



Communication

Cecal Reduction of *Brachyspira* and Lesion Severity in Laying Hens Supplemented with Fermented Defatted 'Alperujo'

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Abstract: Antimicrobial resistance demands the development of therapeutic alternatives such as prebiotics, probiotics, and nutraceuticals. The aim of this study was to assess the antimicrobial properties of the nutraceutical fermented defatted “alperujo”, derived from olive oil production, in a laying hen farm ($n = 122,250$) endemic with avian intestinal spirochetosis (*Brachyspira* spp.). Part of the batch ($n = 1440$) was divided into six groups of 240 hens each that included 80 or 108-week-old laying hens, supplemented with 0%, 2%, or 6% fermented defatted ‘alperujo’ for a month. At the end of the experiment, eight hens from each group were autopsied and cecal content was subjected to (i) *Brachyspira* culture and species identification by PCRs, and (ii) direct DNA extraction and *Brachyspira* qPCR. Furthermore, the ceca were processed for histopathology. Microbiological isolation revealed *B. pilosicoli* and *B. hyodysenteriae* co-infection in all groups. The 80-week-old hen group 2% supplemented showed a reduction in the cecal *Brachyspira* content (qPCR) compared with non-supplemented hens. Cecal histopathology showed a diffuse mild infiltration of lymphocytes, plasma cells, and heterophils; and hyperplasia of the gut-associated lymphoid tissue hyperplasia which decreased in severity in 80-week-old supplemented hens. The reduction in *Brachyspira* colonization and the severity of the lesions observed in supplemented hens highlights a potential protective function against avian intestinal spirochetosis.

Keywords: *Brachyspira*; intestinal health; laying hen; nutraceutical; poultry



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1. Introduction

Avian intestinal spirochetosis is a worldwide neglected disease of domestic avian species and waterfowl caused by *Brachyspira* spp. [1]. Some species are regarded as pathogens of poultry: *B. intermedia*, *B. pilosicoli*, and *B. alvinipulli* [2–4]. Furthermore, *B. pilosicoli* has zoonotic potential leading to food-borne infection in immunocompromised people [5,6]. *Brachyspira* infection of laying hens causes chronic diarrhea, discomfort, and mortality [4,7]. Avian intestinal spirochetosis productive performance impairment and economic importance is due to fecal staining of eggshells, delayed onset of lay in pullets, reduced egg production, poor chick quality from infected breeders, and reduction of feed conversion ratio [4,7,8]. The cost of avian intestinal spirochetosis was estimated at 14 million pounds (over 16 million euro) in the United Kingdom in 2006 [7]. Antimicrobial treatment (ampicillin, tiamulin, lincomycin, zinc bacitracin) is the preferred tool to control *Brachyspira* infections in laying hen farms [2,7–9]. However, relapse after treatment and emerging antimicrobial resistance have been reported for avian spirochetes, demanding re-orientation of avian intestinal spirochetosis management [3,8,10].

Probiotic, prebiotic, and nutraceutical products are compounds with antimicrobial properties as feed additives or supplements in poultry [11–13]. Specifically, nutraceuticals are

compounds derived from natural sources with benefits other than nutrition [14]. The olive oil two-phase extraction system produces the semisolid by-product ‘alperujo’ [15] that can be adapted to animal feed through bacterial fermentation, fat hydrolysis, desiccation, and grinding [16]. The nutraceutical obtained, fermented defatted ‘alperujo’, has been tested in laying hens and broilers showing an improvement of the intestinal health by means of higher bacterial diversity and enhancement of mucosal histological parameters [16,17]. Improvement of some productive parameters has been also reported in laying hens (reduced broken eggs) [16] and broilers (body weight) [17]. In addition, the co-administration of biscuit flour and fermented defatted ‘alperujo’ has also proved beneficial for intestinal health and production performance in laying hens [18].

