta PeEnaA$ ,  $\Delta PeEnaB$ , and  $\Delta PeEnaC$  knockout mutants. WT means wild-type strain.

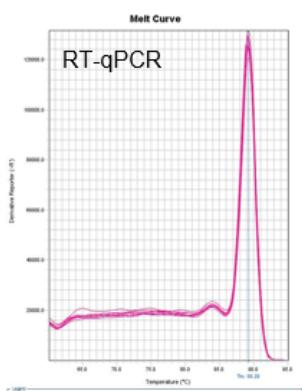


**Figure S2 The transmembrane topology of the PeEna proteins.** The transmembrane topology was determined by manual assertion deduced from a combination of prediction using the transmembrane topology web server TOPCONS.

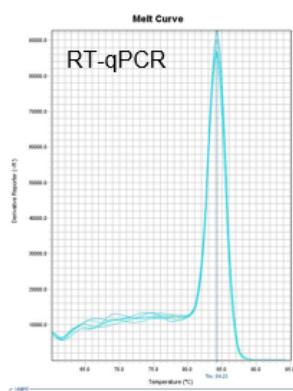


**Figure S3 Functional conserved domain analysis of Ena family proteins via Pfam database.** The cation-ATPase-N (pfam00690) domain is marked in red, the E1-E2\_ATPase (pfam00122) domain is marked in green, the halo acid dehalogenase-like hydrolase (HAD) (COG4087. pfam00702) domain is marked in blue, and Cation\_ATPase\_C (pfam00689) domain is marked in yellow.

