



Review

Diagnostic Management of Gastroenteropancreatic Neuroendocrine Neoplasms: Technique Optimization and Tips and Tricks for Radiologists

Fabio Pellegrino ^{1,*}, Vincenza Granata ², Roberta Fusco ³, Francesca Grassi ^{4,5}, Salvatore Tafuto ⁶, Luca Perrucci ⁷, Giulia Tralli ⁸ and Mariano Scaglione ⁹

¹ Radiology Division, S. Bonifacio Hospital, 37047 Verona, Italy

² Division of Radiology, Istituto Nazionale Tumori IRCCS Fondazione Pascale—IRCCS di Napoli, 80131 Naples, Italy

³ Medical Oncology Division, Igea SpA, 80013 Naples, Italy

⁴ Italian Society of Medical and Interventional Radiology (SIRM), SIRM Foundation, Via della Signora 2, 20122 Milan, Italy

⁵ Division of Radiology, Università degli Studi della Campania Luigi Vanvitelli, 80127 Naples, Italy

⁶ S.C. Sarcomi e Tumori Rari, Istituto Nazionale Tumori, IRCCS, Fondazione “G. Pascale”, 80131 Naples, Italy

⁷ Ferrara Department of Interventional and Diagnostic Radiology, Ospedale di Lagosanto, Azienda AUSL, 44023 Ferrara, Italy

⁸ Department of Radiology, Ospedale Santa Maria della Misericordia, 45100 Rovigo, Italy

⁹ Department of Medical, Surgical and Experimental Sciences, University of Sassari, 07100 Sassari, Italy

* Correspondence: fabio.pellegrino@aulss9.veneto.it



Citation: Pellegrino, F.; Granata, V.; Fusco, R.; Grassi, F.; Tafuto, S.; Perrucci, L.; Tralli, G.; Scaglione, M. Diagnostic Management of Gastroenteropancreatic Neuroendocrine Neoplasms: Technique Optimization and Tips and Tricks for Radiologists. *Tomography* **2023**, *9*, 217–246. <https://doi.org/10.3390/tomography9010018>

Academic Editor: Emilio Quaia

Received: 16 December 2022

Revised: 22 January 2023

Accepted: 23 January 2023

Published: 27 January 2023



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

Abstract: Gastroenteropancreatic neuroendocrine neoplasms (GEP-NENs) comprise a heterogeneous group of neoplasms, which derive from cells of the diffuse neuroendocrine system that specializes in producing hormones and neuropeptides and arise in most cases sporadically and, to a lesser extent, in the context of complex genetic syndromes. Furthermore, they are primarily nonfunctioning, while, in the case of insulinomas, gastrinomas, glucagonomas, vipomas, and somatostatinomas, they produce hormones responsible for clinical syndromes. The GEP-NEN tumor grade and cell differentiation may result in different clinical behaviors and prognoses, with grade one (G1) and grade two (G2) neuroendocrine tumors showing a more favorable outcome than grade three (G3) NET and neuroendocrine carcinoma. Two critical issues should be considered in the NEN diagnostic workup: first, the need to identify the presence of the tumor, and, second, to define the primary site and evaluate regional and distant metastases. Indeed, the primary site, stage, grade, and function are prognostic factors that the radiologist should evaluate to guide prognosis and management. The correct diagnostic management of the patient includes a combination of morphological and functional evaluations. Concerning morphological evaluations, according to the consensus guidelines of the European Neuroendocrine Tumor Society (ENETS), computed tomography (CT) with a contrast medium is recommended. Contrast-enhanced magnetic resonance imaging (MRI), including diffusion-weighted imaging (DWI), is usually indicated for use to evaluate the liver, pancreas, brain, and bones. Ultrasonography (US) is often helpful in the initial diagnosis of liver metastases, and contrast-enhanced ultrasound (CEUS) can solve problems in characterizing the liver, as this tool can guide the biopsy of liver lesions. In addition, intraoperative ultrasound is an effective tool during surgical procedures. Positron emission tomography (PET-CT) with FDG for nonfunctioning lesions and somatostatin analogs for functional lesions are very useful for identifying and evaluating metabolic receptors. The detection of heterogeneity in somatostatin receptor (SSTR) expression is also crucial for treatment decision making. In this narrative review, we have described the role of morphological and functional imaging tools in the assessment of GEP-NENs according to current major guidelines.

Keywords: gastroenteropancreatic; neuroendocrine; neoplasms; diagnosis; radiology; somatostatin receptor imaging; PET; computed tomography; magnetic resonance; ultrasound

1. Introduction

Gastroenteropancreatic neuroendocrine neoplasms (GEP-NENs) account for approximately 1.5% of all gastrointestinal and pancreatic malignancies [1]. The word “neuroendocrine” derives from the similarity of these cells with the neural crest for the expression of, for example, synaptophysin, neurospecific enolase and chromogranin proteins [2,3]. Although it is known that the incidence is low, the growth is slow, and the prevalence is high, in recent decades, there has been an increase in their incidence rate, also due to the improvement of radiological, nuclear medical, and endoscopic imaging techniques [4,5]. Most of these neoplasms arise sporadically, although some cases occur in complex genetic syndromes. In 60–80% of cases, they are nonfunctional, while in 20–30% of cases, these lesions (e.g., insulinomas, gastrinomas, glucagonomas, vipomas, and somatostatinoma) produce hormones responsible for clinical syndromes [6]. With regard to clinical symptoms, in patients with insulinomas, excessive insulin secretion causes hypoglycemia and hypokalemia; pancreatic polypeptide-secreting tumor (Ppomas) patients have weight loss, abdominal pain, and jaundice. In vipomas lesions, we found watery diarrhea, achlorhydria, and hypokalemia. The 4 Ds, diabetes, dermatitis (necrolytic migratory erythema), deep vein thrombosis, and depression, are typical of glucagonomas, while, for somatostatinomas, there are cholelithiasis, hyperglycemia, and steatorrhea.

Regarding enteric lesions, carcinoid syndrome with watery diarrhea, hypotension, bronchospasm, flushing, and right-sided heart disease is due to serotonin hypersecretion. Since serotonin from small bowel lesions is drained through the portal vein in the liver, where monoamine oxidases inactivate it, this syndrome appears if hepatic metabolism is avoided, for example, if the liver or other distant metastases occur (Table 1).

Table 1. Classification of GEP-NETs based on clinical syndromes.

Hypoglycemia and hypokalemia	Insulinomas
Weight loss, abdominal pain, and jaundice	Ppomas
Watery diarrhea, achlorhydria, and hypokalemia	Vipomas
Diabetes, dermatitis (necrolytic migratory erythema), deep vein thrombosis, and depression	Glucagonomas
Cholelithiasis, hyperglycemia, and steatorrhea	Somatostatinomas
Carcinoid syndrome with watery diarrhea, hypotension, bronchospasm, flushing, and right-sided heart disease	Liver metastasis from small bowel lesions

In this context, the diagnosis may be easy for functioning lesions, while nonspecific symptoms often lead to a delayed diagnosis for indolent lesions. Therefore, an accurate diagnosis requires a multidisciplinary approach, including biochemical analyses and multiple imaging techniques [7–11]. The clinical course of these biologically diverse tumors is highly varied, ranging from indolent to aggressive [12], with histological differentiation and grading being the most significant factors for prognosis and treatment planning [13]. In the prognosis and management of GEP-NENs, the Ki-67 labelling index and mitotic count are critical indicators of the state of tumor proliferation [14,15]. The WHO classifications of NENs have been updated from previous classifications differentiating neuroendocrine tumors (NETs) and poorly differentiated neuroendocrine carcinoma (NECs), according to the Ki-67 index and mitotic count [16–22] (Table 2).

Table 2. The World Health Organization (WHO) 2019 classification for neuroendocrine neoplasms of the gastrointestinal tract and hepatopancreatobiliary organs.

Terminology	Differentiation	Grade	Mitotic Rate *	Ki-67 Index
NET, G1	Well differentiated	Low	<2	<3%
NET, G2	Well differentiated	Intermediate	2–20	3–20%
NET, G3	Well differentiated	High	>20	>20%
NEC, small-cell type	Poorly differentiated	High	>20	>20%
NEC, large-cell type			>20	>20%
MinEN	Well or poorly differentiated	Variable	Variable	Variable

* Mitoses/10 hpf.

In this type of tumor, discrepant tumor grades in the same patient can be determined through intratumoral heterogeneity with different Ki-67 indices in different portions of the same mass or between various tumor sites. Therefore, the aggressiveness of the patient's tumor burden could be misinterpreted with small biopsy specimens in a single location of the lesion [23]. Furthermore, histological samples at a given time do not always represent the grade of the tumor during the entire course of the disease, as the behavior of neuroendocrine tumors is complicated by the fact that these tumors change from low to high grade in time with a more aggressive course of the disease [24]. Two critical issues should be considered in the diagnostic workup of NENs: first, the need to identify tumor presence, and second, to define the primary site and assess regional and distant metastases. In fact, the primary site, stage, grade, and functionality are prognostic factors that the radiologist should assess to guide prognosis and management. Proper diagnostic patient management includes the combination of morphological and functional assessments. Regarding morphological evaluations, according to the consensus guidelines of the European Neuroendocrine Tumor Society (ENETS), contrast-enhanced computed tomography (CT) of the neck–thorax–abdomen and pelvis, including a three-phase examination of the liver, is recommended for the diagnosis, staging, treatment response assessment, and surveillance of primary neuroendocrine tumors. Contrast-enhanced magnetic resonance imaging (MRI), including diffusion-weighted imaging (DWI), is indicated for use to evaluate the liver, pancreas, brain, and bones. Ultrasonography (US) is often helpful in the initial diagnosis of liver metastases as this tool can guide liver lesion biopsies for undefined primary tumors, and contrast-enhanced ultrasound (CEUS) may be used for problem-solving in characterizing liver lesions that remain ambiguous on CT/MRI. In addition, intraoperative US, with or without a contrast medium, represents an efficient tool during surgical procedures, favoring lesion detection. Additionally, it is reported that endoscopic US is the most sensitive modality for diagnosing pancreatic NENs (PNENs), also allowing for lesion biopsies [25]. Regarding functional assessments, somatostatin receptor imaging or 18FDG-PET/CT represent essential tools during the staging, response evaluation, and surveillance [25,26]. In this article, we have reported on the state-of-the-art imaging techniques for GEP-NENs, focusing on the role and optimization of morphological and functional techniques in managing these patients. Our aim is to assist the imaging specialist in choosing and using imaging techniques based on the clinical scenario of the GEP-NEN patients.

2. Diagnosis, Staging, and Risk Assessment

A histological diagnosis is mandatory in all patients, and can be carried out on surgical specimens or metastasis biopsies. However, some factors can reduce the adequacy of sampling related to the characteristics of the lesion, such as localization in the pancreatic body or the tail, rather than in the head, or a small size, rich stromal fibrosis, or cystic and necrotic components, but also operative factors, such as the initial experience of

endosonographers and cytopathologists [27]. Disease stage and tumor grade are the two major independent prognostic features that should be evaluated. Therefore, the role of imaging is to detect the primary site to obtain the stage of the disease and to identify the proper patient treatment [28]. In this scenario, functional and morphological evaluations may be used complementarily to overcome the limits of one with the other and vice versa (Table 3). PET/CT or SPECT/CT imaging using adequate tracers allows to accurately evaluate the extension of the disease, both in the initial staging and in the follow-up, and can also identify a primary occult tumor, a task that is sometimes challenging but helpful in optimizing the therapeutic strategy, especially in patients with metastasis disease. Furthermore, functional imaging allows for the noninvasive characterization of the functional status and tumor heterogeneity based on the analysis of the uptake intensity of target-specific radiotracers [29]. Moreover, functional imaging can offer an improved prognostic stratification and refinement of therapeutic strategies, enabling a personalized theranostic approach to managing GEP NETs. Contrast-enhanced CT and MRI supply detailed anatomical information about the primary tumor site and identify regional and distant metastases, information required for optimized surgery, treatment selection, and the identification of persistent or recurrent disease. Finally, endoscopic procedures, such as endoscopic ultrasound, have been used with great success to identify lesions that might otherwise have escaped imaging modalities [30].

Table 3. Comparison of imaging techniques for GEP-NEN diagnosis.

	Advantages	Disadvantages	Use
US	-no ionizing radiations -widely available -inexpensive	-interoperator variability -not recommended for other sites in the gastrointestinal tract -low sensitivity in pancreatic lesions	-initial diagnosis of liver metastases -surveillance in some patients with liver metastases -liver lesion characterization with contrast-enhanced ultrasound (CEUS) -tool to guide liver lesion biopsies -intraoperative use for lesions detection
EUS	-possibility of fine-needle biopsy of the lesion with Ki-67 evaluation -increases the overall PNEN detection rate after a CT scan	-inadequacy of sampling due to site (pancreatic body or tail), small size, rich stromal fibrosis, or cystic and necrotic components -inexperience of endosonologists and cytopathologists	-detection of small pancreatic NENs -histological diagnosis -intratumoral vascularity with contrast-enhanced endoscopic ultrasound
CT contrast enhanced	-good sensitivity and specificity -widely available	-ionizing radiation	-diagnosis, staging, treatment response assessment, and surveillance of primary neuroendocrine tumors -lung lesions assessment
MRI contrast enhanced	-no ionizing radiation -ability to characterize lesions using pre and postcontrast sequences and DWI -cholangiopancreatography sequences to assess the relationship between the lesion and the pancreatic duct	-low sensitivity for detecting small lesions in the duodenum, stomach, and small intestine	-liver, pancreas, brain, and bone evaluation

Table 3. *Cont.*

	Advantages	Disadvantages	Use
Fluorodeoxyglucose PET	-complementary information distinguishing between slowly proliferating and aggressive tumors (higher glucose metabolism in G3 and high G2 NENs)	-ionizing radiation -low sensitivity for neuroendocrine tumors	-optional in NEN evaluation prognostication (worse prognosis if FDG uptake) and post-therapy assessment
68Ga-DOTA-SSA PET/CT	-good sensitivity and specificity	-low sensitivity for insulinomas	-staging with primary tumor location and metastatic detection -restaging with the assessment of residual, recurrent, or progressive disease -patient selection for peptide receptor radionuclide therapy

CT: computed tomography; MRI: magnetic resonance imaging; PET: positron emission tomography; US: ultrasound; EUS: endoscopic ultrasound; SSA: somatostatin analog.

2.1. Functional Assessment

Regarding lesion detection, 68Ga/64Cu-DOTA-somatostatin analogue (SSA) positron emission tomography (PET), in combination with CT (PET-CT), has the higher sensitivity for the detection of most types of NENs, representing, according to the ENETS guidelines, the diagnostic tool of choice to localize the disease in noninsulinoma pancreatic NETs. In a small portion of insulinoma patients (<5–10%), the overexpresses of glucagon-like peptide-1 (GLP-1) can be a target for scintigraphy with radiolabeled GLP-1 receptor analogues [31]. However, this method is confined to research settings. SSTR scintigraphy (SRS) should be carried out only if PET-CT is not available, since this method has a lower sensitivity [25].

According to the European Society of Medical Oncology (ESMO) guidelines [32], the use of PET with [18F] fluorodeoxyglucose (FDG) is optional in NEN evaluation. However, FDG is the tracer of choice for G3 and high G2 NENs, since these entities show a higher glucose metabolism and less SSTR expression compared to low-grade NENs. The presence of FDG uptake in NEN patients at PET-CT suggests a worse prognosis.

2.2. Functional Tools

2.2.1. Somatostatin Receptor Imaging (SRI)

SRI can be obtained through scintigraphy, using a gamma camera or, more recently, PET. It is mandatory to perform SRI to obtain information on the status of the somatostatin receptor in lesions to assess the patient's suitability for treatment with somatostatin analogues. Somatostatin is a regulatory peptide widely distributed in the human body, and its action is mediated by membrane receptors, of which five subclasses (sst1-sst5) have been cloned [32]. The subtype predominantly expressed by GEP-NENs is sst2, and its cellular expression decreases with an increasing proliferation (Ki-67) and, consequently, tumor uptake on somatostatin receptor imaging [33,34]. For 18FDG-PET/CT metabolic imaging, the situation is speculatively opposite, with lesions usually becoming more 18FDG avid as proliferation increases. For the detection of higher-grade NETs, 18FDG-PET/CT may, therefore, be preferred. However, studies show that using both somatostatin receptor imaging and 18FDG-PET/CT in a complementary manner increases sensitivity [34,35].

Somatostatin receptor scintigraphy (SRS) using 111In-DTPA-octreotide (OctreoScan) in its original form provides whole-body planar images, while, in modern practice, this image is merged with single-photon emission computed tomography (SPECT) and CT. Diagnostic accuracy is, thus, improved by exploiting the specificity of OctreoScan and the anatomical details provided with SPECT/CT [36]. 111In-Pentetetrotide, acquired as SPECT/CT, can be used for the routine diagnostic imaging of nonfunctional NEN-GEPs, with a significant impact on treatment planning, as it exhibits higher sensitivity and accuracy values than

conventional imaging, with an added value of 35.6%. It was also shown to change patient classification and management in 27.9% of cases, reducing disease in 9.6% of cases compared to conventional imaging [37].