Olives contain phenolic compounds with beneficial properties (antioxidant, anti-inflammatory, antithrombotic, antimicrobial, and anti-coccidian) that are retained in olive oil by-products [19–21]. The antimicrobial properties of fermented defatted ‘alperujo’ have been previously investigated in broilers experimentally challenged with *Salmonella* Typhimurium [22]. The results of these experiments showed that dietary supplementation delayed and reduced *Salmonella* Typhimurium colonization in the cecum of challenged broilers, contributing to prophylactic and therapeutic measures to reduce salmonellosis prevalence in poultry farms [22]. However, the effectiveness of fermented defatted ‘alperujo’ to reduce colonization by other enteric bacterial pathogens aside from *Salmonella* is unexplored. Therefore, in this study, the potential benefits of fermented defatted ‘alperujo’ were assessed to control avian intestinal spirochetosis in a laying hen farm.

2. Materials and Methods

2.1. Laying Hens and Rearing Conditions

This study was performed in a commercial laying hen farm (Hy-Line 2015, $n = 122,250$), from which part ($n = 1440$) was divided into six groups of 240 hens each (720 80-week-old laying hens and 720 108-week-old laying hens, see below) and monitored for a month. Hens were kept in an intensive housing system under homogeneous environmental conditions (24–32 °C, 50–70% humidity). Feed and water were supplied ad libitum.

2.2. Diet, Groups, and Sampling

For the dietary inclusion, fermented defatted ‘alperujo’ was prepared as previously detailed [16]. ‘Alperujo’ first underwent a controlled anaerobic bacterial fermentation, was then defatted with chemical solvents (fat hydrolysis), followed by desiccation at 80 °C in a low oxygen content atmosphere, and finally ground. Fermented defatted ‘alperujo’ composition is detailed in Table 1.

Table 1. Fermented defatted ‘alperujo’ composition [16].

Determination	Results
Moisture 103° (%w.w.)	12.2
Crude protein (Kjeldahl) (%w.w.)	6.4
Brute fat (%w.w.)	3.0
Ash content (%w.w.)	7.7
Lignin (%w.w.)	23.3
Acid detergent fiber (%w.w.)	39.2
Neutral detergent fiber (%w.w.)	49.3
Tannins (%w.w.)	0.06
Oleic acidity index (%w.w.)	46.1
Peroxide value (%w.w.)	7.9
Total polyphenols (mEq/kg)	0.89
Crude fiber (%w.w.)	27.7

Fermented defatted ‘alperujo’ was added to the commercial feed (Table 2) in a 2% or 6% respect to the feed weight by employing a mixing machine

Table 2. Ingredient composition of the commercial feed.

Ingredients	g/kg)
Yellow corn (grain)	442.00
Barley E 11, 4	225.00
Soybean meal 47	200.00
Sunflower meal 28	120.00
Calcium carbonate	14.10
Tricalcium phosphate	7.90
Soybean oil	9.00
Premix	4.00
Common salt, NaCl	1.97
Sodium bicarbonate	1.98
L-lysine 95%	1.03
Methionine	0.75

All laying hens were fed the same commercial formulation as used in the farm, but some hens were supplemented with 2% or 6% fermented defatted ‘alperujo’ depending on the group: (1) 80-week-old hens infected with *Brachyspira* spp., (2) 80-week-old hens infected with *Brachyspira* spp. and dietary supplemented with 2% fermented defatted ‘alperujo’, (3) 80-week-old hens infected with *Brachyspira* spp. and dietary supplemented with 6% fermented defatted ‘alperujo’, (4) 108-week-old hens infected with *Brachyspira* spp., (5) 108-week-old hens infected with *Brachyspira* spp. and dietary supplemented with 2% fermented defatted ‘alperujo’, and (6) 108-week-old hens infected with *Brachyspira* spp. and dietary supplemented with 6% fermented defatted ‘alperujo’.

One month after dietary supplementation, eight hens from each group ($n = 48$ in total) were euthanized. During autopsies, cecal content was sampled in parallel and subjected to (i) *Brachyspira* culture, DNA extraction, and identification by polymerase chain reaction (PCR), and (ii) direct DNA extraction from cecal content and *Brachyspira* quantification by real time PCR (qPCR) [23]. Additionally, ceca samples were collected for histopathological studies.