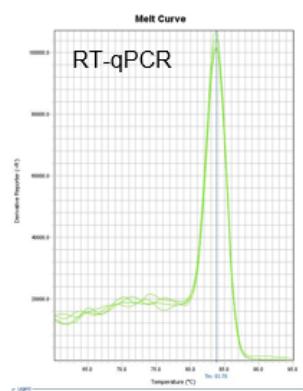
*PeEnaA*



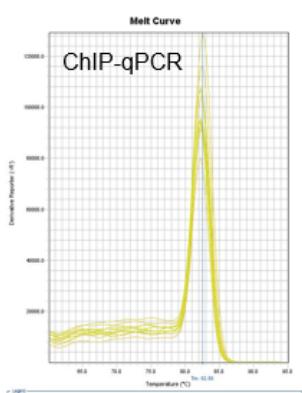
*PeEnaB*



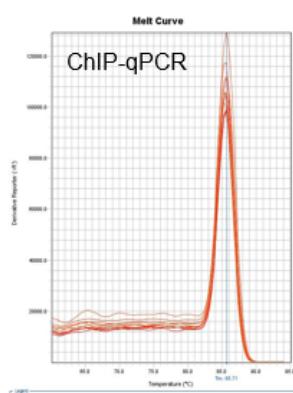
*PeEnaC*



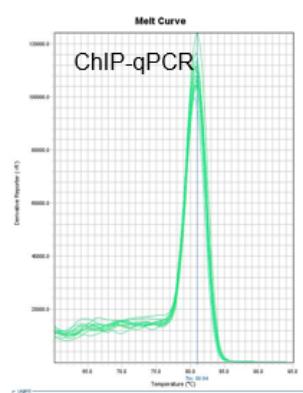
*PeEnaA-1*



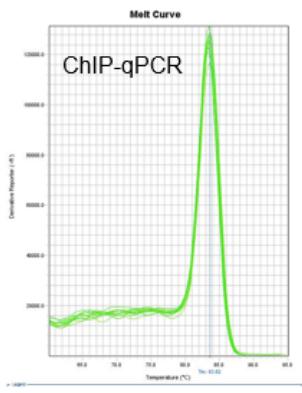
*PeEnaA-2*



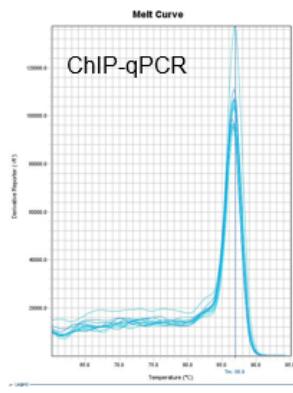
*PeEnaB-1*



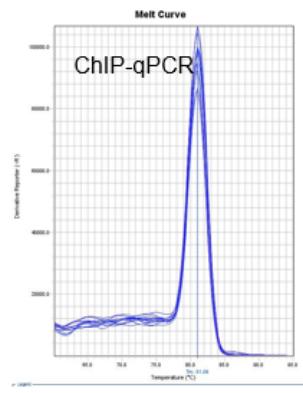
*PeEnaB-2*

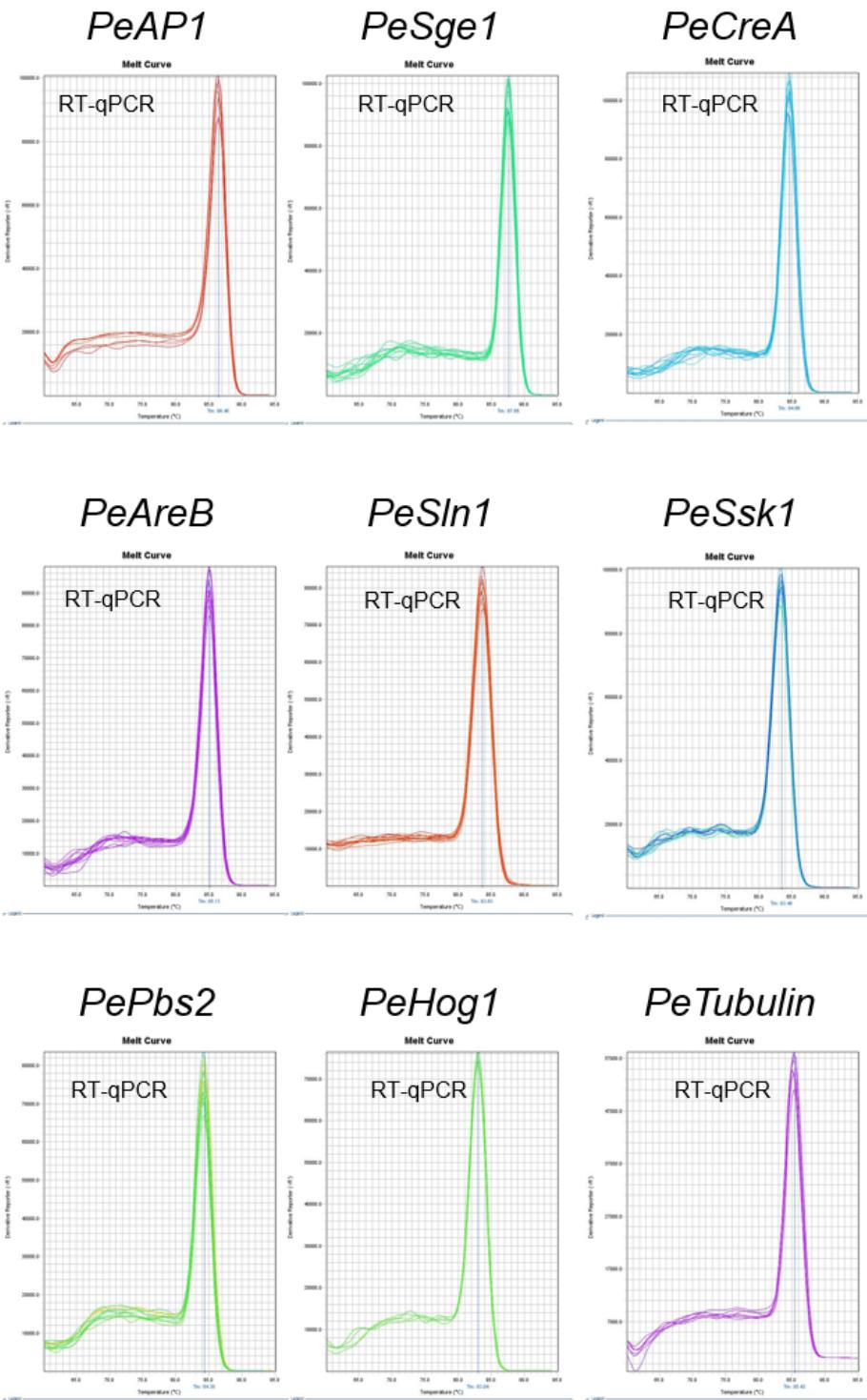


*PeEnaC-1*



*PeEnaC-2*





**Figure S4** The melting curves of representative genes in this study.

**Table S1** The primers used for construction and identification of gene deletion, complementation and eGFP tag strains in this study.