The newest somatostatin receptor-based imaging technique employs the positron emitter 68Ga to label various somatostatin analogues [38]. The 68Ga positron emitter has been used to label somatostatin analogues for PET. Two different preparations of octreotide, 68Ga-DOTATOC(DOTA,1-Nal(3)-octreotide), and 68Ga-DOTANOC(DOTA,1-Nal(3)-octreotide), and one of octreotate, 68Ga-DOTATATE(DOTA, and Tyr(3)-octreotate) are the most commonly used [39–42]. The preparations have different affinities with various somatostatin receptor (sst) subtypes. However, the differences between the three preparations seem marginal for the routine clinical use of PET [43]. The main uses of 68Ga DOTA conjugated peptide-binding SSTR are in staging with primary tumor location and metastatic detection, in restaging with the assessment of residual, recurrent, or progressive disease, and, finally, in patient selection for PRRT [37]. PET with 68Ga-labeled somatostatin analogues has been shown in numerous studies to be superior to SRS [32,44–47] and CT/MRI [23–28], although similar results were found in a study comparing 68Ga-DOTANOC and SRS [48]. PET with 68Ga-labeled somatostatin analogues also showed better results than 18F-DOPA [49,50]. The spatial resolution is higher in PET versus SRS (0.5 cm versus 1.5 cm), and tissue contrast is better. Furthermore, PET offers logistical advantages, because the more favorable kinetics of 68Ga-labeled preparations allows for images to be obtained as early as 30–60 min after injection. Finally, the sensitivity reported in various studies of PET/CT ranged from 86% to 100%, and the specificity ranged from 79% to 100% [50–58] for all panNETs, except for insulinomas, in which case, the sensitivity was only 25% [59]. Accordingly, current major guidelines consider 68Ga-labeled somatostatin analogs PET/CT to be the first-line diagnostic procedure for the staging or restaging of any noninsulinoma panNET cases for detecting an unknown primary tumor site or early recurrence, and for the evaluation of patients undergoing radiometabolic peptide receptor therapy (PRRT) [25,60,61].

2.2.2. Positron Emission Tomography (PET)

In addition to 68Ga-labeled somatostatin analogues, other PET tracers exist for NEN imaging. NENs were previously described as APUDOMAs, and specific PET tracers were developed based on these properties. The most commonly used radiotracer in PET imaging is 18F-FDG, which evaluates the glucose metabolism. It showed imaging results superior to that of octreotide. However, this method is generally considered when other imaging techniques fail or demonstrate contradictory results or limited equivocation. It has been suggested that it may provide complementary information for distinguishing between slowly proliferating and aggressive tumors [62,63]. Additionally, assessing the glucose metabolism with 18F-FDG may allow for a prognostic prediction, including overall and progression-free survival. It may also be useful in the diagnosis of aggressive disease, particularly in high-grade tumors, with better prognostic values and greater sensitivity in identifying the extent of the disease. As in all 18F-FDG-avid tumors, the analogue of glucose accumulates only in high-grade neuroendocrine neoplasms, and, therefore, FDG-PET has not been considered useful for imaging in most patients with NENs [64–66]. In more recent studies, however, FDG-PET has been shown to provide additional information when combined with 68Ga-DOTATATE [67,68] and SRS [34], also providing helpful information in the preoperative diagnostic workup regarding the behavior and aggressiveness of the tumor [69]. It has also been reported that FDG may positivity predict early tumor progression [70] and be associated with a higher risk of death [71]. In a recent study on pancreatic NENs, FDG-PET was used to define tumors with a higher malignant potential [72].

2.3. Morphologic Imaging and Tools

Although the limitations of morphological imaging are well known in the management of patients with NENs, the optimization of traditional techniques, technological improvements, and the introduction of new functional sequences in magnetic resonance imaging have achieved high sensitivity in detecting these entities. Furthermore, morphological imaging is essential in local staging to identify patients suitable for surgical resection, since this approach is considered, according to the ESMO guidelines, the first-line treatment for several GEP-NENs.

2.3.1. Computed Tomography

CT is usually the initial imaging technique in the oncology field, where the CT examination is often repeated in the course of treatments to conduct a response assessment due to its good sensitivity, specificity, and availability [73–76]. Optimizing techniques are mandatory for assessing GEP-NENs, requiring multiphase CT protocols with a contrast medium [77]. Scans should be performed before the injection (useful for discriminating calcifications), in the arterial phase (typical NEN enhancement and mandatory to increase the detection of small functioning pancreatic NENs, so as to assess the encasement of the hepatic, splenic, and mesenteric artery), and the portal venous phase (for liver imaging and the delineation of parenchymatous organs, as for the encasement of mesenteric and portal veins) [78]. For pancreatic lesions, the contrast study protocol should include the delayed phase for the detection of delayed enhancement presented by some fibrous tumors. CT is an excellent method for detecting complications secondary to GEP-NETs, such as a bowel obstruction, intussusception, or desmoplastic reaction, which are life-threatening diseases [79–87]. The right timing of the arterial phase is crucial for appropriate imaging. NENs have a strictly arterial blood supply and begin taking the contrast medium as soon as it reaches the lesion through the arterial system. With regard to pancreatic lesions, in order to obtain an optimal contrast between the lesion and parenchyma, a flow rate of 4 mL/s has been suggested, with bolus triggering on the descending aorta, and a delay between the trigger and arterial scan of 12 s. When focusing on liver imaging, a slightly later time may be chosen for a more intense impregnation of the lesions. Early scanning after the initiation of a contrast injection (first-stage arterial or CT angiography) may occasionally be required prior to surgery to show arterial anatomy. With postprocessing, the vascular anatomy can also be shown in three-dimensional images, MIP (maximum intensity projection), and using the volume rendering technique. In the late arterial phase (or portal inflow phase), after approximately 10–15 s, the small arterial branches are enhanced and this is the best phase to delineate hypervascular liver metastases and pancreatic NENs. In the venous (or portal venous) phase, approximately 60–90 s after the start of the injection, the contrast medium passes through the capillaries and reaches the veins, including the portal vein, with improved visualization of normal hepatic parenchyma. In this phase, hypovascular liver metastases are better visualized. Since current CT scanners produce images of 1 mm or less, coronal and sagittal two-dimensional images can be obtained with high resolution through multiplanar reconstructions (MPRs) [25,88–90] (Table 4). Such reconstructions can be helpful, for example, to show the relationship between a pancreatic NEN and the pancreatic duct. The use of dual-energy computed tomography (DECT) in diagnosis and follow-up in patients with NETs is growing. Thanks to the use of different X-ray spectra, e.g., 80 and 140 kV, DECT can differentiate tissue samples simultaneously, providing iodine density quantification on a single contrast-enhanced scan without increasing the patient's radiation dose and, as a result, produces images with greater contrast between the lesion and surrounding tissue [91–96]. Noah et al. showed that CT iodine maps could increase radiologists' confidence in studying these tumors [92]. DECT has shown a higher sensitivity detecting pancreatic insulinomas compared to conventional CT (95.7 vs. 68.8%) [97]. According to the ENETS guidelines for jejunum and ileum NENs, CT enteroclysis is the diagnostic tool that should be chosen for small bowel lesions if CT is inconclusive. CT enteroclysis showed a sensitivity of approximately 92%, and a positive predictive value

(PPV) of 95% [98–100]. The patient's preparation is critical for the CT enteroclysis, and involves the use of a neutral contrast medium (water and methylcellulose) with a nasojejunal conduit and the administration of spasmolytics [25]. The contrast study includes an arterial phase (approximately 25 s after the start of the intravenous contrast injection) and a venous phase (60 s after). In addition, extraluminal findings may be assessed as metastases in solid organs or in lymph nodes as a desmoplastic reaction. When the neutral contrast is administrated through drinking approximately 1L of oral negative contrast, we could perform a CT enterography that shows a sensitivity similar to CT enteroclysis [101,102] (Figure 1).

Table 4. CT protocols for diagnosis and staging of GEP-NENs.

Abdomen Multidetector CT Protocol	
Voltage	120 kVp
Effective amperage	200 mAs
Rotation time	0.5 s
Detector collimation	1.5 mm
Section thickness	3.0 mm
Pitch	0.75
Increment	1.5 mm
Coverage	Image from the 11th vertebral body through the iliac crest
Oral contrast material	Negative oral contrast material (500 mL of water 30 min before examination and 250 mL of water immediately before examination)
Nonenhanced CT	
Contrast-enhanced CT	100–125 mL isomolar or osmolar iodinated contrast material (370 mg/mL) at 4–5 mL/s
Image Acquisition Phase	
Arterial phase	15–25 s (CT angiography in preoperative setting) 25–30 s
Pancreatic parenchymal phase	40–45 s
Portal venous phase	60–70 s
Multiplanar reformation	Axial, sagittal, and coronal planes; section thickness, 3 mm
Thorax/Neck Multidetector CT Protocol	
Thorax–Neck–Abdomen	Amount of contrast media and injection rates adjusted to what is required to perform a proper CT of the abdomen
Neck	1.5–2 mL/kg body weight of contrast media 300–350 mg/mL injected at 2.5 mL/s using a 40 s scanning delay
Thorax	1–1.5 mL/kg body weight of contrast media 300–350 mg/mL injected at 1.5 mL/s using a 60 s scanning delay
CT Enteroclysis	
Patient preparation	2000 mL of hyperosmolar fluid, such as mannitol, with a 50% concentration, or, alternatively, warm tap water administered through a nasogastrojejunal tube by using a dedicated (150–200 mL/min rate) power injector Intravenous glucagon or anticholinergic drug (recommended)
Contrast-enhanced CT	120–150 mL isomolar or osmolar iodinated contrast media at 3 mL/s Arterial phase using a 25 s scanning delay Venous phase using a 60 s scanning delay

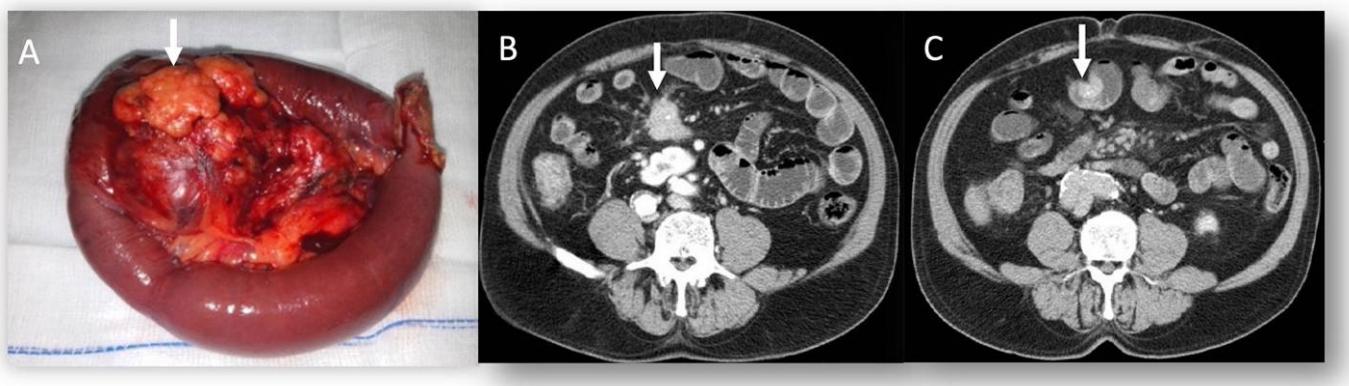


Figure 1. Sample size (A) of small bowel NEN with desmoplastic reaction (arrows in (A) and (B)), evaluated on CT (B) in portal phase of contrast study. In (C), arrow shows the intraluminal small tumor as an enhanced polypoid lesion.

2.3.2. Magnetic Resonance Imaging

According to the ESMO guidelines, MRI is the diagnostic tool of choice for liver and pancreas assessments due to its accuracy in the differential diagnosis between primary hepatic malignancy, liver metastases, and benign lesions, such as hemangiomas [103]. However, during the presurgical phase, given the better accuracy of CT in vascular evaluations, the two methods should be complementary. MRI also has a higher accuracy than CT for bone and brain evaluations [104–106]. Otherwise, CT is preferred for the lungs. The MRI sensitivity for PNETs detection is 79% (range 54%–100%) [107–109], while the mean sensitivity of MRI for the detection of liver metastases is 91% (range 82%–98%) [110–114]. In 2015, Flechsig et al. demonstrated the greater sensitivity of MRI in the search for liver metastases, comparing it with both CT and PET with ⁶⁸Ga-DOTATOC. MRI, where PET fails, would seem capable of detecting metastases thanks to DWI, although a differential diagnosis with focal nodular hyperplasia (FNH) can sometimes be challenging. The sensitivity obtained with DWI and T2-weighted images would seem comparable to those obtained with contrast, which, therefore, can be used to complete the investigation [115,116]. The standard abdominal study protocol can be correlated to the target.

Regarding pancreatic lesions, it generally includes T1- and T2-W conventional sequences, T2-W fat-suppressed (FS) sequences, DWI, cholangiopancreatography sequences, and pre and postcontrast T1-W sequences. Conventional sequences allow for the identification of a lesion's structural characteristics (solid vs. cystic). T2-W FS and DWI increase the detection rate and allow for a better characterization of small lesions, allowing to identify multiple lesions in familial syndromes. Cholangiopancreatography sequences assess the relationship between the lesion and the pancreatic duct for correct characterization and better surgical management. Regarding postcontrast sequences, for pancreatic lesions, due to the retroperitoneal side, it is possible to consider studying these protocols: contrast-enhanced (CE) MRI and dynamic contrast enhancement (DCE) MRI. In CE-MRI, T1-W FS sequences are acquired in specific phases (e.g., arterial, pancreatic, and venous phases), while, in DCE-MRI, more sequences are acquired continuously, without pause, for approximately 5 min. This method allows for a qualitative, semiquantitative, and quantitative evaluation. Usually, CE-MRI is the standard protocol, while DCE-MRI is preferred in research settings.

Regarding liver metastasis assessments, the standard protocol includes T1- and T2-W conventional sequences, T1-W in-phase and opposed phase, T2-W FS sequence, DWI, and postcontrast T1-W sequences. Similarly to the above, conventional sequences allow for identifying a lesion's structural characteristics. T2-W FS and DWI increase the detection rate of small lesions and allow for better characterization. Regarding contrast protocol studies, two subtypes of contrast agents can be employed: a nonspecific contrast medium that distributes into the vascular and extravascular extracellular spaces, and liver-specific agents, taken up by liver cells. The advantage of the liver-specific agent is correlated with the possibility of increasing the detection rate of small liver metastases, considering the pharmacokinetics of these media; during the hepatobiliary phase, normal liver parenchyma is uniformly hyperintense, while primitive or secondary lesions are not hyperintense due to missing normal hepatocytes [117,118]. Despite the proven advantages of hepatospecific contrast agents, recent studies showed that a suboptimal image quality is frequently observed in arterial phase imaging with Gd-EOB-DTP, which could negatively affect the characterization of hepatic lesions. Consequently, these agents should be chosen in presurgical settings, while, during characterization, we must carefully consider the pros and cons [119–125].

Regarding small bowel lesions, MR enteroclysis provides a better assessment of mucosal lesions compared to MR enterography [126]. The study protocol should include, in axial and coronal planes, steady-state precession sequences, since these have a lower sensitivity to motion artifacts. These sequences allow for a rapid overview of the entire abdomen. In addition, coronal and axial T2-W, fast or turbo spin echo sequences, based on the half-Fourier reconstruction technique, should be performed to obtain high contrast between the lumen and the bowel wall. However, these sequences are susceptible to intraluminal motion due to flow void, so a radiologist should have familiarity with these artifacts.

A precontrast and postcontrast coronal T1-W FS gradient-echo sequence are applied during the arterial and the venous phase of the contrast study, followed by similar axial sequences covering the entire abdomen. The addition of DWI has been suggested, with at least two b values. Although, currently, CT enteroclysis is the primary imaging modality in small bowel tumors, few data are available on the comparison of the effectiveness of CT and MR enterography techniques.

Regarding rectal lesions, the study protocol includes T2-W sequences and DWI, while T1-W contrast sequences are optional. Specific parameters should be followed to achieve optimal high-resolution sequences, including a small FOV, slice thickness (no more than 3 mm), and correct scan plane alignment (perpendicular to the rectal wall at the level of the tumor).

Concerning bone metastases, commonly used protocols include a combination of T1-W spin echo and fat-suppressed short-tau inversion recovery (STIR) sequences. Recently, obtaining data in whole-body DWI studies has been attracting significant interest, although most studies are aimed at prostate metastases. According to a neuro-oncology study group [127], in brain metastases, the “minimum standard” recommended pulse sequences include: (i) parameter matched pre- and postcontrast inversion recovery (IR)-prepared isotropic 3D T1-weighted gradient echo (IR-GRE); (ii) axial 2D T2-weighted turbo spin echo acquired after the injection of a gadolinium-based contrast agent and before postcontrast 3D T1-weighted images; (iii) axial 2D or 3D T2-weighted fluid-attenuated inversion recovery; (iv) axial 2D, three-directional diffusion-weighted images; and (v) postcontrast 2D T1-weighted spin echo images for increased lesion conspicuity. Recommended sequence parameters were provided for both 1.5T and 3T MR systems. An “ideal” protocol was also provided, replacing IR-GRE with 3D TSE T1-weighted imaging pre- and post-gadolinium, and is best performed at 3T, for which a dynamic susceptibility contrast perfusion is included.

Regardless of the target, to improve the temporal resolution (number of phases) and avoid artefacts, acquisition acceleration techniques and motion compression techniques are commonly used. Multiphasic imaging and apnea acquisition can be used, but the time between the contrast injection and acquisition is critical following the times described in CT [128]. Furthermore, if we are looking for a PNEN, the use of thin-layer acquisitions is recommended [129]. The use of diffusion also involves the use of respiratory gating or breath compensation techniques with parallel imaging and a b-value above 500, but also the acquisition of at least two b-values or more, for example, (0, 50, 600) [128,130], since it is, thus, possible to calculate ADC (apparent diffusion coefficient) maps [115,131]. The literature on this subject is still limited, but the increased ADC values seem to be linked to those of Ki-67 and also to the evaluation of responses to TACE (transarterial chemoembolization) [131].

Conventional DWI is based on a monoexponential model and assesses the water molecule motion according to a Gaussian approach [132–135].

However, it is known that water molecules diffusion within tissue follows a non-Gaussian model; thus, a non-Gaussian approach, named diffusion kurtosis imaging (DKI), was proposed by Jensen et al. [136].