2.3. *Brachyspira* Culture and Confirmation by PCR

For *Brachyspira* culture and confirmation, cecal samples were cultured onto MacConkey agar, MacConkey agar supplemented with cefotaxime (CTX, 1 mg/L), and trypto-casein soy agar (TSA) plates. Plates were incubated at 37 °C in anaerobic conditions (80% N₂, 10% H₂, and 10% CO₂). Plates were inspected after 72 h and, if negative, daily for 10 days. Morphologically compatible cultures were sub-cultured until pure, and DNA was extracted for further identification by polymerase chain reaction (PCR). *Brachyspira* isolates were identified using previously described PCR protocols for *B. intermedia*, *B. pilosicoli*, and *B. hyodysenteriae*. Briefly, a PCR was performed based on the amplification of the NADH oxidase (nox) gene of *B. intermedia* (567 base pair region) and *B. hyodysenteriae* (1268 base pair region) [24], and the 16S rRNA gene (823 base pair region) of *B. pilosicoli* [25].

2.4. Quantitative Real-Time PCR for *Brachyspira*

Direct DNA extraction from cecal samples was carried out using a commercial kit (FASTI001-1 FavorPrep Stool DNA Isolation Mini Kit, Favorgen-Europe, Vienna, Austria), following the manufacturer’s specifications (elution volume of 200 µL), coupled with a specific qPCR assay for quantitative detection of *Brachyspira* spp., as described previously [23]. Briefly, a multiplex qPCR assay was performed based on the amplification of a 198 base pair portion of the NADH oxidase gene, using TaqMan probes for detecting and quantifying *B. hyodysenteriae*, *B. pilosicoli*, and *B. intermedia*. The detection limit for all three targeted species was 1–10 viable cells and 10 fg DNA per reaction [23]. Genome equivalent/g was calculated performing a standard curve based on 10-fold serial dilutions of known amounts of positive control DNA.

2.5. Histological Processing

Ceca samples obtained during the autopsies were fixed in 10% buffered formalin, automatically dehydrated in ethanol series and xylene substitute, and embedded in synthetic paraffin (Citadel 2000 Tissue Processor, Thermo Fisher Scientific, Waltham, MA, USA). Paraffin blocks (Histo Star Embedding Workstation, Thermo Fisher Scientific) were cut at 4 μ m sections (Finesse ME+ Microtome, Thermo Fisher Scientific) and stained with hematoxylin-eosin (Gemini AS Automated Slide Stainer, Thermo Fisher Scientific), mounted (CTM6 Coverslipper, Thermo Fisher Scientific), and examined under light microscopy (DM2000 LED, Leica, Wetzlar, Germany).

2.6. Statistical Analysis

The qPCR results (genome equivalent/g) were compared according to the percentage of supplementation using a Kruskal–Wallis test, followed by a Holm–Bonferroni post hoc test. In addition, this comparison was performed for each age group.

3. Results

3.1. *Brachyspira* Culture and Confirmation on Cecal Content

Regardless of the group, *Brachyspira* was cultured from all cecal contents as detailed in Table 3. Species identification revealed *B. pilosicoli* and *B. hyodysenteriae* co-infection in all the groups (Table 3). *B. intermedia* was not detected in any sample.

Table 3. *Brachyspira* detection in cecal content according to the groups.

Hens Age	Dietary Inclusion	<i>Brachyspira</i> Culture	<i>Brachyspira intermedia</i>	<i>Brachyspira pilosicoli</i>	<i>Brachyspira hyodysenteriae</i>
80 weeks	0%	8/8 (100%)	0/8 (0.0%)	6/8 (75.0%)	8/8 (100%)
	2%	8/8 (100%)	0/8 (0.0%)	7/8 (87.5%)	7/8 (87.5%)
	6%	8/8 (100%)	0/8 (0.0%)	8/8 (100%)	8/8 (100%)
108 weeks	0%	8/8 (100%)	0/8 (0.0%)	8/8 (100%)	8/8 (100%)
	2%	8/8 (100%)	0/8 (0.0%)	7/8 (87.5%)	6/8 (75.0%)
	6%	8/8 (100%)	0/8 (0.0%)	8/8 (100%)	8/8 (100%)

3.2. Quantitative Real-Time PCR for *Brachyspira*

In 80-week-old laying hens, dietary supplementation with fermented defatted ‘alpe-rujo’ showed a reduction in the cecal *Brachyspira* content (Figure 1) in the 2% group (median: 1.59×10^6) compared with non-supplemented hens (median: 3.30×10^6). No statistical differences were observed in the 2% and in 6% group (median: 3.65×10^6) compared with controls ($p > 0.05$).