Primer name	Primer sequence (5'-3')
EnaA-up-F	GAATTGAGCTCGGTACCCGGGTGCCGCATGGCTGTGC GTCGTCG
EnaA-up-R	CCACAGCTGCAGGTCGACTCTAGATTGCGATTTCGCAA ATGGTAAGA
EnaA-down-F	AGTAACCATGCATGGTTGCCTAACTCAATAGATTACTC AAATGAACAAC
EnaA-down-R	GCCAGTGCCAAGCTCGGCGGCCAAATGCCGGAAGA GTTTGCAGAA
EnaC-up-F	GAATTGAGCTCGGTACCCGGGTTGTGTTGAGCAAAA AAGTAGC
EnaC-up-R	CCACAGCTGCAGGTCGACTCTAGATCAATAAGATTGTAG GAAAGAAAG
EnaC-down-F	AGTAACCATGCATGGTTGCCTAACTCTGGAAAATTGTC TGACTGAT
EnaC-down-R	GCCAGTGCCAAGCTCGGCGCGCCTGATCATTCAACGTT GAATAAACG
EnaB-up-F	GAATTGAGCTCGGTACCCGGGAGGAACAGCGTCTATGT TGATGCG
EnaB-up-R	CCACAGCTGCAGGTCGACTCTAGATGGATATTGTCGTTC GATGGGTAG
EnaB-down-F	AGTAACCATGCATGGTTGCCTAACTCCGAATGCTTCCCT GGCCCAACAT
EnaB-down-R	GCCAGTGCCAAGCTCGGCGCGCCTATGGCGATGTAATT CAAATTCGC
EnaA-test-F	GTATGCATGAAGCGCGGGGGTTG
EnaC-test-F	ACAGGAAGATGCAGTGGCTTCGC
EnaB-test-F	TGGCCCAGCTGAGATGAGCCATT
HPH-F	TCACCCCCATCTCAACTCCA
HPH-R	TGCTCCATACAAGCCAACAC
NEO-F	ATGATTGAACAAGATGGATTGC
NEO-R	TCAGAAGAACTCGTCAAGAAG
Nat-F	TCCTTCACCACCGACACCGTCTTCC
Nat-R	CGTCCGATTGTCGTCGATTGTC
EnaA-Com-F	AACTCCATCACATCACAAGAGCTCGCAGGAAGCCAA AAAGGCTCT
EnaA-Com-R	CTCCTGCCCTTGCTCACTCTAGATTGAAGATTACTTGT TCATTA
EnaB-Com-F	AACTCCATCACATCACAAGAGCTCATGTCAGACGAGAAG CCTGGCA

EnaB-Com-R	CTCCTGCCCTTGCTCACTCTAGATTAGGGTTGCTCCGGG ATCTC
EnaC-Com-F	AACTCCATCACATCACAAAGAGCTCATAGAGGTAACCTCCG ATGTCGA
EnaC-Com-R	CTCCTGCCCTTGCTCACTCTAGACTTCAGTTCCACGCTT GCATTCGC

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**Table S2 Formulation of the citrate-phosphate buffer**

<b>pH</b>	<b>0.2 mol/L Na<sub>2</sub>HPO<sub>4</sub> (mL)</b>	<b>0.1 mol/L Citric acid (mL)</b>
3	4.11	15.89
5	10.30	9.70
8	19.45	0.55

**Note:**

Preparation of MM media at pH 5 and 8: Prepare 2× MM medium and the above citrate-phosphate buffer, and preheat them at 80°C, respectively. Mix the medium and the buffer in a ratio of 1:1 and spread the plates.

Preparation of CY media at pH 3, 5, and 8: Prepare 2× CY liquid medium and the above citrate-phosphate buffer, Mix them in a ratio of 1:1.