Using the DKI, it is possible to calculate the kurtosis median coefficient (MK) and to assess the variation of diffusion behavior with a Gaussian to a non-Gaussian approach and the diffusion coefficient (MD), which evaluates the correction of the non-Gaussian bias. DKI allows us to obtain more data on tissue structures than DWI does. However, DKI requires high-quality images at b -values greater than 1000 s/mm^2 and a high signal-to-noise ratio (SNR) [137].

Shi et al. [107] assessed DKI performance in differentiating pancreatic ductal adenocarcinomas (PDACs) from solid pseudo papillary neoplasms (SPNs) and PNETs, showing that the accuracy rate with DKI was higher than that of a subjective diagnosis alone [138]. In addition, Le Bihan et al. [139,140] proposed a biexponential model, the intravoxel incoherent motion (IVIM), to obtain data on tissue perfusion. By using the IVIM model and multiple, sufficiently low b values ($<200 \text{ mm}^2/\text{s}$), not only can pure diffusion characteristics (D) be separated from pseudodiffusion caused by microscopic circulation in tissue, but perfusion characteristics (pseudodiffusion coefficient (D^*)) and their proportions (perfusion fraction (f)) can also be extracted.

Zeng et al. showed that conventional DWI and IVIM models are valuable tools to differentiate nonhypervascular PNETs from PDACs. D^* showed better performance than f and ADC [141]. Improvements in scanner performance and the optimization of pulse sequences have reduced acquisition times and increased the use of whole-body MRI (WB-MRI) in several clinical scenarios [142]. Few studies report [58,143] on the use of WB-MRI and PET in PET/MRI hybrid scanners for NET imaging [144,145]. In a total study time of 1h or less, the exam protocols include the neck–thorax–abdomen (and brain when needed), with the acquisition of DWI and IV contrast-enhanced images of the liver and pancreas. Finally, although MRI is used less frequently than US or CT as a guide for minimally invasive interventional procedures [146–158], its use has increased. It has many advantages, such as a lack of ionizing radiation, real-time MR fluoroscopy placement, high resolution, capacity to display small tumors with increased sensitivity, and the monitoring of thermal effects. It can be combined with diffusion-weighted imaging or MRI contrast agents to visualize more difficult lesions [159,160] (Figure 2).

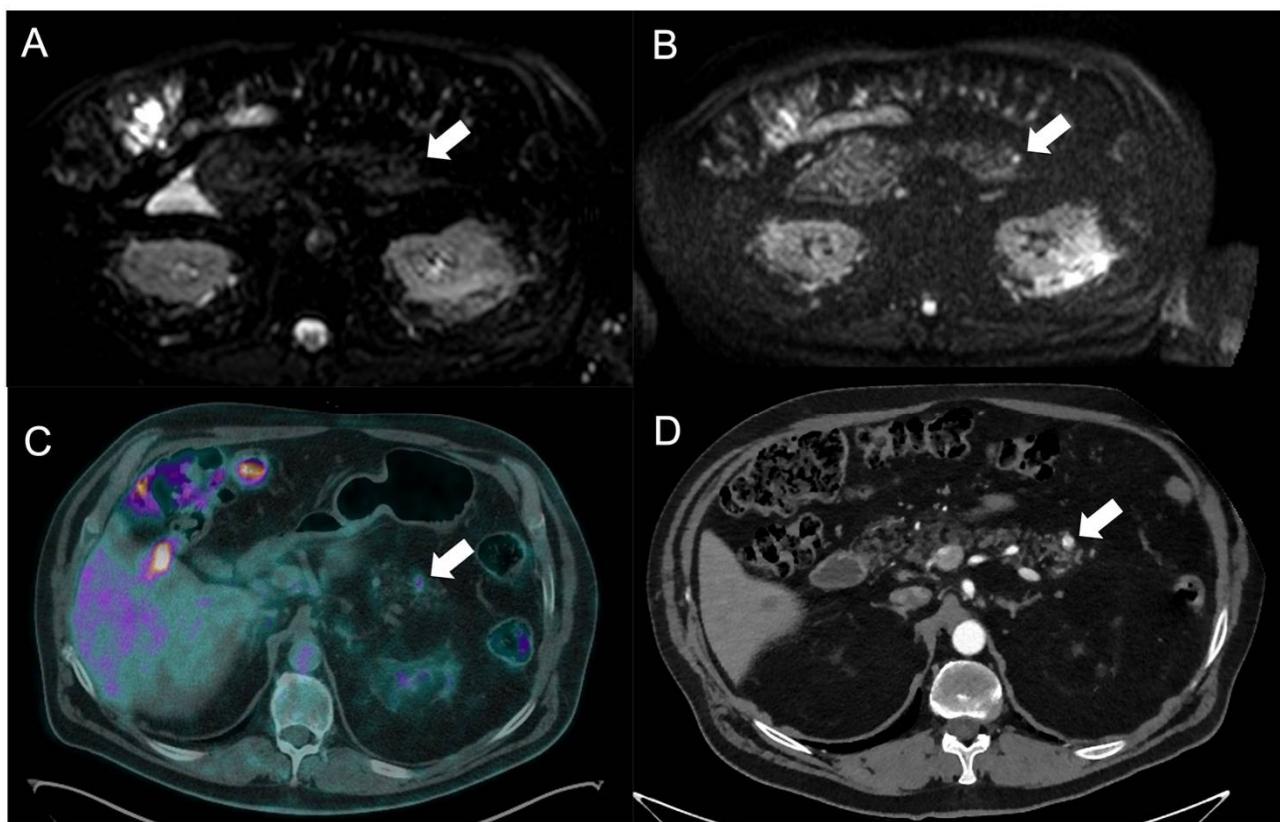


Figure 2. The pancreatic NEN lesion at the tail is highlighted with an arrow. DWI at low and high b-values, respectively, in Subfigures (A,B), showing the restricted diffusion detecting the little lesion, typically characterized by ^{68}Ga uptake in PET/CT (C) and high arterious enhancement at CT (D).

2.3.3. Ultrasonography and Contrast-Enhanced Ultrasound (CEUS)

Although the abdominal US is a widely used, easy-to-perform, and inexpensive diagnostic tool [161], its role in GEP-NEN evaluations is limited and, in some cases, controversial.

According to the Polish Network of Neuroendocrine Tumours, abdominal US is an excellent tool in liver metastasis assessments with a sensitivity of 85–90%, and the possibility to use a contrast medium (CEUS) increases the sensibility and allows to characterize liver lesions that remain equivocal on CT/MRI [162]. Additionally, although US is not a technical standard for therapy monitoring, it may be helpful in surveillance in some patients with liver metastases not clearly visible on CT and MRI or in young patients, to reduce the radiation dose. The differences in sensitivity are due to anatomical conditions, cooperation with the patient and the experience of the physician [162]. In addition, it may be challenging in patients with a large tumor burden to obtain an overview of an enlarged liver in which most of the normal parenchyma is replaced with the tumor. Of contrast, abdominal US should not be proposed in pancreatic lesions since its sensitivity is of 13% to 27%. In comparison, endoscopic ultrasound (EUS) is the current optimal diagnostic tool to detect small PNETs with 86% (range 82%–93%) sensitivity and 92% (range 86%–95%) specificity [163]. EUS plays a decisive role in defining GEP-NENs of the gastrointestinal wall, as it provides information on the size, depth of invasion, and local-regional metastases [164]. The EUS-guided biopsy can also provide a definitive diagnosis and useful information (for example, the evaluation of the nuclear protein Ki-67, which appears to be a proliferative index) for correctly managing this type of lesion [165]. In addition, the EUS can accurately select the ideal candidates for the endoscopic resection. EUS plays a decisive role in the setting of pancreatic NENs. It can help locate the tumor correctly when other noninvasive procedures have failed and provide useful additional information (e.g., the distance to the

pancreatic duct and the Ki-67) for the best therapeutic management (surgery, conservative approach, and type of anticancer therapy in cases of unresectable tumors). EUS is an essential method for the detection of small PNENs [166]. In addition, contrast-enhanced endoscopic ultrasound using second-generation contrast agents allows for the assessment of pancreatic tumor intratumoral vascularity by permitting the differentiation of pancreatic ductal adenocarcinoma from PNENs, with the latter generally showing a hypoenhancement pattern, while PNENs typically show a pattern of hyperenhancement [167]. Indeed, it has been shown that CT fails to find 68.4% of PNENs <10 mm and 15% of those ≤20 mm in diameter [168]. In a meta-analysis, James et al. reported that preoperative EUS increases the overall PNEN detection rate by >25% after a CT scan, with or without additional investigative examinations, such as MRI or ultrasound [169]. Additionally, the ability to perform EUS-guided tattooing in pancreatic lesions can help surgeons find cancer and avoid primarily destructive surgeries. Finally, EUS-guided therapies (e.g., alcohol ablation), especially in patients unsuitable for surgery, are under investigation, and could represent a future field of interest [131]. The transducer is placed directly on the organ's surface in perioperative ultrasound (IOUS) and is mandatory in the surgical resection of a pancreatic NEN to facilitate the diagnosis of liver metastases during surgery. IOUS facilitates lesion detection and localization in the pancreas and liver, and, according to the ESMO guidelines, it is mandatory before pancreatic resection in MEN1 syndrome patients [170].

Generally, on US images, panNENs present as well-defined, solid, and heterogeneous hypoechoic lesions, and some of them can present with cystic regions [171,172]. Liver metastases often appear hyperechoic compared to the surrounding liver parenchyma, although they can also manifest with a hypoechoic and targetoid appearance with increased vascularity on Doppler US (Figure 3).

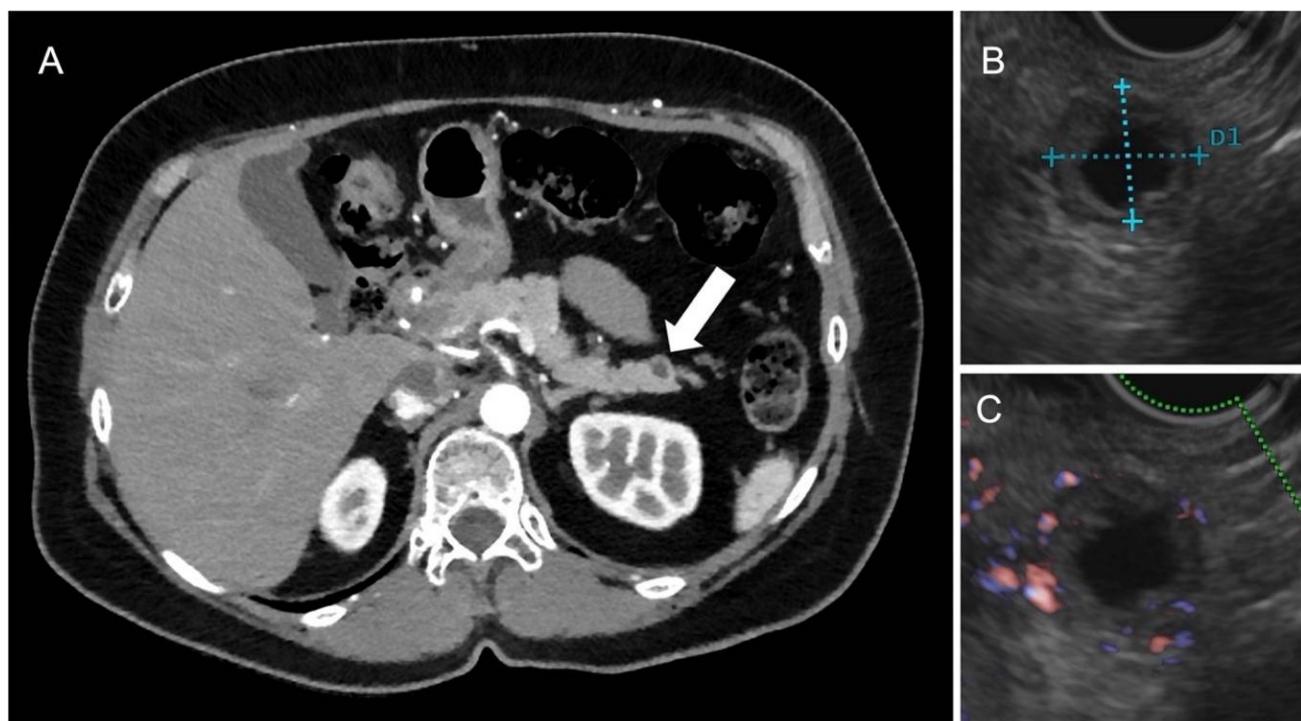


Figure 3. Arterious CT scan (A) demonstrating a hypodense lesion at pancreatic tail (arrow), which, at echoendoscopy, showed anechoic content (B) with hypervascularized thick walls at Doppler, and after a bolus of ultrasound contrast agent (C). The tumor was surgically removed and diagnosed as G1 NEN tumor with cystic appearance.

3. Clinical Setting

3.1. Pancreatic NENs

According to the ESMO guidelines, surgical resection is the treatment of choice in G1 and G2 NENs. In functional lesions, clinical symptoms should be managed before any approach.

Regarding PNENs, several studies showed the safety of a watch-and-wait approach instead of surgical resection for asymptomatic NF-PNETs <2 cm [60,173,174].

Regardless, surgical resection is the front-line treatment in young patients, in cases of local invasiveness, and in the presence of functioning PNETs, irrespective of tumor size. A parenchyma-sparing resection (e.g., enucleation or central pancreatectomy) should be considered in cases of the absence of signs of invasiveness, as the dilation of the main pancreatic duct and nodal involvement. If there is nodal involvement, a standard pancreatectomy with lymphadenectomy is mandatory [170].

In this scenario, imaging may identify single or multiple lesions and define the localization, size, functional activity, and signs of local invasiveness. It, therefore, appears clear that there is not one technique better than another, but there is the necessity of multimodal imaging, where the morphological ones and vice versa support the functioning techniques. Indeed, while functional techniques allow to establish the presence or absence of one or more lesions and the functional status, MRI efficiently identifies the localization, the relationship with the main duct and the presence of liver metastases [112,175,176]. CT allows a better vascular assessment even in the presence of anatomical variants, so as pulmonary metastases [177,178] (Figure 4).

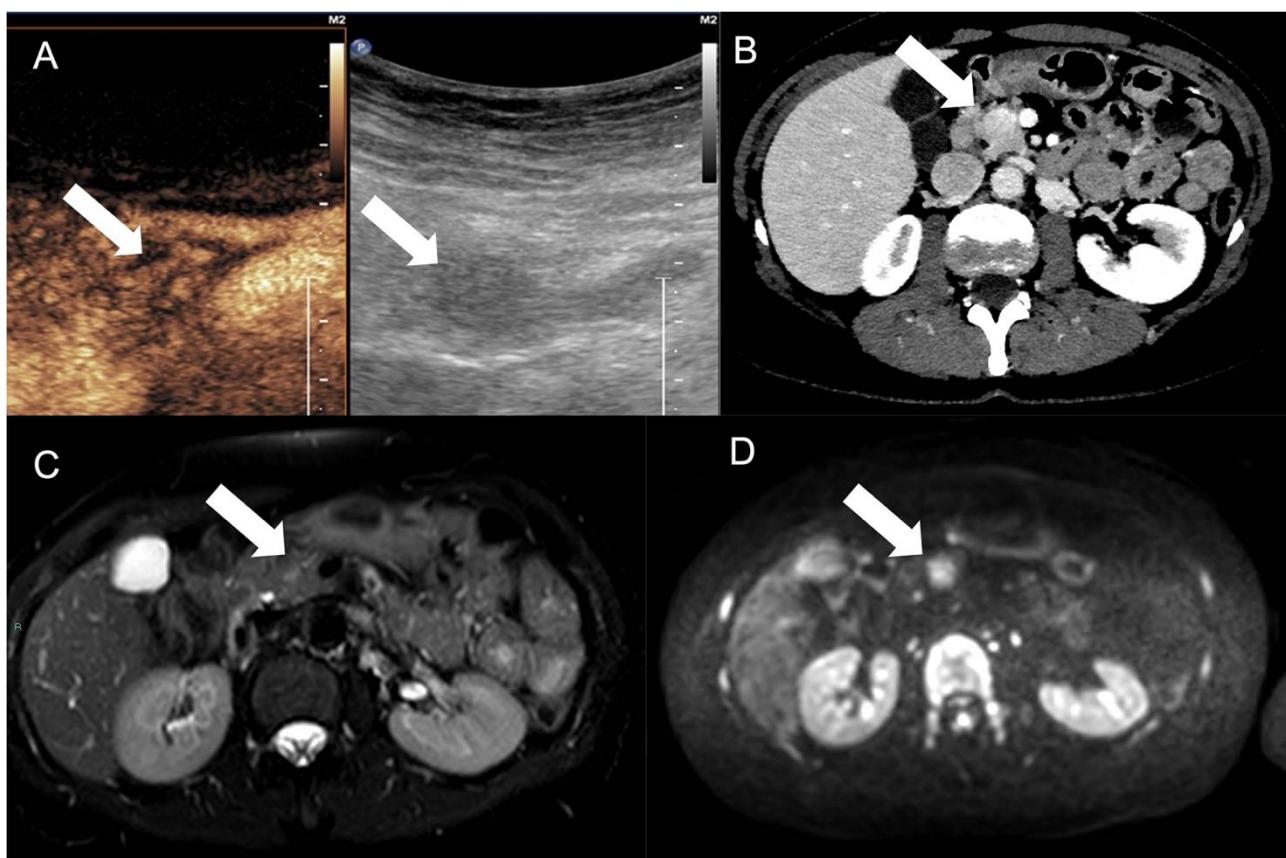


Figure 4. A pancreatic NEN tumor (arrow) was discovered as hypoechoic mass at ultrasound, showing high vascularization at CEUS (A) and CT (B). At MRI examination, the mass was not clearly defined with T2W fat-suppression sequence (C), while the restriction at diffusion imaging improved the definition of the mass (D).