In 108-week-old laying hens, dietary supplementation with fermented defatted ‘alpe-rujo’ did not show any reduction in the cecal *Brachyspira* content (Figure 1) in 2% (median: 6.10×10^6) or 6% (median: 4.15×10^6) groups compared with non-supplemented hens (median: 3.75×10^6). No statistically significant differences were found among groups ($p > 0.05$).

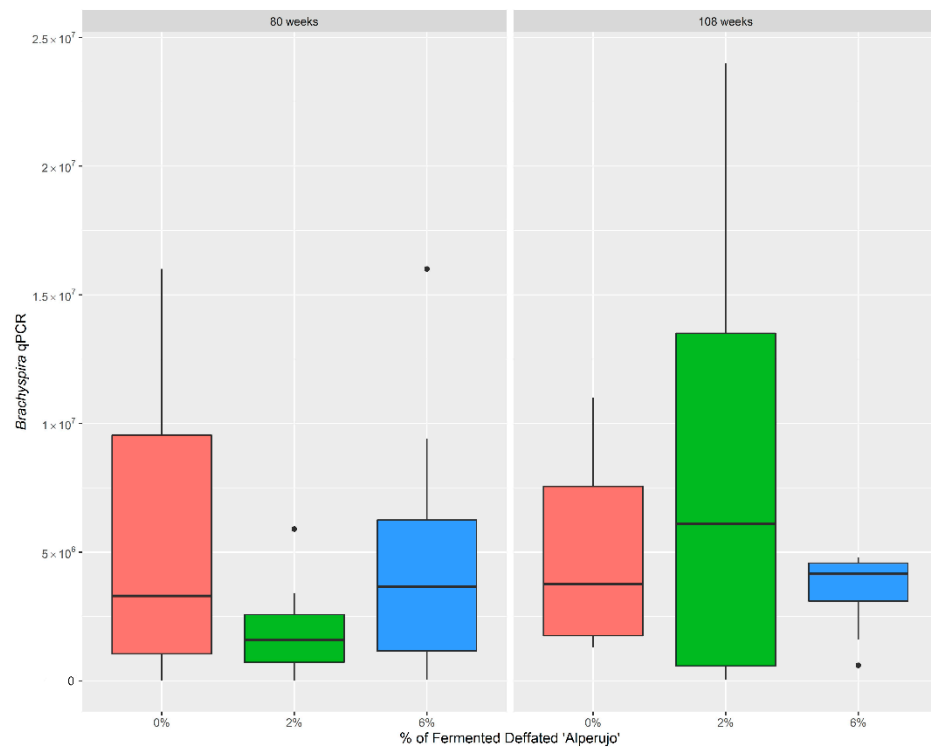


Figure 1. Distribution of the *Brachyspira* spp. qPCR (genome equivalent/g) observed among groups. Boxes indicate the interquartile range, middle highlighted bars inside boxes indicate median values, and top and bottom of the box indicate the 75th and 25th percentile. Whiskers denote 97.5th and 2.5th percentile. Dots represent outliers. Dietary supplementation with fermented defatted ‘alperujo’ showed a reduction in the cecal *Brachyspira* content in 80-week-old laying hens.

3.3. Histopathology

Histopathological study of the cecum in non-supplemented laying hens regardless of the age group showed a moderate chronic diffuse lymphoplasmacytic and heterophilic typhlitis, displaying also mild to moderate epithelial hyperplasia and moderate gut-associated lymphoid tissue (GALT) hyperplasia (Figure 2a).