**Table S3** The primers used for RT-qPCR in this study.

Name	Sequence (5'-3')	Annealing temperatures (°C)	Accession number
<i>EnaA</i> -RegionA	F: CTGGAGAAGTAGGTGGCTTGAA R: CTCGCCGGTCCCCACGCGACAAT	60	PEX2_013210 (PEG01338)
<i>EnaA</i> -RegionB	F: ATGTGCCAGAGTCTTTGCGTCA R: TCGATGGCGAAAGATAAGCGGGCT	60	PEX2_013210 (PEG01338)
<i>EnaB</i> -RegionA	F: TGCTCCTTCCCAGTGTGTTGTG R: GCTGTGAACCTGCCTCGTCTTGC	60	PEX2_039950 (PEG09496)
<i>EnaB</i> -RegionB	F: TGGAGCAAGACTGTTGAAGTGGC R: GACAGACATCGGAATCTGGTGGG	60	PEX2_039950 (PEG09496)
<i>EnaC</i> -RegionA	F: CCACGACTGCCTAGAACGCAAGT R: TCCGACGGGAGTGTCAAAGATA	60	PEX2_085740 (PEG10401)
<i>EnaC</i> -RegionB	F: CCCTCTAGGGTTGGAAGCAAGT R: GTAGCAAGGCGGACGAAGGTGTA	60	PEX2_085740 (PEG10401)
<i>PeLaeA</i>	F: TCGAAGCTCTCCGGACATG R: AATGCAATGCGGTCAATCTG	60	PEX2_035880 (PEG08484)
<i>PeVeA</i>	F: GAAGATACTTCGGCCTTGATGA R: TCGCGATAAGCAGGATAAGGA	60	PEX2_043190 (PEG11106)
<i>PeVelB</i>	F: TTGTTGTCTGGGTCCGTCAA R: CCTGGGATGAGGGATTCTG	60	PEX2_031850 (PEG02453)
<i>PeAP1</i>	F: GAGCCTGTTGCCAGATTGATA R: GTTCACGACTACACCACCTCA	60	PEX2_065210 (PEG01534)
<i>PeSge1</i>	F: ATGGAATGTACTTCGGGCTGCTT R: TAGTGGGTCGTTGGATGGGATAA	60	PEX2_061550 (PEG11642)
<i>PeCreA</i>	F: TCACGATACTTCGCCCCACTCAA R: GAAGGCACGGTCGCAAAGAGGAC	60	PEX2_022240 (PEG01401)
<i>PeAreB</i>	F: CTATACCCTCATATGGCGACCAC R: TTCAAGAAGAGTCCACATGCGTTG	60	PEX2_003790 (PEG07853)
<i>PeSln1</i>	F: CCCAGACCGACTCAAGTATCAAC R: GCTATCTAGCAGAAATGTCACCTCCC	60	PEX2_054790 (PEG04458)
<i>PeSsk1</i>	F: CGATCAATACAATTCTCCTTGCC R: GCTGGGATTGGAGTTGAGCTTTC	60	PEX2_072810 (PEG02228)
<i>PePbs2</i>	F: GGAACTGGGTTATCTGCCTCGTC R: TGCCCTGTTGAAGCAGCAGTATG	60	PEX2_005030 (PEG09483)
<i>PeHog1</i>	F: CACCGCTCTCAAATCACCGACAG R: TGCCATCAAACGGCTCCCTATC	60	PEX2_060120 (PEG11493)
<i>PePatA</i>	F: AAAGGCCGGTGCATTGATC R: TTGGAGGGCTTGGTGAGCAT	60	PEX2_082810 (PEG03683)
<i>PePatB</i>	F: GCCAGGCTATGCGATTGAGT R: GCTGGAACCCTGTCCATTGT	60	PEX2_082800 (PEG03684)

<i>PePatC</i>	F: TCCACCTGCGAATATCCCTTA R: CATGCCAGTGCCATTTC	60	PEX2_082790 (PEG03685)
<i>PePatD</i>	F: ATGAGATTGCGTCTGCGCAAAG R: CTACCCAAGCGGGATGAGATT	60	PEX2_082780 (PEG03686)
<i>PePatE</i>	F: CATTCTCATGGGCCTGAGT R: TCGAAGCTCTTCCGGACATG	60	PEX2_082770 (PEG03687)
<i>PePatF</i>	F: GCGAGTGAATTGGCCAAT R: GTCCGACCAAAGGATGAAG	60	PEX2_082760 (PEG03688)
<i>PePatG</i>	F: CGGCCGTCTTGAAGGAAAT R: CTTGCCGTAGCGGGTGAATA	60	PEX2_082750 (PEG03689)
<i>PePatH</i>	F: CATTATCGCGGGTGTCTGA R: GATCAACGCTTGCACGATAGC	60	PEX2_082740 (PEG03690)
<i>PePatI</i>	F: GCAAACCTATTCCGCAAGGA R: TGGTTCTGCCATCGATCAC	60	PEX2_082860 (PEG03676)
<i>PePatJ</i>	F: CGCCAGACATACCGCCATA R: TTTGGTCGATCGGGACTGTT	60	PEX2_082870 (PEG03675)
<i>PePatK</i>	F: GACGCTGGCTACTGGATTG R: TCGTGCCTGAGGCCAGTAT	60	PEX2_082880 (PEG03674)
<i>PePatL</i>	F: GCAGGAGATCCGTTTCAGACA R: CCACTGACCGACGGTTACAAC	60	PEX2_082850 (PEG03677)
<i>PePatM</i>	F: ACCCACAGCTGCACATGGA R: AGCGAGAAGAGGCCAGAAGA	60	PEX2_082820 (PEG03681)
<i>PePatN</i>	F: CGTCGATGTCGCTAGCAA R: GGCGATAATCACGTCAATTG	60	PEX2_082830 (PEG03680)
<i>PePatO</i>	F: TCGCCTCCTGGTGTATCTT R: AAGCGTGCCAGTCATTGAG	60	PEX2_082840 (PEG03678)
$\beta$ -tubulin	F: CTCCAGCTCGAGCGTATGAAC R: GGCTCAAATCGACGAGAAC	60	PEX2_027410 (PEG11155)