3.2. Liver Metastases

Surgery is a recommended treatment option for most well-differentiated G1 and G2 tumor neuroendocrine liver metastases (NELMs) in unilobar and limited diseases [14]. The surgical approach is influenced, in fact, by lesion size and location, disease stage, and the patient's symptoms [179]. Due to the frequent bilobar and multifocal localization, less than 20–30% of patients are candidates for liver resection with curative intent. Liver transplantation as a therapeutic option is feasible in selected patients with unresectable NELMs [180–183]. Moreover, cytoreductive ablative therapies, in addition to surgical resection, can offer improved survival and quality of life at 5 years compared to patients who do not undergo surgery (70%–90% vs. 50%) [184]. For patients with NELMs who are not candidates for surgery for high liver involvement or sites that are inaccessible or refractory to medical therapy, intra-arterial treatments, such as transarterial embolization (TAE), transarterial chemoembolization (TACE), and selective internal radiation therapy (SIRT), are safe and effective options for gaining control of the disease. These techniques allow for biological and symptom improvements in selected patients, or are palliative treatments in patients with multiple, bilateral, or unresectable NELMs, reducing tumor volume and endocrine secretion [185]. Intra-arterial treatments are effective because NELMs are usually hypervascular tumors with a predominant arterial supply. Intra-arterial therapies in NELMs have demonstrated efficacy in a subset of NELMs (i.e., those from midgut origin) [186], while they must be associated with intra-arterial chemotherapy for liver metastases from colorectal cancer [187]. NELMs may be linked to biological syndromes due to their endocrine secretion. Hence, in addition to the local control of tumor growth, one of the goals of the intra-arterial treatment is to control the endocrine secretion of NELMs, which is not the case for liver metastases of other origins. An improvement in symptoms was observed in 60–90% of patients [188–191]. The mass effect due to liver involvement through NELMs decreased in 100% of patients [192,193]. The therapeutic decision should be discussed in dedicated multidisciplinary meetings [194], and at least one dedicated liver CT or MRI examination should be performed to assess the arterial anatomy, the extent of liver involvement, portal vein patency, and bile duct dilatation. Thoracoabdominal-pelvic CT is indicated for use when looking for extrahepatic metastases from NETs. Ideal candidates have a nonpancreatic NET with a resected primary tumor site, no extrahepatic metastases, and between a 30% and 50% liver involvement. However, some benefits may be observed in other patients, and all patients should be discussed individually. SIRT consists of the transarterial deposition of a radioactive source within tumors [89], based on the predominant arterial vascularization in NELMs [195], and has shown promising results with good tolerance compared to TAE and TACE. Indeed, several studies reported that a complete response occurred in 1–8% of patients, which was not found in TAE/TACE [196–200].

RECIST 1.1 or mRECIST criteria were performed in radiological follow-ups [197,201] after SIRT, and treated lesions appeared hypovascular or necrotic in 97% of patients [202]. Some meta-analyses have shown an objective radiological response rate with a weighted mean of 51% (95% CI: 47–54%) [196,203] and a mean disease control rate of 86–88% [196,204]. Symptom improvement has been reported in 20–80% of SIRT-treated NELM patients and decreased lesion diameters, low enhancement, and a high proportion of necrosis post-SIRT compared to pre-SIRT MRI are prognostic factors of survival after SIRT [205,206]. A decrease in lesion diameters, low enhancement, and a high portion of necrosis on post-SIRT MRI compared to pre-SIRT MRI are prognostic survival factors after SIRT [205,206]. In molecular imaging, a decrease in the standardized uptake value (SUV) ratio of the liver-spleen uptake using ⁶⁸Ga-DOTATOC PET between pre- and post-SIRT exams has been associated with improved overall survival and progression-free survival [207]. Several retrospective studies suggest that the resection of metastatic lesions improves the 5-year survival rate from 30–40 to 75% [208,209] with a radical resection (R0) of NELMs correlating with a 5-year overall survival (OS) rate of approximately 85% [210]. However, NELMs are frequently more extensive than those identified on presurgical imaging and intraoperatively,

and an accurate curative resection is difficult to achieve. Several pieces of research showed as a multimodality approach with surgical and ablative treatments, allowing for better disease control. So, it is clear the necessity to choose the imaging tool that allows the identification of all lesions so as the relationship with the vascular and biliary duct for proper patient management. Although MRI with liver-specific contrast agents allows to identify very small lesions, this tool should be combined with CT to obtain an optimal vascular evaluation [211–218] (Figure 5).

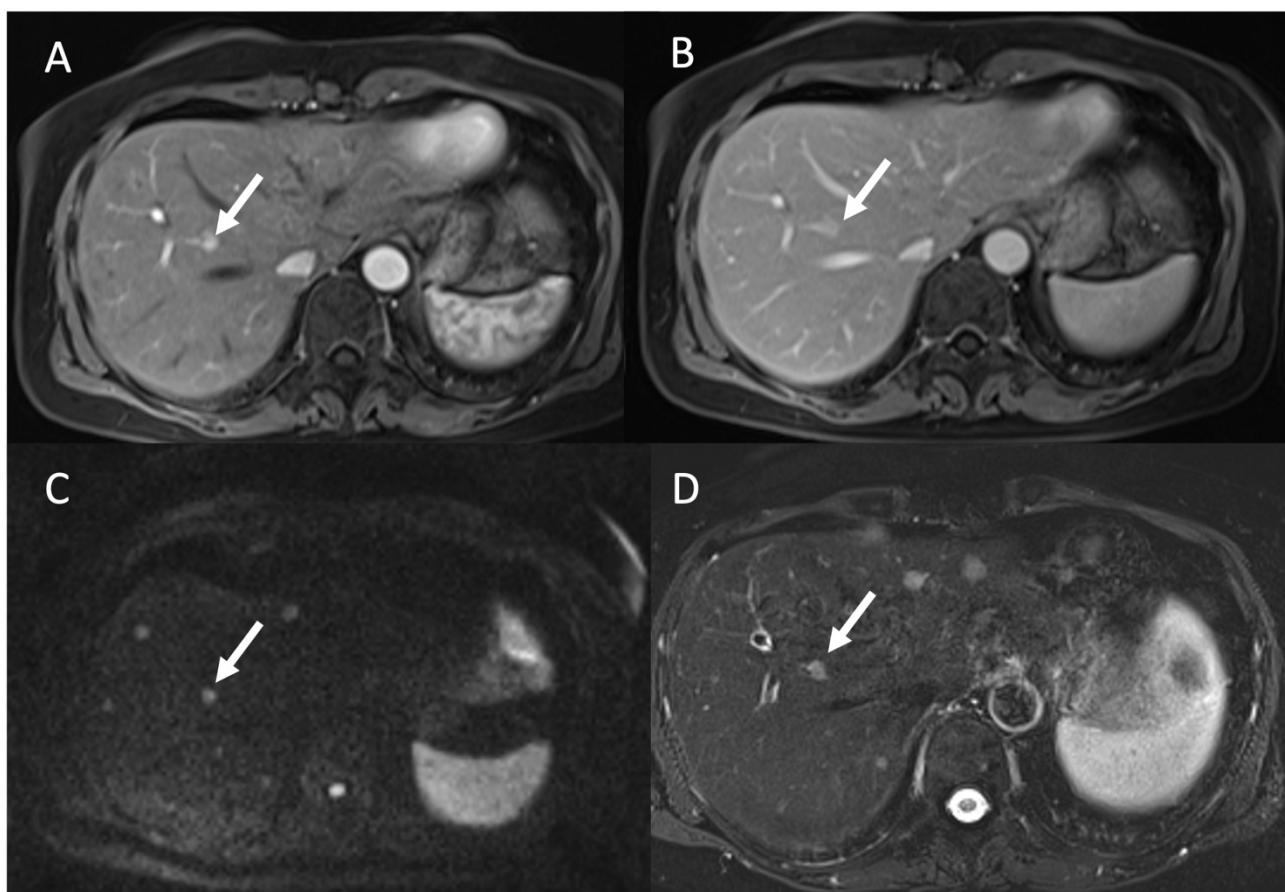


Figure 5. MRI evaluation of liver NEN metastases. Contrast study ((A): arterial; (B): portal phase) show a single lesion (arrow), while, in DWI (C) and T2-W FS (D), more lesions were detected compared to (A,B).

3.3. Gastrointestinal Tract NEN

In the gastrointestinal tract, neuroendocrine neoplasms are found most frequently in the small bowel (26%) and rectum (34%), less frequently in the appendix (6%), cecum (6%), stomach (12%), and duodenum (8%), with the colon being a rare site of onset [219–223]. Most gastroenteropancreatic neuroendocrine neoplasms are small (<2 cm), and, therefore, conventional imaging techniques fail to define primary cancer in patients with metastatic diseases [7]. For these reasons, other small bowel imaging techniques, such as CT and MRI enterography, or enteroclysis and endoscopic techniques, such as capsule endoscopy and double-balloon enteroscopy, may improve the detection of primary occult tumors. Furthermore, these neoplasms stimulate a fibrotic reaction in the surrounding tissue, which can lead to functional obstruction or vascular compromise [224]. Presently, the use of entero-MRI for diagnosing small intestinal neuroendocrine tumors has not yet been established, considering the high cost of this investigation. The significant advantage of entero-CT compared to conventional CT is the ability to identify lesions of 1–2 cm thanks to the high difference in contrast between neoplasm and intestinal lumen obtained through

the oral administration of the hyperosmolar contrast medium, thus, being able to identify neuroendocrine tumors in a small size also [7]. At the duodenum level, they appear as small intramural masses, polypoid lesions that protrude into the lumen, or circumferential thickenings of the wall. In contrast, when they arise in the small bowel, they can appear as solitary or multiple polypoid lesions presenting an intense enhancement after the intravenous administration of an iodinated contrast medium, or as “plaque” wall thickening [225]. Colorectal GEP-NENs are associated with a worse prognosis when compared with neuroendocrine tumors occurring elsewhere, as they are often advanced at the time of diagnosis. At the colic level, neuroendocrine tumors tend to be larger (>2 cm) and often involve the cecum and ascending colon. Approximately 34% of gastrointestinal neuroendocrine tumors are located in the rectum and often present as small solitary submucosal masses, as multiple nodules, or as single ulcerated polypoid lesions. A virtual colonoscopy cannot differentiate a neuroendocrine tumor from the more frequent adenocarcinoma, since both can appear as circumferential thickening of the wall or as polypoid formations protruding into the intestinal lumen, which are associated, in most cases, with locoregional lymphadenopathies [226] (Figure 6).

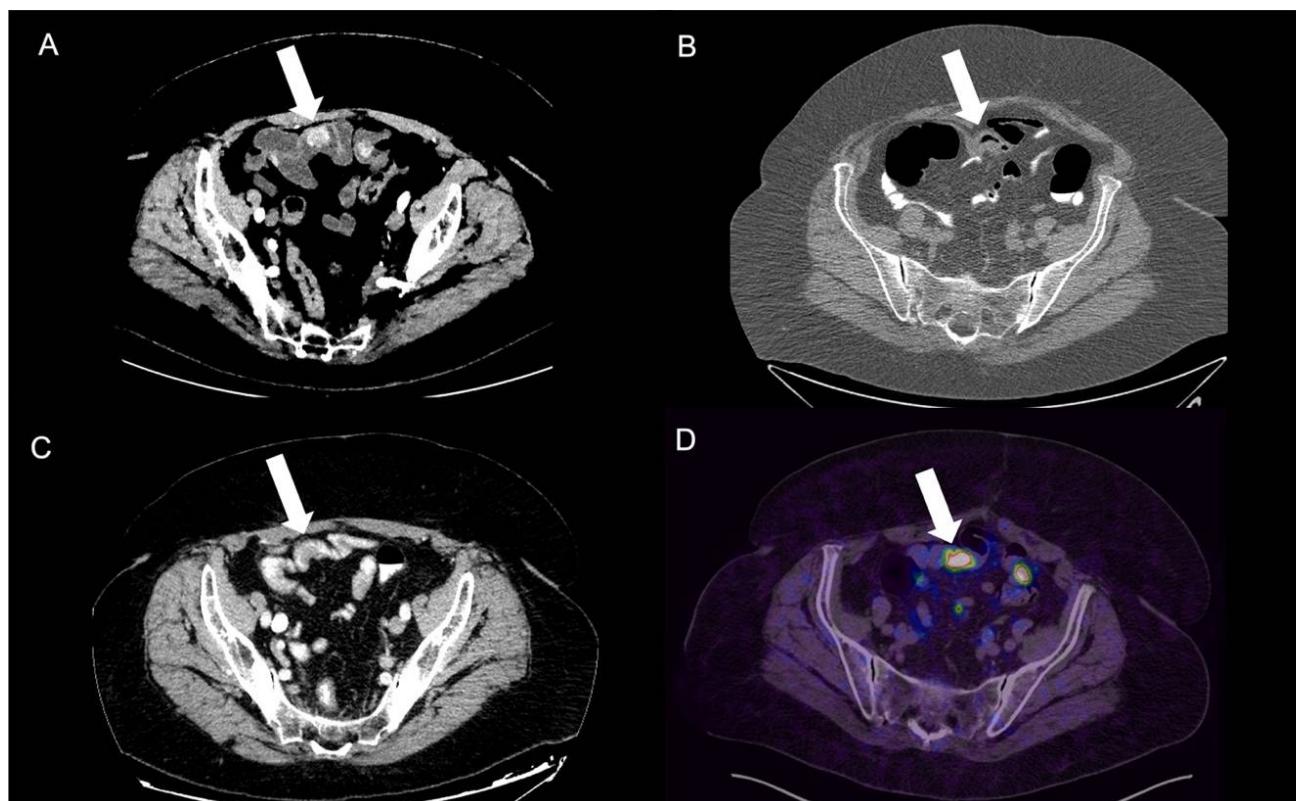


Figure 6. Ileal NEN tumors shown with different endoluminal contrasts and in PET-CT exam. The hypervasculatized tumors are clearly different from normal bowel walls due to being highlighted in relation to endoluminal hypodense and hyperosmolar contrast media. The same patients previously had a colon CT performed, showing a slight air distension of lumen with minimal wall thickening that did not improve the tumor evidence, as well as a later contrast-enhanced CT with oral hyperosmolar iodinated contrast, which improved the intestinal lumen visualization but strongly limited the enhancement of tumors. PET/CT-⁶⁸Gallium showed the same highly uptaking lesions, with the highest sensitivity compared to others, but this last one is not routinely performed without neuroendocrine suspicion being specific for highly probable or known NEN tumors. (A) CT-enterography, (B) virtual colonoscopy, (C) CECT with oral contrast, and (D) PET-⁶⁸Ga.

4. Follow-Up and Treatment Assessment

There is no consensus on the optimal follow-up for completely resected gastroenteropancreatic neuroendocrine tumors. Published guidelines for follow-up are complex and emphasize closer surveillance in the first 2 years after resection [100,227–229]. The recommendations are summarized in Table 5.

Table 5. Follow-up recommendations after NET resection in current guidelines.

	ESMO	NCCN	ENETS
Small-bowel NEN	Grades 1–2: Laboratory tests and CT or MRI every 3–6 months. Grade 3: Every 2–3 months. Octreoscan after 18–24 months if SRS positive.	Clinical review at 3–12 months with biomarkers and CT or MRI as clinically indicated; then, review every 6–12 months for maximum of 10 years.	Grade 1: US, CT, or MRI at 6 and 12 months, then yearly or longer; octreoscan (or gallium-68-based PET) at baseline and every 2 years. Grade 2–3: US, CT, or MRI every 3 months indefinitely; octreoscan (or gallium-68-based PET) at 3 months and yearly.
Pancreatic NEN	Grades 1–2: Laboratory tests and CT or MRI every 3–6 months. Grade 3: Every 2–3 months. Octreoscan after 18–24 months if SSTR positive.	Clinical review at 3–12 months with biomarkers and CT or MRI as clinically indicated; then, review every 6–12 months for maximum of 10 years.	Grade 1: US, CT, or MRI at 6 and 12 months, then yearly or longer; octreoscan (or gallium-68-based PET) at baseline and every 2 years. Grades 2–3: US, CT, or MRI every 3 months indefinitely; octreoscan (or gallium-68-based PET) at 3 months and yearly.
Appendiceal NEN	Not reported	Not follow-up: <2 cm completely resected by appendicectomy “as clinically indicated”.	No follow-up: <1 cm completely resected with appendicectomy; appendiceal NET >1 cm completely resected with right hemicolectomy without lymph node involvement.
Rectal NEN	Not reported	Not follow-up: <1 cm with negative margins.	No follow-up: completely resected rectal NETs <1 cm.

ENETS, European Neuroendocrine Tumor Society; ESMO, European Society for Medical Oncology; NCCN, National Comprehensive Cancer Network; NET, neuroendocrine tumor; CT, computed tomography; MRI, magnetic resonance imaging; PET, positron emission tomography; SRS, somatostatin receptor scintigraphy; US, ultrasonography.

However, NENs have a different pattern and timescale of recurrence and, thus, require a more practical and tailored follow-up, including improvements in cost, patient flow, and the elimination of excessive demand for inefficient tests [230]. CT and MRI should be preferred for G1 and G2, while the follow-up of G3 tumors should also include gallium-68-based PET. Regardless, the follow-up for NENs requires a multidisciplinary approach, including biochemical (chromogranin A, hormones, and vasoactive amines), radiologic, and histologic investigations.

Regarding treatment assessments, medical therapies in NEN patients vary from somatostatin analogs, targeted agents, such as everolimus and sunitinib, and PRRT to ablation treatments. In this scenario, the limits of Recist 1.1 are clear, which evaluates the tumor shrinkage and the necessity to use functional criteria.