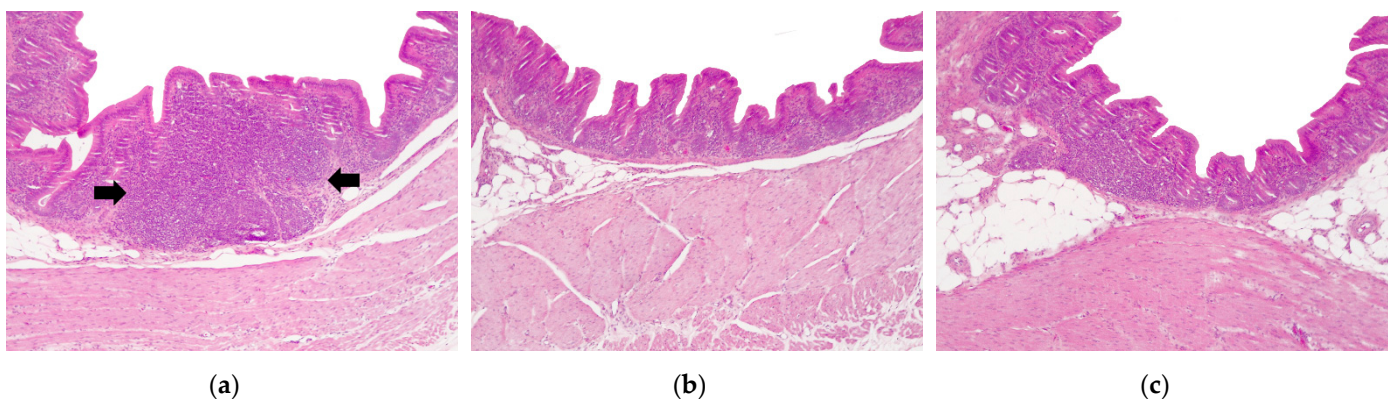


Figure 2. Histopathological study of the cecum of 80 and 108-week-old laying hens infected with *Brachyspira* and supplemented with fermented defatted ‘alperujo’. (a) Non-supplemented groups. Moderate chronic diffuse lymphoplasmacytic and heterophilic typhlitis and marked GALT hyperplasia (arrows). (b) Two percent supplemented groups. There is a marked reduction in the lesion severity compared with non-supplemented hens. (c) Six percent supplemented groups. There is a reduction in the inflammation compared with non-supplemented hens. Hematoxylin-eosin, 100×, scale bar: 200 μm.

Supplemented laying hens showed a marked reduction in the intestinal lesions, displaying a reduction in the severity of the lymphoplasmacytic and heterophilic inflammation in the cecal lamina propria and less/absence of GALT hyperplasia. Histopathological changes were milder in 2% supplemented group (Figure 2b) compared with 6% supplemented group (Figure 2c).

4. Discussion

Avian intestinal spirochetosis is an emerging disease which includes the genus *Brachyspira*, which groups different species recognized as poultry pathogens. Here, before investigating the effectiveness of the nutraceutical tested against *Brachyspira*, we confirmed the natural infection and revealed two major species infecting (co-infecting) the laying hens of the flock: *B. pilosicoli* and *B. hyodysenteriae*. Studies performed in the United States of America (USA) also revealed *B. pilosicoli*, but in a reduced number of flocks (23.8%) and with a low within-flock prevalence (10.7%) compared with other *Brachyspira* spp. [26]. Although *B. intermedia* seem to be more prevalent in the USA, we did not find *B. intermedia* in any cecal sample [26]. On the other hand, *B. hyodysenteriae* has been previously reported in poultry [27]. This suggests that different *Brachyspira* spp. may be involved in avian intestinal spirochetosis, depending on the geographic region. In addition, studies focusing on the diversity of *Brachyspira* spp. in laying hen flocks also found the presence of different *Brachyspira* spp. in fecal samples as reported here [25].