5. Conclusions

The radiological approach to GEP-NENs is variable and depends on symptoms, laboratory conditions, and clinical suspicions. In the absence of these, the sensitivity and specificity of instrumental examinations are limited, above all, due to the difficulty in finding primitive lesions, which are often small in size. When symptoms are subtle due to the absence or poor secretion of hormones or neuropeptides, diagnosis is delayed and the prognosis worsens.

The choice of accurate imaging of NETs is fundamental for the diagnosis, staging, and assessment of suitability for surgery, choice of therapy, and response to treatment. The radiologist should be aware of each method's strengths and limitations and their complementarity to recommend the best test for the given clinical scenario. Whichever test is chosen, attention to detail in acquiring and interpreting the study is critical to achieving the best possible results.

Author Contributions: Conceptualization, F.P. and V.G.; methodology, M.S.; validation, R.F. and F.G.; formal analysis, V.G.; investigation, L.P., G.T. and S.T.; writing—original draft preparation, F.P., V.G., L.P., G.T., R.F., S.T. and F.G.; writing—review and editing, F.P. and V.G.; supervision, M.S. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Conflicts of Interest: The authors declare no conflict of interest.

References

- Yao, J.; Hassan, M.; Phan, A.; Dagohoy, C.; Leary, C.; Mares, J.; Abdalla, E.; Fleming, J.; Vauthey, J.-N.; Rashid, A.; et al. One hundred years after “carcinoid”: Epidemiology of and prognostic factors for neuroendocrine tumors in 35,825 cases in the United States. *J. Clin. Oncol.* **2008**, *26*, 3063–3072. [[CrossRef](#)] [[PubMed](#)]
- Eh, T.; Ch, T. Imaging of gastroenteropancreatic neuroendocrine tumors. *World J. Clin. Oncol.* **2011**, *2*, 28. [[CrossRef](#)]
- Pantelis, A.; Panagopoulou, P.; Lapatsanis, D. Artificial Intelligence and Machine Learning in the Diagnosis and Management of Gastroenteropancreatic Neuroendocrine Neoplasms—A Scoping Review. *Diagnostics* **2022**, *12*, 874. [[CrossRef](#)] [[PubMed](#)]
- Chiti, G.; Grazzini, G.; Flammia, F.; Matteuzzi, B.; Tortoli, P.; Bettarini, S.; Pasqualini, E.; Granata, V.; Busoni, S.; Messserini, L.; et al. Gastroenteropancreatic neuroendocrine neoplasms (GEP-NENs): A radiomic model to predict tumor grade. *Radiol. Med.* **2022**, *127*, 1234. [[CrossRef](#)]
- Benedetti, G.; Mori, M.; Panzeri, M.; Barbera, M.; Palumbo, D.; Sini, C.; Muffatti, F.; Andreasi, V.; Steidler, S.; Doglioni, C.; et al. CT-derived radiomic features to discriminate histologic characteristics of pancreatic neuroendocrine tumors. *Radiol. Med.* **2021**, *126*, 745–760. [[CrossRef](#)]

6. Sahani, D.; Bonaffini, P.; Castillo, C.F.-D.; Blake, M. Gastroenteropancreatic Neuroendocrine Tumors: Role of Imaging in Diagnosis and Management. *Radiology* **2013**, *11*, 38–61. [[CrossRef](#)]
7. Ganeshan, D.; Bhosale, P.; Yang, T.; Kundras, V. Imaging Features of Carcinoid Tumors of the Gastrointestinal Tract. *AJR Am. J. Roentgenol.* **2013**, *201*, 773–786. [[CrossRef](#)]
8. Granata, V.; Fusco, R.; Setola, S.V.; de Lutio di Castelguidone, E.; Camera, L.; Tafuto, S.; Avallone, A.; Belli, A.; Incollingo, P.; Palaia, R.; et al. The multidisciplinary team for gastroenteropancreatic neuroendocrine tumours: The radiologist's challenge. *Radiol. Oncol.* **2019**, *53*, 373–387. [[CrossRef](#)] [[PubMed](#)]
9. Danti, G.; Flammia, F.; Matteuzzi, B.; Cozzi, D.; Berti, V.; Grazzini, G.; Pradella, S.; Recchia, L.; Brunese, L.; Miele, V. Gastrointestinal neuroendocrine neoplasms (GI-NENs): Hot topics in morphological, functional, and prognostic imaging. *Radiol. Med.* **2021**, *126*, 1497–1507. [[CrossRef](#)]
10. De Felice, F.; Boldrini, L.; Greco, C.; Nardone, V.; Salvestrini, V.; Desideri, I. ESTRO vision 2030: The young Italian Association of Radiotherapy and Clinical Oncology (yAIRO) commitment statement. *Radiol. Med.* **2021**, *126*, 1374–1376. [[CrossRef](#)]
11. Granata, V.; Faggioni, L.; Grassi, R.; Fusco, R.; Reginelli, A.; Rega, D.; Maggioretti, N.; Buccicardi, D.; Frittoli, B.; Rengo, M.; et al. Structured reporting of computed tomography in the staging of colon cancer: A Delphi consensus proposal. *Radiol. Med.* **2022**, *127*, 21–29. [[CrossRef](#)] [[PubMed](#)]
12. Wang, R.; Zheng-Powell, R.; Chen, H.; Bibb, J.; Chen, H.; Rose, J. Management of Gastrointestinal Neuroendocrine Tumors. *Clin. Med. Insights Endocrinol. Diabetes* **2019**, *12*, 1179551419884058. [[CrossRef](#)]
13. Liang, W.; Yang, P.; Huang, R.; Xu, L.; Wang, J.; Liu, W.; Zhang, L.; Wan, D.; Huang, Q.; Lu, Y.; et al. A Combined Nomogram Model to Preoperatively Predict Histologic Grade in Pancreatic Neuroendocrine Tumors. *Clin. Cancer Res.* **2019**, *25*, 584–594. [[CrossRef](#)] [[PubMed](#)]
14. Saleh, M.; Bhosale, P.; Yano, M.; Itani, M.; Elsayes, A.; Halperin, D.; Bergsland, E.; Morani, A. New frontiers in imaging including radiomics updates for pancreatic neuroendocrine neoplasms. *Abdom. Radiol.* **2020**, *47*, 3078–3100. [[CrossRef](#)]
15. Faviana, P.; Boldrini, L.; Gentile, C.; Erba, P.; Sammarco, E.; Bartoli, F.; Esposito, E.; Galli, L.; Lippolis, P.; Bardi, M. Proposal for a New Diagnostic Histopathological Approach in the Evaluation of Ki-67 in GEP-NETs. *Diagnostics* **2022**, *12*, 1960. [[CrossRef](#)]
16. Coriat, R.; Walter, T.; Terris, B.; Couvelard, A.; Ruszniewski, P. Gastroenteropancreatic Well-Differentiated Grade 3 Neuroendocrine Tumors: Review and Position Statement. *Oncologist* **2016**, *21*, 1191. [[CrossRef](#)] [[PubMed](#)]
17. Heetfeld, M.; Chougnat, C.; Olsen, I.; Rinke, A.; Borbath, I.; Crespo, G.; Barriuso, J.; Pavel, M.; O'Toole, D.; Walter, T. Characteristics and treatment of patients with G3 gastroenteropancreatic neuroendocrine neoplasms. *Endocr. Relat. Cancer* **2015**, *22*, 657–664. [[CrossRef](#)]
18. Caruso, D.; Polici, M.; Rinzivillo, M.; Zerunian, M.; Nacci, I.; Marasco, M.; Magi, L.; Tarallo, M.; Gargiulo, S.; Iannicelli, E.; et al. CT-based radiomics for prediction of therapeutic response to Everolimus in metastatic neuroendocrine tumors. *Radiol. Med.* **2022**, *127*, 691–701. [[CrossRef](#)] [[PubMed](#)]
19. Karmazanovsky, G.; Gruzdev, I.; Tikhonova, V.; Kondratyev, E.; Revishvili, A. Computed tomography-based radiomics approach in pancreatic tumors characterization. *Radiol. Med.* **2021**, *126*, 1388–1395. [[CrossRef](#)]
20. Cozzi, D.; Bicci, E.; Cavigli, E.; Danti, G.; Bettarini, S.; Tortoli, P.; Lorenzo; Mazzoni, N.; Busoni, S.; Pradella, S.; et al. Chest Radiology Radiomics in pulmonary neuroendocrine tumours (NETs). *Radiol. Med.* **2022**, *127*, 609–615. [[CrossRef](#)] [[PubMed](#)]
21. Karlafti, E.; Charalampidou, M.; Fotiadou, G.; Deka, I.A.; Raptou, G.; Kyriakidis, F.; Panidis, S.; Ioannidis, A.; Protopapas, A.; Netta, S.; et al. Ampullary Large-Cell Neuroendocrine Carcinoma, a Diagnostic Challenge of a Rare Aggressive Neoplasm: A Case Report and Literature Review. *Diagnostics* **2022**, *12*, 1797. [[CrossRef](#)] [[PubMed](#)]
22. Mitrovic-Jovanovic, M.; Grubor, N.; Milosevic, S.; Jankovic, A.; Stosic, K.; Ostojic, S.; Ninic, A.; Micev, M.; Kovac, J. Total Pancreatectomy for Multicentric Cystic Neuroendocrine Tumor of the Pancreas: A Case Report. *Diagnostics* **2022**, *12*, 1003. [[CrossRef](#)]
23. Yang, Z.; Tang, L.; Klimstra, D. Effect of tumor heterogeneity on the assessment of Ki67 labeling index in well-differentiated neuroendocrine tumors metastatic to the liver: Implications for prognostic stratification. *Am. J. Surg. Pathol.* **2011**, *35*, 853–860. [[CrossRef](#)] [[PubMed](#)]
24. Oronsky, B.; Ma, P.; Morgensztern, D.; Carter, C. Nothing But NET: A Review of Neuroendocrine Tumors and Carcinomas. *Neoplasia* **2017**, *19*, 991–1002. [[CrossRef](#)]
25. Sundin, A.; Arnold, R.; Baudin, E.; Cwikla, J.; Eriksson, B.; Fanti, S.; Fazio, N.; Giammarile, F.; Hicks, R.; Kjaer, A.; et al. ENETS Consensus Guidelines for the Standards of Care in Neuroendocrine Tumors: Radiological, Nuclear Medicine and Hybrid Imaging. *Neuroendocrinology* **2017**, *105*, 212–244. [[CrossRef](#)]
26. El Ghannudi, S.; Ouvrard, E.; Mikail, N.; Freschini, B.; Schindler, T.; Imperiale, A. Cutting-Edge Imaging of Cardiac Metastases from Neuroendocrine Tumors: Lesson from a Case Series. *Diagnostics* **2022**, *12*, 1182. [[CrossRef](#)]
27. Zilli, A.; Arcidiacono, P.; Conte, D.; Massironi, S. Clinical impact of endoscopic ultrasonography on the management of neuroendocrine tumors: Lights and shadows. *Dig. Liver Dis.* **2018**, *50*, 6–14. [[CrossRef](#)]

28. Vicini, S.; Bortolotto, C.; Rengo, M.; Ballerini, D.; Bellini, D.; Carbone, I.; Preda, L.; Laghi, A.; Coppola, F.; Faggioni, L. A narrative review on current imaging applications of artificial intelligence and radiomics in oncology: Focus on the three most common cancers. *Radiol. Med.* **2022**, *127*, 819–836. [\[CrossRef\]](#) [\[PubMed\]](#)
29. Baumann, T.; Rottenburger, C.; Nicolas, G.; Wild, D. Gastroenteropancreatic neuroendocrine tumours (GEP-NET)—Imaging and staging. *Best Pract. Res. Clin. Endocrinol. Metab.* **2016**, *30*, 45–57. [\[CrossRef\]](#)
30. Ramage, J.; Davies, A.H.G.; Ardill, J.; Bax, N.; Caplin, M.; Grossman, A.; Hawkins, R.; McNicol, A.; Reed, N.; Sutton, R. Watkinson, Guidelines for the management of gastroenteropancreatic neuroendocrine (including carcinoid) tumours. *Gut* **2005**, *54*, iv1–iv16. [\[CrossRef\]](#)
31. Luo, Y.; Pan, Q.; Yao, S.; Yu, M.; Wu, W.; Xue, H.; Kiesewetter, D.; Zhu, Z.; Li, F.; Zhao, Y.; et al. Glucagon-Like Peptide-1 Receptor PET/CT with 68Ga-NOTA-Exendin-4 for Detecting Localized Insulinoma: A Prospective Cohort Study. *J. Nucl. Med.* **2016**, *57*, 715. [\[CrossRef\]](#) [\[PubMed\]](#)
32. Frilling, A.; Sotiropoulos, G.; Radtke, A.; Malago, M.; Bockisch, A.; Kuehl, H.; Li, J.; Broelsch, C. The impact of 68Ga-DOTATOC positron emission tomography/computed tomography on the multimodal management of patients with neuroendocrine tumors. *Ann. Surg.* **2010**, *252*, 850–855. [\[CrossRef\]](#) [\[PubMed\]](#)
33. Olsen, I.; Langer, S.; Federspiel, B.; Oxbøl, J.; Loft, A.; Berthelsen, A.K.; Mortensen, J.; Oturai, P.; Knigge, U.; Kjaer, A. 68Ga-DOTATOC PET and gene expression profile in patients with neuroendocrine carcinomas: Strong correlation between PET tracer uptake and gene expression of somatostatin receptor subtype 2. *Am. J. Nucl. Med. Mol. Imag.* **2016**, *6*, 59.
34. Binderup, T.; Knigge, U.; Loft, A.; Mortensen, J.; Pfeifer, A.; Federspiel, B.; Hansen, C.; Højgaard, L.; Kjaer, A. Functional imaging of neuroendocrine tumors: A head-to-head comparison of somatostatin receptor scintigraphy, 123I-MIBG scintigraphy, and 18F-FDG PET. *J. Nucl. Med.* **2010**, *51*, 704–712. [\[CrossRef\]](#)
35. Simsek, D.; Kuyumcu, S.; Turkmen, C.; Sanli, Y.; Aykan, F.; Unal, S.; Adalet, I. Can Complementary 68Ga-DOTATATE and 18F-FDG PET/CT Establish the Missing Link Between Histopathology and Therapeutic Approach in Gastroenteropancreatic Neuroendocrine Tumors? *J. Nucl. Med.* **2014**, *55*, 1811–1817. [\[CrossRef\]](#)
36. Maxwell, J.; Howe, J. Imaging in neuroendocrine tumors: An update for the clinician. *Int. J. Endocr. Oncol.* **2015**, *2*, 159–168. [\[CrossRef\]](#)
37. Kaewput, C.; Vinjamuri, S. Role of Combined 68Ga DOTA-Peptides and 18F FDG PET/CT in the Evaluation of Gastroenteropancreatic Neuroendocrine Neoplasms. *Diagnostics* **2022**, *12*, 280. [\[CrossRef\]](#)
38. Shetty, D.; Patel, D.; Le, K.; Bui, C.; Mansberg, R. Pitfalls in Gallium-68 PSMA PET/CT Interpretation—A Pictorial Review. *Tomography* **2018**, *4*, 182–193. [\[CrossRef\]](#)
39. de Herder, W.; Kwekkeboom, D.; Feelders, R.; van Aken, M.; Lamberts, S.W.J.; van der Lely, A.; Krenning, E. Somatostatin receptor imaging for neuroendocrine tumors. *Pituitary* **2006**, *9*, 243–248. [\[CrossRef\]](#)
40. Ambrosini, V.; Campana, D.; Tomassetti, P.; Grassetto, G.; Rubello, D.; Fanti, S. PET/CT with 68Gallium-DOTA-peptides in NET: An overview. *Eur. J. Radiol.* **2011**, *80*, e116–e119. [\[CrossRef\]](#)
41. Bauckneht, M.; Albano, D.; Annunziata, S.; Santo, G.; Guglielmo, P.; Frantellizzi, V.; Branca, A.; Ferrari, C.; Vento, A.; Mirabile, A.; et al. Somatostatin Receptor PET/CT Imaging for the Detection and Staging of Pancreatic NET: A Systematic Review and Meta-Analysis. *Diagnostics* **2020**, *10*, 598. [\[CrossRef\]](#)
42. Hussein, M.; Cafarelli, F.; Paparella, M.; Rennie, W.; Guglielmi, G. Phosphaturic mesenchymal tumors: Radiological aspects and suggested imaging pathway. *Radiol. Med.* **2021**, *126*, 1609–1618. [\[CrossRef\]](#) [\[PubMed\]](#)
43. Ambrosini, V.; Campana, D.; Tomassetti, P.; Fanti, S. 68Ga-labelled peptides for diagnosis of gastroenteropancreatic NET. *Eur. J. Nucl. Med.* **2012**, *39*, 52–60. [\[CrossRef\]](#)
44. Poeppel, T.; Binse, I.; Petersenn, S.; Lahner, H.; Schott, M.; Antoch, G.; Brandau, W.; Bockisch, A.; Boy, C. 68Ga-DOTATOC versus 68Ga-DOTATATE PET/CT in functional imaging of neuroendocrine tumors. *J. Nucl. Med.* **2011**, *52*, 1864–1870. [\[CrossRef\]](#) [\[PubMed\]](#)
45. Gabriel, M.; Decristoforo, C.; Kendler, D.; Dobrozemsky, G.; Heute, D.; Uprimny, C.; Kovacs, P.; Von Guggenberg, E.; Bale, R.; Virgolini, I. 68Ga-DOTA-Tyr3-octreotide PET in neuroendocrine tumors: Comparison with somatostatin receptor scintigraphy and CT. *J. Nucl. Med.* **2007**, *48*, 508–518. [\[CrossRef\]](#) [\[PubMed\]](#)
46. Buchmann, I.; Henze, M.; Engelbrecht, S.; Eisenhut, M.; Runz, A.; Schäfer, M.; Schilling, T.; Haufe, S.; Herrmann, T.; Haberkorn, U. Comparison of 68Ga-DOTATOC PET and 111In-DTPAOC (Octreoscan) SPECT in patients with neuroendocrine tumours. *Eur. J. Nucl. Med. Mol.* **2007**, *34*, 1617–1626. [\[CrossRef\]](#)
47. Srirajaskanthan, R.; Kayani, I.; Quigley, A.; Soh, J.; Caplin, M.; Bomanji, J. The role of 68Ga-DOTATATE PET in patients with neuroendocrine tumors and negative or equivocal findings on 111In-DTPA-octreotide scintigraphy. *J. Nucl. Med.* **2010**, *51*, 875–882. [\[CrossRef\]](#)
48. Naswa, N.; Sharma, P.; Kumar, A.; Nazar, A.; Kumar, R.; Chumber, S.; Bal, C. Gallium-68-DOTA-NOC PET/CT of patients with gastroenteropancreatic neuroendocrine tumors: A prospective single-center study. *AJR. Am. J. Roentgenol.* **2011**, *197*, 1221–1228. [\[CrossRef\]](#)

49. Krausz, Y.; Freedman, N.; Rubinstein, R.; Lavie, E.; Orevi, M.; Tshori, S.; Salmon, A.; Glaser, B.; Chisin, R.; Mishani, E.; et al. 68Ga-DOTA-NOC PET/CT imaging of neuroendocrine tumors: Comparison with 111In-DTPA-octreotide (OctreoScan®). *Mol. Imaging Biol.* **2011**, *13*, 583–593. [CrossRef] [PubMed]
50. Ambrosini, V.; Tomassetti, P.; Castellucci, P.; Campana, D.; Montini, G.; Rubello, D.; Nanni, C.; Rizzello, A.; Franchi, R.; Fanti, S. Comparison between 68Ga-DOTA-NOC and 18F-DOPA PET for the detection of gastro-entero-pancreatic and lung neuro-endocrine tumours. *Eur. J. Nucl. Med. Mol.* **2008**, *35*, 1431–1438. [CrossRef]
51. Calabro, D.; Argalia, G.; Ambrosini, V. Role of PET/CT and Therapy Management of Pancreatic Neuroendocrine Tumors. *Diagnostics* **2020**, *10*, 1059. [CrossRef]
52. Sharma, P.; Arora, S.; Mukherjee, A.; Pal, S.; Sahni, P.; Garg, P.; Khadgawat, R.; Thulkar, S.; Bal, C.; Kumar, R. Predictive value of 68Ga-DOTANOC PET/CT in patients with suspicion of neuroendocrine tumors: Is its routine use justified? *Clin. Nucl. Med.* **2014**, *39*, 37–43. [CrossRef]
53. Sharma, P.; Naswa, N.; KC, S.; Alvarado, L.; Dwivedi, A.; Yadav, Y.; Kumar, R.; Ammini, A.; Bal, C. Comparison of the prognostic values of 68Ga-DOTANOC PET/CT and 18F-FDG PET/CT in patients with well-differentiated neuroendocrine tumor. *Eur. J. Nucl. Med. Mol.* **2014**, *41*, 2194–2202. [CrossRef] [PubMed]
54. Sharma, P.; Arora, S.; Dhull, V.; Naswa, N.; Kumar, R.; Ammini, A.; Bal, C. Evaluation of (68)Ga-DOTANOC PET/CT imaging in a large exclusive population of pancreatic neuroendocrine tumors. *Abdom. Imag.* **2015**, *40*, 299–309. [CrossRef] [PubMed]
55. Rufini, V.; Baum, R.; Castaldi, P.; Treglia, G.; De Gaetano, A.; Carreras, C.; Kaemmerer, D.; Hommann, M.; Hörsch, D.; Bonomo, L.; et al. Role of PET/CT in the functional imaging of endocrine pancreatic tumors. *Abdom. Imag.* **2012**, *37*, 1004–1020. [CrossRef]
56. Treglia, G.; Castaldi, P.; Rindi, G.; Giordano, A.; Rufini, V. Diagnostic performance of Gallium-68 somatostatin receptor PET and PET/CT in patients with thoracic and gastroenteropancreatic neuroendocrine tumours: A meta-analysis. *Endocrine* **2012**, *42*, 80–87. [CrossRef] [PubMed]
57. Wild, D.; Bomanji, J.; Benkert, P.; Maecke, H.; Ell, P.; Reubi, J.; Caplin, M. Comparison of 68Ga-DOTANOC and 68Ga-DOTATATE PET/CT Within Patients with Gastroenteropancreatic Neuroendocrine Tumors. *J. Nucl. Med.* **2013**, *54*, 364–372. [CrossRef]
58. De Camargo Etchebehere, E.C.S.; De Oliveira Santos, A.; Gumz, B.; Vicente, A.; Hoff, P.; Corradi, G.; Ichiki, W.; De Almeida Filho, J.; Cantoni, S.; Camargo, E.; et al. 68Ga-DOTATATE PET/CT, 99mTc-HYNIC-Octreotide SPECT/CT, and Whole-Body MR Imaging in Detection of Neuroendocrine Tumors: A Prospective Trial. *J. Nucl. Med.* **2014**, *55*, 1598–1604. [CrossRef]
59. Durgapal, P.; Sharma, R.; Kandasamy, D.; Bal, C. Somatostatin receptor based PET/CT imaging with 68Ga-DOTA-Nal3-octreotide for localization of clinically and biochemically suspected insulinoma. *Q. J. Nucl. Med. Mol. Imag.* **2016**, *60*, 69–76.
60. Falconi, M.; Eriksson, B.; Kaltsas, G.; Bartsch, D.; Capdevila, J.; Caplin, M.; Kos-Kudla, B.; Kwekkeboom, D.; Rindi, G.; Klöppel, G.; et al. ENETS Consensus Guidelines Update for the Management of Patients with Functional Pancreatic Neuroendocrine Tumors and Non-Functional Pancreatic Neuroendocrine Tumors. *Neuroendocrinology* **2016**, *103*, 153–171. [CrossRef]
61. Bozkurt, M.; Virgolini, I.; Balogova, S.; Beheshti, M.; Rubello, D.; Decristoforo, C.; Ambrosini, V.; Kjaer, A.; Delgado-Bolton, R.; Kunikowska, J.; et al. Guideline for PET/CT imaging of neuroendocrine neoplasms with 68Ga-DOTA-conjugated somatostatin receptor targeting peptides and 18F-DOPA. *Eur. J. Nucl. Med. Mol. Imag.* **2017**, *44*, 1588–1601. [CrossRef]
62. Severi, S.; Nanni, O.; Bodei, L.; Sansovini, M.; Ianniello, A.; Nicoletti, S.; Scarpi, E.; Matteucci, F.; Gilardi, L.; Paganelli, G. Role of 18FDG PET/CT in patients treated with 177Lu-DOTATATE for advanced differentiated neuroendocrine tumours. *Eur. J. Nucl. Med. Mol. Imag.* **2013**, *40*, 881–888. [CrossRef]
63. Delpassand, E.; Samarghandi, A.; Mourtada, J.; Zamanian, S.; Espenan, G.; Sharif, R.; MacKenzie, S.; Kosari, K.; Barakat, O.; Naqvi, S.; et al. Long-Term Survival, Toxicity Profile, and role of F-18 FDG PET/CT scan in Patients with Progressive Neuroendocrine Tumors Following Peptide Receptor Radionuclide Therapy with High Activity In-111 Pentetreotide. *Theranostics* **2012**, *2*, 472–480. [CrossRef] [PubMed]
64. Orlefors, H.; Sundin, A.; Garske, U.; Juhlin, C.; Oberg, K.; Skogseid, B.; Langstrom, B.; Bergstrom, M.; Eriksson, B. Whole-body (11)C-5-hydroxytryptophan positron emission tomography as a universal imaging technique for neuroendocrine tumors: Comparison with somatostatin receptor scintigraphy and computed tomography. *J. Clin. Endocrinol. Metab.* **2005**, *90*, 3392–3400. [CrossRef] [PubMed]
65. Marchetti, L.; Perrucci, L.; Pellegrino, F.; Baroni, L.; Merlo, A.; Tilli, M.; Rambaldi, I.; Maietti, E.; Carnevale, A.; Bartolomei, M.; et al. Diagnostic contribution of contrast-enhanced CT as compared to unenhanced low-dose CT in PET/CT staging and treatment response assessment of 18FDG-avid lymphomas: A prospective study. *J. Nucl. Med.* **2021**, *62*, 1372–1379. [CrossRef]
66. Pellegrino, F.; Scabbia, F.; Merlo, A.; Perrucci, L.; Aliberti, L.; Urso, A.; Ambrosio, M.; Cuneo, A.; Galeotti, R.; Giganti, M. Spontaneously reversible adrenal nodules in primary diffuse large B-cell testicular lymphoma mimicking an extranodal involvement: A case report. *Radiol. Case Rep.* **2021**, *16*, 2168–2173. [CrossRef]
67. Koopmans, K.; Neels, O.; Kema, I.; Elsinga, P.; Sluiter, W.; Vanghillewe, K.; Brouwers, A.; Jager, P.; De Vries, E. Improved staging of patients with carcinoid and islet cell tumors with 18F-dihydroxy-phenylalanine and 11C-5-hydroxy-tryptophan positron emission tomography. *J. Clin. Oncol.* **2008**, *26*, 1489–1495. [CrossRef]
68. Adams, S.; Baum, R.; Rink, T.; Schumm-Dräger, P.; Usadel, K.; Hör, G. Limited value of fluorine-18 fluorodeoxyglucose positron emission tomography for the imaging of neuroendocrine tumours. *Eur. J. Nucl. Med.* **1998**, *25*, 79–83. [CrossRef]

69. Mapelli, P.; Partelli, S.; Salgarello, M.; Doraku, J.; Muffatti, F.; Lena, M.S.; Pasetto, S.; Bezzi, C.; Bettinardi, V.; Andreasi, V.; et al. Dual Tracer 68Ga-DOTATOC and 18F-FDG PET Improve Preoperative Evaluation of Aggressiveness in Resectable Pancreatic Neuroendocrine Neoplasms. *Diagnostics* **2021**, *11*, 192. [[CrossRef](#)]
70. Garin, E.; Le Jeune, F.; Devillers, A.; Cuggia, M.; De Lajarte-Thirouard, A.; Bouriel, C.; Boucher, E.; Raoul, J. Predictive value of 18F-FDG PET and somatostatin receptor scintigraphy in patients with metastatic endocrine tumors. *J. Nucl. Med.* **2009**, *50*, 858–864. [[CrossRef](#)] [[PubMed](#)]
71. Binderup, T.; Knigge, U.; Loft, A.; Federspiel, B.; Kjaer, A. 18F-fluorodeoxyglucose positron emission tomography predicts survival of patients with neuroendocrine tumors. *Clin. Cancer Res.* **2010**, *16*, 978–985. [[CrossRef](#)]
72. Masui, T.; Doi, R.; Ito, T.; Kami, K.; Ogawa, K.; Harada, D.; Uemoto, S. Diagnostic value of 18F-fluorodeoxyglucose positron emission tomography for pancreatic neuroendocrine tumors with reference to the World Health Organization classification. *Oncol. Lett.* **2010**, *1*, 155. [[CrossRef](#)]
73. Cloyd, J.; Poulsides, G. Non-functional neuroendocrine tumors of the pancreas: Advances in diagnosis and management. *World J. Gastroenterol.* **2015**, *21*, 9512. [[CrossRef](#)]
74. Fusco, R.; Setola, S.; Raiano, N.; Granata, V.; Cerciello, V.; Pecori, B.; Petrillo, A. Analysis of a monocentric computed tomography dosimetric database using a radiation dose index monitoring software: Dose levels and alerts before and after the implementation of the adaptive statistical iterative reconstruction on CT images. *Radiol. Medica* **2022**, *127*, 733–742. [[CrossRef](#)]
75. Han, D.; Yu, N.; Yu, Y.; He, T.; Duan, X. Performance of CT radiomics in predicting the overall survival of patients with stage III clear cell renal carcinoma after radical nephrectomy. *Radiol. Med.* **2022**, *127*, 837–847. [[CrossRef](#)]
76. Masci, G.; Ciccarelli, F.; Mattei, F.; Grasso, D.; Accarpi, F.; Catalano, C.; Laghi, A.; Sammartino, P.; Iafrate, F. Role of CT texture analysis for predicting peritoneal metastases in patients with gastric cancer. *Radiol. Med.* **2022**, *127*, 251–258. [[CrossRef](#)]
77. Gregucci, F.; Fiorentino, A.; Mazzola, R.; Ricchetti, F.; Bonaparte, I.; Surgo, A.; Figlia, V.; Carbonara, R.; Caliandro, M.; Ciliberti, M.; et al. Radiomic analysis to predict local response in locally advanced pancreatic cancer treated with stereotactic body radiation therapy. *Radiol. Med.* **2022**, *127*, 100–107. [[CrossRef](#)]
78. Ruf, J.; Schiefer, J.; Furth, C.; Kosiek, O.; Kropf, S.; Heuck, F.; Denecke, T.; Pavel, M.; Pascher, A.; Wiedenmann, B. 68Ga-DOTATOC PET/CT of neuroendocrine tumors: Spotlight on the CT phases of a triple-phase protocol. *J. Nucl. Med.* **2011**, *52*, 697–704. [[CrossRef](#)]
79. Carnevale, M.A.; D’Amato, E.; Pellegrino, F.; Toma, I.; Perrucci, L.; Di Ciesco, C.A.; Labaj, O. Pneumatosis cystoides intestinalis during the treatment with paclitaxel for metastatic ovarian cancer. *EuroMediter. Biomed. J.* **2020**, *15*, 130–133. [[CrossRef](#)]
80. Pinto, A.; Scaglione, M.; Romano, L. Perforation of a gastrojejunal anastomosis due to acute pancreatitis revealed by helical computed tomography. *Acta Radiol.* **2003**, *44*, 572–573. [[CrossRef](#)]
81. Scaglione, M.; Grassi, R.; Pinto, A.; Giovine, S.; Gagliardi, N.; Stavolo, C.; Romano, L. Positive predictive value and negative predictive value of spiral CT in the diagnosis of closed loop obstruction complicated by intestinal ischemia. *Radiol. Med.* **2004**, *107*, 69–77.
82. Scaglione, M.; Romano, S.; Pinto, F.; Flagiello, F.; Farina, R.; Acampora, C.; Romano, L. Helical CT diagnosis of small bowel obstruction in the acute clinical setting. *Eur. J. Radiol.* **2004**, *50*, 15–22. [[CrossRef](#)]
83. Lassandro, F.; Scaglione, M.; Rossi, G.; Grassi, R.; Romano, L. Portomesenteric vein gas: Diagnostic and prognostic value. *Emerg. Radiol.* **2002**, *9*, 96–99. [[CrossRef](#)] [[PubMed](#)]
84. Scaglione, M.; Romano, L.; Bocchini, G.; Sica, G.; Guida, F.; Pinto, A.; Grassi, R. Multidetector computed tomography of pancreatic, small bowel, and mesenteric traumas. *Semin. Roentgenol.* **2012**, *47*, 362–370. [[CrossRef](#)]
85. Patlas, M.; Alabousi, A.; Scaglione, M.; Romano, L.; Soto, J. Cross-sectional imaging of nontraumatic peritoneal and mesenteric emergencies. *Can. Assoc. Radiol. J.* **2013**, *64*, 148–153. [[CrossRef](#)]
86. Flacobellis; Narese, D.; Beritto, D.; Brillantino, A.; Di Serafino, M.; Guerrini, S.; Grassi, R.; Scaglione, M.; Mazzei, M.; Romano, L. Large Bowel Ischemia/Infarction: How to Recognize It and Make Differential Diagnosis? A Review. *Diagnostics* **2021**, *11*, 998. [[CrossRef](#)]
87. Scaglione, M.; Galluzzo, M.; Santucci, D.; Trinci, M.; Messina, L.; Laccetti, E.; Faiella, E.; Zobel, B.B. Small bowel obstruction and intestinal ischemia: Emphasizing the role of MDCT in the management decision process. *Abdom. Radiol.* **2022**, *47*, 1541–1555. [[CrossRef](#)]
88. Scaglione, M.; Pinto, A.; Romano, S.; Scialpi, M.; Volterrani, L.; Rotondo, A.; Romano, L. Using multidetector row computed tomography to diagnose and stage pancreatic carcinoma: The problems and the possibilities. *J. Pancreas* **2005**, *6*, 278.
89. Tamm, E.; Bhosale, P.; Lee, J.; Rohren, E. State-of-the-art Imaging of Pancreatic Neuroendocrine Tumors. *Surg. Oncol. Clin. N. Am.* **2016**, *25*, 375–400. [[CrossRef](#)]
90. Scialpi, M.; Reginelli, A.; D’Andrea, A.; Gravante, S.; Falcone, G.; Baccari, P.; Manganaro, L.; Palumbo, B.; Cappabianca, S. Pancreatic tumors imaging: An update. *Int. J. Surg.* **2016**, *28*, S142–S155. [[CrossRef](#)]