Antimicrobials are the main control strategy for avian intestinal spirochetosis in poultry [2,7–9]. Despite the current antibiotic use, antimicrobial resistance and subsequent use restriction in the European Union demands development of therapeutic alternatives in animal health, such as prebiotics, probiotics, and nutraceuticals. Dietary composition has been studied in the context of *Brachyspira* infection in laying hens. For example, the use of different diets based on wheat, barley, or barley and sorghum showed that laying hens fed wheat had significantly more colonization by *B. intermedia* than the other hens fed with other cereals [28]. Furthermore, the wheat variety also seems to influence *B. intermedia* intestinal colonization in laying hens [29]. Few natural compounds have been tested as antimicrobial alternatives in poultry models of *Brachyspira* infection. Essential oils have been tested in vitro and in vivo in a study [30]. Of them, cinnamaldehyde was proven inhibitory for *B. intermedia* in vitro and was shown to reduce *B. intermedia* in the cecum of pullets in vivo [30]. However, the underlying mechanisms were not clearly defined, with microbiota and mucosal structure changes considered to be the main mechanisms involved in *B. intermedia* reduction [30]. The use of the probiotic *Lactobacillus reuteri* LM1 in drinking water was demonstrated useful to reduce *B. pilosicoli* clinical signs, histological intestinal lesions, and improve performance in laying hens, possibly through bacterial competition in the gut lumen and/or intestinal microenvironment acidification [31].

Here, the nutraceutical fermented defatted ‘alperujo’, derived from the olive oil production, was assessed to control *Brachyspira* infection in a laying hen farm endemic with avian intestinal spirochetosis. Our results showed a reduction in *Brachyspira* spp. cecal content in 80-week-old laying hens supplemented at 2%. Similar to our results, the employment of this nutraceutical at 2% has proven valuable to control cecal *Salmonella* Typhimurium colonization in broilers under experimental conditions [22]. The bacteriostatic capacity of this nutraceutical has been attributed to the inhibition of bacterial division via reduction of the ATP intracellular concentration through the synergistic action of phenols and polyphenols, particularly hydroxytyrosol [22].

The effects on the intestinal health of the nutraceutical tested here also comprise microbiota modulation and morphologic changes in the intestine [16–18,22]. The evaluation of the intestinal mucosa provides valuable information on health and disease in laying hens. Herein, the histopathological results in *Brachyspira*-infected non-supplemented laying hens revealed evident cecal inflammation. While some authors did not report histological lesions in *Brachyspira*-infected laying hens [9], others describe the attachment of *Brachyspira* to the intestinal epithelium before invading the mucosa and inducing a variable

lymphoplasmacytic typhlocolitis [8,27,31]. Here, the supplementation with fermented defatted ‘alperujo’ markedly reduced intestinal inflammation regardless of the age range, especially at 2% supplementation. These results coincide with those obtained in broilers infected with *Salmonella* Typhimurium and supplemented with fermented defatted ‘alperujo’, which display a reduction in the intestinal inflammation compared with non-supplemented *Salmonella*-infected controls [22]. Our results also coincide with those obtained in laying hens 21 days after *B. pilosicoli* challenge and supplemented with the probiotic *Lactobacillus reuteri* LM1, which showed a marked reduction in the severity of histopathological lesions [31]. This highlights the need to monitor histopathological changes during efficacy trials testing antimicrobial compounds.

The age of the animal is known to influence epidemiology and pathogenesis in avian intestinal spirochetosis, as initial infection of pullets leads to amplification and transmission within the farm [5,25,32]. Therefore, two age groups (80- and 108-week-old laying hens) were independently studied to assess age differences. The results showed no positive effects in 108-week-old hens. This may be explained by the increased rate of colonization of *Brachyspira* in older animals [32]. Concordantly, some authors have set that intestinal spirochetes significantly increase in chickens older than 40 weeks of age [5]. The expected increased *Brachyspira* content in the ceca of older hens may explain the decreased effectiveness of the tested nutraceutical in this age range.

5. Conclusions

This study showed that dietary supplementation with fermented defatted ‘alperujo’ at 2%, but not 6%, promotes the cecal reduction of *Brachyspira* (*B. pilosicoli* and *B. hyodysenteriae*) and diminished cecal inflammation in 80-week-old laying hens. This nutraceutical can contribute to the control of avian intestinal spirochetosis, thereby reducing the use of antimicrobials in poultry.

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