91. Atwi, N.; Sabottke, C.; Pitre, D.; Smith, D.; Danrad, R.; Dharaiya, E.; Kambadakone, A.; Pandharipande, P.; Toshav, A. Follow-up Recommendation Rates Associated with Spectral Detector Dual-Energy CT of the Abdomen and Pelvis: A Retrospective Comparison to Single-Energy CT. *J. Am. Coll. Radiol.* **2020**, *17*, 940–950. [[CrossRef](#)]
92. Johnson, T.; Krauß, B.; Sedlmair, M.; Grasruck, M.; Bruder, H.; Morhard, D.; Fink, C.; Weckbach, S.; Lenhard, M.; Schmidt, B.; et al. Material differentiation by dual energy CT: Initial experience. *Eur. Radiol.* **2007**, *17*, 1510–1517. [[CrossRef](#)] [[PubMed](#)]
93. Cicero, G.; Mazzuotti, S.; Silipigni, S.; Blandino, A.; Cantisani, V.; Pergolizzi, S.; D’Angelo, T.; Stagno, A.; Maimone, S.; Squadrito, G.; et al. Dual-energy CT quantification of fractional extracellular space in cirrhotic patients: Comparison between early and delayed equilibrium phases and correlation with oesophageal varices. *Radiol. Med.* **2021**, *126*, 761–767. [[CrossRef](#)]
94. Nakamura, Y.; Higaki, T.; Honda, Y.; Tatsugami, F.; Tani, C.; Fukumoto, W.; Narita, K.; Kondo, S.; Akagi, M.; Awai, K. Advanced CT techniques for assessing hepatocellular carcinoma. *Radiol. Med.* **2021**, *126*, 925–935. [[CrossRef](#)] [[PubMed](#)]
95. Assadsangabi, R.; Babaei, R.; Songco, C.; Ivanovic, V.; Bobinski, M.; Chen, Y.; Nabavizadeh, S. Multimodality oncologic evaluation of superficial neck and facial lymph nodes. *Radiol. Med.* **2021**, *126*, 1074–1084. [[CrossRef](#)]
96. Salaffi, F.; Carotti, M.; Di Matteo, A.; Ceccarelli, L.; Farah, S.; Villota-Eraso, C.; Di Carlo, M.; Giovagnoni, A. Ultrasound and magnetic resonance imaging as diagnostic tools for sarcopenia in immune-mediated rheumatic diseases (IMRDs). *Radiol. Med.* **2022**, *127*, 1277–1291. [[CrossRef](#)]
97. Lin, X.; Wu, Z.; Tao, R.; Guo, Y.; Li, J.; Zhang, J.; Chen, K. Dual energy spectral CT imaging of insulinoma—Value in preoperative diagnosis compared with conventional multi-detector CT. *Eur. J. Radiol.* **2012**, *81*, 2487–2494. [[CrossRef](#)]
98. Kamaoui, I.; De-Luca, V.; Ficarelli, S.; Mennesson, N.; Lombard-Bohas, C.; Pilleul, F. Value of CT enteroclysis in suspected small-bowel carcinoid tumors. *Am. J. Roentgenol.* **2010**, *194*, 629–633. [[CrossRef](#)]
99. Morse, B.; Al-Toubah, T.; Montilla-Soler, J. Anatomic and Functional Imaging of Neuroendocrine Tumors. *Curr. Treat. Options Oncol.* **2020**, *21*, 1–16. [[CrossRef](#)]
100. Niederle, B.; Pape, U.; Costa, F.; Gross, D.; Kelestimir, F.; Knigge, U.; Öberg, K.; Pavel, M.; Perren, A.; Toumpanakis, C.; et al. ENETS Consensus Guidelines Update for Neuroendocrine Neoplasms of the Jejunum and Ileum. *Neuroendocrinology* **2016**, *103*, 125–138. [[CrossRef](#)]
101. Soyer, P.; Aout, M.; Hoeffel, C.; Vicaut, E.; Placé, V.; Boudiaf, M. Helical CT-enteroclysis in the detection of small-bowel tumours: A meta-analysis. *Eur. Radiol.* **2013**, *23*, 388–399. [[CrossRef](#)]
102. Hakim, F.; Alexander, J.; Huprich, J.; Grover, M.; Enders, F. CT-enterography may identify small bowel tumors not detected by capsule endoscopy: Eight years experience at Mayo Clinic Rochester. *Dig. Dis. Sci.* **2011**, *56*, 2914–2919. [[CrossRef](#)]
103. Granata, V.; Grassi, R.; Fusco, R.; Setola, S.; Belli, A.; Ottiano, A.; Nasti, G.; La Porta, M.; Danti, G.; Cappabianca, S.; et al. Intrahepatic cholangiocarcinoma and its differential diagnosis at MRI: How radiologist should assess MR features. *Radiol. Med.* **2021**, *126*, 1584–1600. [[CrossRef](#)] [[PubMed](#)]
104. Granata, V.; Simonetti, I.; Fusco, R.; Sergio; Setola, V.; Izzo, F.; Scarpato, L.; Vanella, V.; Festino, L.; Simeone, E.; et al. Management of cutaneous melanoma: Radiologists challenging and risk assessment. *Radiol. Med.* **2022**, *127*, 899–911. [[CrossRef](#)]
105. Scola, E.; Desideri, I.; Bianchi, A.; Gadda, D.; Busto, G.; Fiorenza, A.; Amadori, T.; Mancini, S.; Miele, V.; Fainardi, E. Assessment of brain tumors by magnetic resonance dynamic susceptibility contrast perfusion-weighted imaging and computed tomography perfusion: A comparison study. *Radiol. Med.* **2022**, *127*, 664–672. [[CrossRef](#)] [[PubMed](#)]
106. Liu, J.; Wang, C.; Guo, W.; Zeng, P.; Liu, Y.; Lang, N.; Yuan, H. A preliminary study using spinal MRI-based radiomics to predict high-risk cytogenetic abnormalities in multiple myeloma. *Radiol. Med.* **2021**, *126*, 1226–1235. [[CrossRef](#)] [[PubMed](#)]
107. Putzer, D.; Gabriel, M.; Henninger, B.; Kendler, D.; Uprimny, C.; Dobrozemsky, G.; Decristoforo, C.; Bale, R.; Jaschke, W.; Virgolini, I. Bone metastases in patients with neuroendocrine tumor: 68Ga-DOTA-Tyr3-octreotide PET in comparison to CT and bone scintigraphy. *J. Nucl. Med.* **2009**, *50*, 1214–1221. [[CrossRef](#)]
108. Schmid-Tannwald, C.; Schmid-Tannwald, C.; Morelli, J.; Neumann, R.; Haug, A.; Jansen, N.; Nikolaou, K.; Schramm, N.; Reiser, M.; Rist, C. Comparison of abdominal MRI with diffusion-weighted imaging to 68Ga-DOTATATE PET/CT in detection of neuroendocrine tumors of the pancreas. *Eur. J. Nucl. Mol. Imag.* **2013**, *40*, 897–907. [[CrossRef](#)]
109. Brenner, R.; Metens, T.; Bali, M.; Demetter, P.; Matos, C. Pancreatic neuroendocrine tumor: Added value of fusion of T2-weighted imaging and high b-value diffusion-weighted imaging for tumor detection. *Eur. J. Radiol.* **2012**, *81*, e746–e749. [[CrossRef](#)] [[PubMed](#)]
110. d’Assignies, G.; Fina, P.; Bruno, O.; Vullierme, M.; Tubach, F.; Paradis, V.; Sauvanet, A.; Ruszniewski, P.; Vilgrain, V. High sensitivity of diffusion-weighted MR imaging for the detection of liver metastases from neuroendocrine tumors: Comparison with T2-weighted and dynamic gadolinium-enhanced MR imaging. *Radiology* **2013**, *268*, 390–399. [[CrossRef](#)]
111. Ronot, M.; Clift, A.; Baum, R.; Singh, A.; Kulkarni, H.; Frilling, A.; Vilgrain, V. Morphological and Functional Imaging for Detecting and Assessing the Resectability of Neuroendocrine Liver Metastases. *Neuroendocrinology* **2018**, *106*, 74–88. [[CrossRef](#)]
112. Dromain, C.; de Baere, T.; Lumbroso, J.; Caillet, H.; Laplanche, A.; Boige, V.; Ducreux, M.; Duvillard, P.; Elias, D.; Schlumberger, M.; et al. Detection of liver metastases from endocrine tumors: A prospective comparison of somatostatin receptor scintigraphy, computed tomography, and magnetic resonance imaging. *J. Clin. Oncol.* **2005**, *23*, 70–78. [[CrossRef](#)] [[PubMed](#)]

113. Cwikla, J.; Buscombe, J.; Caplin, M.; Watkinson, A.; Walecki, J.; Gorczyca-Wisniewska, E.; Hilson, A. Diagnostic imaging of carcinoid metastases to the abdomen and pelvis. *Med. Sci. Monit.* **2004**, *10*, 9–16. [PubMed]
114. Chambers, A.; Pasieka, J.; Dixon, E.; Rorstad, O. Role of imaging in the preoperative staging of small bowel neuroendocrine tumors. *J. Am. Coll. Surg.* **2010**, *211*, 620–627. [CrossRef] [PubMed]
115. Sundin, Radiological and nuclear medicine imaging of gastroenteropancreatic neuroendocrine tumours. *Best Pract. Res. Clin. Gastroenterol.* **2012**, *26*, 803–818. [CrossRef]
116. Fushimi, Y.; Yoshida, K.; Okawa, M.; Maki, T.; Nakajima, S.; Sakata, A.; Okuchi, S.; Hinoda, T.; Kanagaki, M.; Nakamoto, Y. Vessel wall MR imaging in neuroradiology. *Radiol. Med.* **2022**, *127*, 1032–1045. [CrossRef] [PubMed]
117. Granata, V.; Fusco, R.; De Muzio, F.; Cutolo, C.; Setola, S.; Aversana, F.D.; Ottaiano, A.; Avallone, A.; Nasti, G.; Grassi, F.; et al. Contrast MR-Based Radiomics and Machine Learning Analysis to Assess Clinical Outcomes following Liver Resection in Colorectal Liver Metastases: A Preliminary Study. *Cancers* **2022**, *14*, 1110. [CrossRef] [PubMed]
118. Renzulli, M.; Brandi, N.; Argalia, G.; Brocchi, S.; Farolfi, A.; Fanti, S.; Golfieri, R. Morphological, dynamic and functional characteristics of liver pseudolesions and benign lesions. *Radiol. Med.* **2022**, *127*, 129–144. [CrossRef] [PubMed]
119. Granata, V.; Fusco, R.; Maio, F.; Avallone, A.; Nasti, G.; Palaia, R.; Albino, V.; Grassi, R.; Izzo, F.; Petrillo, A. Qualitative assessment of EOB-GD-DTPA and Gd-BT-DO3A MR contrast studies in HCC patients and colorectal liver metastases. *Infect. Agent. Cancer* **2019**, *14*, 1–9. [CrossRef]
120. Granata, V.; Fusco, R.; Avallone, A.; Filice, F.; Tatangelo, F.; Piccirillo, M.; Grassi, R.; Izzo, F.; Petrillo, A. Critical analysis of the major and ancillary imaging features of LI-RADS on 127 proven HCCs evaluated with functional and morphological MRI: Lights and shadows. *Oncotarget* **2017**, *8*, 51224. [CrossRef]
121. Granata, V.; Fusco, R.; De Muzio, F.; Cutolo, C.; Setola, S.; Dell'aversana, F.; Belli, A.; Romano, C.; Ottaiano, A.; Nasti, G.; et al. Magnetic Resonance Features of Liver Mucinous Colorectal Metastases: What the Radiologist Should Know. *J. Clin. Med.* **2022**, *11*, 2221. [CrossRef]
122. Granata, V.; Fusco, R.; Avallone, A.; Cassata, A.; Palaia, R.; Delrio, P.; Grassi, R.; Tatangelo, F.; Grazzini, G.; Izzo, F.; et al. Abbreviated MRI protocol for colorectal liver metastases: How the radiologist could work in pre surgical setting. *PLoS ONE* **2020**, *15*, e0241431. [CrossRef]
123. Granata, V.; Fusco, R.; de Lutio di Castelguidone, E.; Avallone, A.; Palaia, R.; Delrio, P.; Tatangelo, F.; Botti, G.; Grassi, R.; Izzo, F.; et al. Diagnostic performance of gadoxetic acid-enhanced liver MRI versus multidetector CT in the assessment of colorectal liver metastases compared to hepatic resection. *BMC Gastroenterol.* **2019**, *19*, 129. [CrossRef]
124. Zerunian, M.; Pucciarelli, F.; Caruso, D.; Polici, M.; Masci, B.; Guido, G.; De Santis, D.; Polverari, D.; Principessa, D.; Benvenga, A.; et al. Artificial intelligence based image quality enhancement in liver MRI: A quantitative and qualitative evaluation. *Radiol. Med.* **2022**, *127*, 1098–1105. [CrossRef]
125. Pecoraro, M.; Cipollari, S.; Marchitelli, L.; Messina, E.; Del Monte, M.; Galea, N.; Ciardi, M.; Francone, M.; Catalano, C.; Panebianco, V. Cross-sectional analysis of follow-up chest MRI and chest CT scans in patients previously affected by COVID-19. *Radiol. Med.* **2021**, *126*, 1273. [CrossRef]
126. Cappabianca; Di Grezia, G.; Grassi, V.; Rotondo, A. The role of nasoenteric intubation in the MR study of patients with Crohn's disease: Our experience and literature review. *Radiol. Med.* **2011**, *116*, 389–406. [CrossRef] [PubMed]
127. Kaufmann, T.; Smits, M.; Boxerman, J.; Huang, R.; Barboriak, D.; Weller, M.; Chung, C.; Tsien, C.; Brown, P.; Shankar, L.; et al. Consensus recommendations for a standardized brain tumor imaging protocol for clinical trials in brain metastases. *Neuro. Oncol.* **2020**, *22*, 757–772. [CrossRef] [PubMed]
128. Kiess, W.; Werther, G. Best Practice & Research Clinical Endocrinology and Metabolism. *Best Pract. Res. Clin. Endocrinol. Metab.* **2016**, *30*, 591–601. [CrossRef]
129. Orditura, M.; Petrillo, A.; Ventriglia, J.; Diana, A.; Laterza, M.; Fabozzi, A.; Savastano, B.; Franzese, E.; Conzo, G.; Santini, L. Pancreatic neuroendocrine tumors: Nosography, management and treatment. *Int. J. Surg.* **2016**, *1*, S156–S162. [CrossRef]
130. Halefoglu, A.; Ozagari, A. Tumor grade estimation of clear cell and papillary renal cell carcinomas using contrast-enhanced MDCT and FSE T2 weighted MR imaging: Radiology-pathology correlation. *Radiol. Med.* **2021**, *126*, 1139–1148. [CrossRef]
131. Attili, F.; Capurso, G.; Vanella, G.; Fuccio, L.; Fave, G.; Costamagna, G.; Larghi, A. Diagnostic and therapeutic role of endoscopy in gastroenteropancreatic neuroendocrine neoplasms. *Dig. Liver Dis.* **2014**, *46*, 9–17. [CrossRef] [PubMed]
132. Granata, V.; Fusco, R.; Amato, D.; Albino, V.; Patrone, R.; Izzo, F.; Petrillo, A. Beyond the vascular profile: Conventional DWI, IVIM and kurtosis in the assessment of hepatocellular carcinoma. *Eur. Rev. Med. Pharmacol. Sci.* **2020**, *24*, 7284–7293. [CrossRef]
133. Granata, V.; Fusco, R.; Belli, A.; Danti, G.; Bicci, E.; Cutolo, C.; Petrillo, A.; Izzo, F. Diffusion weighted imaging and diffusion kurtosis imaging in abdominal oncological setting: Why and when. *Infect. Agent. Cancer* **2022**, *17*, 25. [CrossRef]
134. Bruno, F.; Granata, V.; Bellisari, F.; Sgalambro, F.; Tommasino, E.; Palumbo, P.; Arrigoni, F.; Cozzi, D.; Grassi, F.; Brunese, M.; et al. Advanced Magnetic Resonance Imaging (MRI) Techniques: Technical Principles and Applications in Nanomedicine. *Cancers* **2022**, *14*, 1626. [CrossRef]

135. Satake, H.; Ishigaki, S.; Ito, R.; Naganawa, S. Breast Radiology Radiomics in breast MRI: Current progress toward clinical application in the era of artificial intelligence. *Radiol. Med.* **2022**, *127*, 39–56. [[CrossRef](#)]
136. Jensen, J.; Helpern, J. MRI quantification of non-Gaussian water diffusion by kurtosis analysis. *NMR Biomed.* **2010**, *23*, 698–710. [[CrossRef](#)]
137. Chianca, V.; Albano, D.; Messina, C.; Vincenzo, G.; Rizzo, S.; Filippo; Grande, D.; Luca; Sconfienza, M. An update in musculoskeletal tumors: From quantitative imaging to radiomics. *Radiol. Med.* **2021**, *126*, 1095–1105. [[CrossRef](#)]
138. Shi, Y.; Li, X.; Zhang, X.; Zhu, H.; Liu, Y.; Wei, Y.; Sun, Y. Non-gaussian models of 3-Tesla diffusion-weighted MRI for the differentiation of pancreatic ductal adenocarcinomas from neuroendocrine tumors and solid pseudopapillary neoplasms. *Magn. Reson. Imag.* **2021**, *83*, 68–76. [[CrossRef](#)]
139. Le Bihan, D.; Breton, E.; Lallemand, D.; Aubin, M.; Vignaud, J.; Laval-Jeantet, M. Separation of diffusion and perfusion in intravoxel incoherent motion MR imaging. *Radiology* **1988**, *168*, 497–505. [[CrossRef](#)]
140. Le Bihan, D.; Breton, E.; Lallemand, D.; Grenier, P.; Cabanis, E.; Laval-Jeantet, M. MR imaging of intravoxel incoherent motions: Application to diffusion and perfusion in neurologic disorders. *Radiology* **1986**, *161*, 401–407. [[CrossRef](#)]
141. Zeng, P.; Ma, L.; Liu, J.; Song, Z.; Liu, J.; Yuan, H. The diagnostic value of intravoxel incoherent motion diffusion-weighted imaging for distinguishing nonhypervascular pancreatic neuroendocrine tumors from pancreatic ductal adenocarcinomas. *Eur. J. Radiol.* **2022**, *150*. [[CrossRef](#)]
142. Petralia, G.; Zugni, F.; Paul; Summers, E.; Colombo, A.; Pricolo, P.; Grazioli, L.; Colagrande, S.; Giovagnoni, A.; Anwar; et al. Whole-body magnetic resonance imaging (WB-MRI) for cancer screening: Recommendations for use on behalf of the Italian Working Group on Magnetic Resonance. *Radiol. Med.* **2021**, *126*, 1434–1450. [[CrossRef](#)]
143. Schraml, C.; Schwenzer, N.; Sperling, O.; Aschoff, P.; Lichy, M.; Uller, M.M.; Brendle, C.; Werner, M.; Claussen, C.; Pfannenberg, C. Staging of neuroendocrine tumours: Comparison of [68Ga]DOTATOC multiphase PET/CT and whole-body MRI. *Cancer Imag.* **2013**, *13*, 63–72. [[CrossRef](#)]
144. Hope, T.; Pampaloni, M.; Nakakura, E.; VanBrocklin, H.; Slater, J.; Jivan, S.; Aparici, C.; Yee, J.; Bergsland, E. Simultaneous (68)Ga-DOTA-TOC PET/MRI with gadoxetate disodium in patients with neuroendocrine tumor. *Abdom. Imag.* **2015**, *40*, 1432–1440. [[CrossRef](#)]
145. Beiderwellen, K.; Poeppel, T.; Hartung-Knemeyer, V.; Buchbender, C.; Kuehl, H.; Bockisch, A.; Lauenstein, T. Simultaneous 68Ga-DOTATOC PET/MRI in patients with gastroenteropancreatic neuroendocrine tumors: Initial results. *Investig. Radiol.* **2013**, *48*, 273–279. [[CrossRef](#)]
146. Ierardi, A.; Stellato, E.; Pellegrino, G.; Bonelli, C.; Cellina, M.; Renzulli, M.; Biondetti, P.; Carrafiello, G. Fluid-dynamic control microcatheter used with glue: Preliminary experience on its feasibility and safety. *Radiol. Med.* **2022**, *127*, 272–276. [[CrossRef](#)]
147. Bozkurt, M.; Eldem, G.; Bozbulut, U.; Bozkurt, M.; Kılıçkap, S.; Peynircioğlu, B.; Çil, B.; Ergün, E.L.; Volkan-Salancı, B. Factors affecting the response to Y-90 microsphere therapy in the cholangiocarcinoma patients. *Radiol. Med.* **2021**, *126*, 323–333. [[CrossRef](#)]
148. Ierardi, A.; Carnevale, A.; Angileri, S.; Pellegrino, F.; Renzulli, M.; Golfieri, R.; Zhang, D.; Sun, H.; Giganti, M.; Dionigi, G.; et al. Outcomes following minimally invasive imagine-guided percutaneous ablation of adrenal glands. *Gland Surg.* **2020**, *9*, 859–866. [[CrossRef](#)]
149. Ierardi, A.M.; Carnevale, A.; Pellegrino, F.; Di Stefano, G.; Bonelli, C.; Renzulli, M.; Giganti, M.; Carrafiello, G. Uterine Myomas: Extravascular Treatment. *Semin. Ultrasound CT. MR.* **2021**, *42*, 56–74. [[CrossRef](#)]
150. Francica, G.; Meloni, M.; De Sio, I.; Smolock, A.; Brace, C.; Iadevaia, M.; Santambrogio, R.; Sironi, S.; Scaglione, M.; Lee, F. Radiofrequency and microwave ablation of subcapsular hepatocellular carcinoma accessed by direct puncture: Safety and efficacy. *Eur. J. Radiol.* **2016**, *85*, 739–743. [[CrossRef](#)]
151. Li, N.; Wakim, J.; Koethe, Y.; Huber, T.; Schenning, R.; Gade, T.; Hunt, S.; Park, B. Multicenter assessment of augmented reality registration methods for image-guided interventions. *Radiol. Med.* **2022**, *127*, 857–865. [[CrossRef](#)]
152. Spinelli, M.; Balbaa, M.F.; Mauro; Gallazzi, B.; Mohamed; Eid, E.-E.; Hesham; Kotb, T.; Shafei, E.; Anna; et al. Role of percutaneous CT-guided radiofrequency ablation in treatment of intra-articular, in close contact with cartilage and extra-articular osteoid osteomas: Comparative analysis and new classification system. *Radiol. Med.* **2022**, *127*, 1142–1150. [[CrossRef](#)]
153. Gurgitano, M.; Salvatore; Angileri, A.; Giovanni; Rodà, M.; Liguori, A.; Pandolfi, M.; Ierardi, A.; Bradford; Wood, J.; et al. Vascular and Interventional Radiology Interventional Radiology ex-machina: Impact of Artificial Intelligence on practice. *Radiol. Med.* **2021**, *126*, 998–1006. [[CrossRef](#)]
154. Giurazza, F.; Contegiacomo, A.; Calandri, M.; Mosconi, C.; Modestino, F.; Corvino, F.; Anna; Scrofani, R.; Marra, P.; Coniglio, G.; et al. IVC filter retrieval: A multicenter proposal of two score systems to predict application of complex technique and procedural outcome. *Radiol. Med.* **2021**, *126*, 1007–1016. [[CrossRef](#)]
155. De Filippo, M.; Puglisi, S.; D'amuri, F.; Gentili, F.; Paladini, I.; Carrafiello, G.; Maestroni, U.; Paolo; Rio, D.; Ziglioli, F.; et al. CT-guided percutaneous drainage of abdominopelvic collections: A pictorial essay. *Radiol. Med.* **2021**, *126*, 1561–1570. [[CrossRef](#)]
156. Agazzi, G.; Ravanello, M.; Roca, E.; Medicina, D.; Balzarini, P.; Pessina, C.; Vermi, W.; Berruti, A.; Maroldi, R.; Farina, D.; et al. CT texture analysis for prediction of EGFR mutational status and ALK rearrangement in patients with non-small cell lung cancer. *Radiol. Med.* **2021**, *126*, 786–794. [[CrossRef](#)]

157. Qin, H.; Que, Q.; Lin, P.; Li, X.; Wang, X.-R.; He, Y.; Chen, J.-Q.; Yang, H. Magnetic resonance imaging (MRI) radiomics of papillary thyroid cancer (PTC): A comparison of predictive performance of multiple classifiers modeling to identify cervical lymph node metastases before surgery. *Radiol. Med.* **2021**, *126*, 1312–1327. [[CrossRef](#)]
158. Carnevale, A.; Pellegrino, F.; Cossu, A.; Ierardi, A.; Parenti, G.; Carrafiello, G.; Giganti, M. Current concepts in ablative procedures for primary benign liver lesions: A step forward to minimize the invasiveness of treatment when deemed necessary. *Med. Oncol.* **2020**, *37*, 31. [[CrossRef](#)]
159. Stefanini, M.; Simonetti, G. Interventional Magnetic Resonance Imaging Suite (IMRIS): How to build and how to use. *Radiol. Med.* **2022**, *127*, 1063–1067. [[CrossRef](#)]
160. Mansur, A.; Garg, T.; Shrigiriwar, A.; Etezadi, V.; Georgiades, C.; Habibollahi, P.; Huber, T.; Camacho, J.; Nour, S.; Sag, A.; et al. Image-Guided Percutaneous Ablation for Primary and Metastatic Tumors. *Diagnostics* **2022**, *12*, 1300. [[CrossRef](#)]
161. Argalia, G.; Tarantino, G.; Ventura, C.; Campioni, D.; Tagliati, C.; Guardati, P.; Kostandini, A.; Marzioni, M.; Giuseppetti, G.; Giovagnoni, A. Shear wave elastography and transient elastography in HCV patients after direct-acting antivirals. *Radiol. Med.* **2021**, *126*, 894–899. [[CrossRef](#)]
162. Kos-Kudla, B.; Blicharz-Dorniak, J.; Strzelczyk, J.; Baldys-Waligórska, A.; Bednarczuk, T.; Bolanowski, M.; Boratyn-Nowicka, A.; Borowska, M.; Cichocki, A.; Cwikla, J.; et al. Diagnostic and therapeutic guidelines for gastro-entero-pancreatic neuroendocrine neoplasms (recommended by the Polish Network of Neuroendocrine Tumours). *Endokrynol. Pol.* **2017**, *68*, 79–110. [[CrossRef](#)]
163. Anderson, M.; Carpenter, S.; Thompson, N.; Nostrant, T.; Elta, G.; Scheiman, J. Endoscopic ultrasound is highly accurate and directs management in patients with neuroendocrine tumors of the pancreas. *Am. J. Gastroenterol.* **2000**, *95*, 2271–2277. [[CrossRef](#)] [[PubMed](#)]
164. Tanaka, H.; Matsusaki, S. The Utility of Endoscopic-Ultrasonography-Guided Tissue Acquisition for Solid Pancreatic Lesions. *Diagnostics* **2022**, *12*, 753. [[CrossRef](#)] [[PubMed](#)]
165. Takahashi, K.; Yasuda, I.; Hanaoka, T.; Hayashi, Y.; Araki, Y.; Motoo, I.; Kajiura, S.; Ando, T.; Fujinami, H.; Tajiri, K.; et al. Diagnostic Fine-Needle Biopsy of Small Solid Pancreatic Lesions Using a Franseen Needle during Endoscopic Ultrasound Examination. *Diagnostics* **2020**, *11*, 27. [[CrossRef](#)]
166. Ishii, T.; Katanuma, A.; Toyonaga, H.; Chikugo, K.; Nasuno, H.; Kin, T.; Hayashi, T.; Takahashi, K. Role of Endoscopic Ultrasound in the Diagnosis of Pancreatic Neuroendocrine Neoplasms. *Diagnostics* **2021**, *11*, 316. [[CrossRef](#)] [[PubMed](#)]
167. Tamura, T.; Sugihara, Y.; Yamazaki, H.; Koutani, H.; Tamura, T.; Tsuda, I.; Emori, T.; Kawaiji, Y.; Hatamaru, K.; Yamashita, Y.; et al. Contrast-Enhanced Harmonic Endoscopic Ultrasound for Diagnosis of the Aggressiveness of Pancreatic Neuroendocrine Neoplasm. *Diagnostics* **2022**, *12*, 2988. [[CrossRef](#)]
168. Manta, R.; Nardi, E.; Pagano, N.; Ricci, C.; Sica, M.; Castellani, D.; Bertani, H.; Piccoli, M.; Mullineris, B.; Tringali, A.; et al. Pre-operative Diagnosis of Pancreatic Neuroendocrine Tumors with Endoscopic Ultrasonography and Computed Tomography in a Large Series. *J. Gastrointestin. Liver Dis.* **2016**, *25*, 317–321. [[CrossRef](#)]
169. James, P.; Tsolakis, A.; Zhang, M.; Bellettrutti, P.; Mohamed, R.; Roberts, D.; Heitman, S. Incremental benefit of preoperative EUS for the detection of pancreatic neuroendocrine tumors: A meta-analysis. *Gastrointest. Endosc.* **2015**, *81*, 848–856.e1. [[CrossRef](#)]
170. Pavel, M.; Öberg, K.; Falconi, M.; Krenning, E.; Sundin, A.; Perren, A.; Berruti, A. Gastroenteropancreatic neuroendocrine neoplasms: ESMO Clinical Practice Guidelines for diagnosis, treatment and follow-up. *Ann. Oncol. Off. J. Eur. Soc. Med. Oncol.* **2020**, *31*, 844–860. [[CrossRef](#)] [[PubMed](#)]
171. Ciaravino, V.; De Robertis, R.; Martini, P.T.; Cardobi, N.; Cingarlini, S.; Amodio, A.; Landoni, L.; Capelli, P.; D’Onofrio, M. Imaging presentation of pancreatic neuroendocrine neoplasms. *Insights Imag.* **2018**, *9*, 943. [[CrossRef](#)] [[PubMed](#)]
172. Lee, D.; Kim, M.; Kim, H. Diagnosis of Pancreatic Neuroendocrine Tumors. *Clin. Endosc.* **2017**, *50*, 537. [[CrossRef](#)]
173. Partelli, S.; Cirocchi, R.; Crippa, S.; Cardinali, L.; Fendrich, V.; Bartsch, D.; Falconi, M. Systematic review of active surveillance versus surgical management of asymptomatic small non-functioning pancreatic neuroendocrine neoplasms. *Br. J. Surg.* **2017**, *104*, 34–41. [[CrossRef](#)] [[PubMed](#)]
174. Fusco, R.; Granata, V.; Sansone, M.; Rega, D.; Delrio, P.; Tatangelo, F.; Romano, C.; Avallone, A.; Pupo, D.; Giordano, M.; et al. Validation of the standardized index of shape tool to analyze DCE-MRI data in the assessment of neo-adjuvant therapy in locally advanced rectal cancer. *Radiol. Med.* **2021**, *126*, 1044–1054. [[CrossRef](#)] [[PubMed](#)]
175. Ichikawa, T.; Peterson, M.; Federle, M.; Baron, R.; Haradome, H.; Kawamori, Y.; Nawano, S.; Araki, T. Islet cell tumor of the pancreas: Biphasic CT versus MR imaging in tumor detection. *Radiology* **2000**, *216*, 163–171. [[CrossRef](#)] [[PubMed](#)]
176. Sundin, A.; Vullierme, M.; Kaltsas, G.; Plöckinger, U.; Åkerström, G.; Annibale, B.; Arnold, R.; Bajetta, E.; Barkmanova, J.; Chen, Y.; et al. ENETS Consensus Guidelines for the Standards of Care in Neuroendocrine Tumors: Radiological examinations. *Neuroendocrinology* **2009**, *90*, 167–183. [[CrossRef](#)] [[PubMed](#)]
177. Bhosale, P.; Shah, A.; Wei, W.; Varadhachary, G.; Johnson, V.; Shah, V.; Kundra, V. Carcinoid tumours: Predicting the location of the primary neoplasm based on the sites of metastases. *Eur. Radiol.* **2012**, *23*, 400–407. [[CrossRef](#)]
178. Kang, Y.; Cho, J.-H.; Se, Hwang, H. Diagnostic value of various criteria for deep lobe involvement in radiologic studies with parotid mass: A systematic review and meta-analysis. *Radiol. Med.* **2022**, *127*, 1124–1133. [[CrossRef](#)]
179. De Robertis, R.; Geraci, L.; Tomaiuolo, L.; Bortoli, L.; Beleù, A.; Malleo, G.; D’Onofrio, M. Liver metastases in pancreatic ductal adenocarcinoma: A predictive model based on CT texture analysis. *Radiol. Med.* **2022**, *127*, 1079–1084. [[CrossRef](#)]

180. Chan, M.; Ma, K.A. Chan, Surgical management of neuroendocrine tumor-associated liver metastases: A review. *Gland Surg.* **2018**, *7*, 28–35. [[CrossRef](#)]
181. Alagusundaramoorthy, S.; Gedaly, R. Role of surgery and transplantation in the treatment of hepatic metastases from neuroendocrine tumor. *World J. Gastroenterol.* **2014**, *20*, 14348. [[CrossRef](#)]
182. Mazzaferro, V.; Sposito, C.; Coppa, J.; Miceli, R.; Bhoori, S.; Bongini, M.; Camerini, T.; Milione, M.; Regalia, E.; Spreafico, C.; et al. The Long-Term Benefit of Liver Transplantation for Hepatic Metastases from Neuroendocrine Tumors. *Am. J. Transpl.* **2016**, *16*, 2892–2902. [[CrossRef](#)] [[PubMed](#)]
183. Ravaioli, M.; Ercolani, G.; Neri, F.; Cescon, M.; Stacchini, G.; Del Gaudio, M.; Cucchetti, A.; Pinna, A. Liver transplantation for hepatic tumors: A systematic review. *World J. Gastroenterol.* **2014**, *20*, 5345–5352. [[CrossRef](#)]
184. Veenendaal, L.; Rinkes, I.B.; Lips, C.; van Hillegersberg, R. Liver metastases of neuroendocrine tumours; early reduction of tumour load to improve life expectancy. *World J. Surg. Oncol.* **2006**, *4*, 35. [[CrossRef](#)]
185. Barat, M.; Cottreau, A.; Kedra, A.; Dermine, S.; Palmieri, L.; Coriat, R.; Dautry, R.; Tselikas, L.; Soyer, P.; Dohan, A. The Role of Interventional Radiology for the Treatment of Hepatic Metastases from Neuroendocrine Tumor: An Updated Review. *J. Clin. Med.* **2020**, *9*, 2302. [[CrossRef](#)] [[PubMed](#)]
186. Maire, F.; Lombard-Bohas, C.; O'Toole, D.; Vullierme, M.; Rebours, V.; Couvelard, A.; Pelletier, A.; Zappa, M.; Pilleul, F.; Hentic, O.; et al. Hepatic arterial embolization versus chemoembolization in the treatment of liver metastases from well-differentiated midgut endocrine tumors: A prospective randomized study. *Neuroendocrinology* **2012**, *96*, 294–300. [[CrossRef](#)]
187. Fiorentini, G.; Sarti, D.; Nani, R.; Aliberti, C.; Fiorentini, C.; Guadagni, S. Updates of colorectal cancer liver metastases therapy: Review on DEBIRI. *Hepatic Oncol.* **2020**, *7*, HEP16. [[CrossRef](#)] [[PubMed](#)]
188. Pitt, S.; Knuth, J.; Keily, J.; McDermott, J.; Weber, S.; Chen, H.; Rilling, W.; Quebbeman, E.; Agarwal, D.; Pitt, H. Hepatic neuroendocrine metastases: Chemo- or bland embolization? *J. Gastrointest. Surg.* **2008**, *12*, 1951–1960. [[CrossRef](#)]
189. Bloomston, M.; Al-Saif, O.; Klemanski, D.; Pinzone, J.; Martin, E.; Palmer, B.; Guy, G.; Khabiri, H.; Ellison, E.; Shah, M. Hepatic artery chemoembolization in 122 patients with metastatic carcinoid tumor: Lessons learned. *J. Gastrointest. Surg.* **2007**, *11*, 264–271. [[CrossRef](#)] [[PubMed](#)]
190. Strosberg, J.; Choi, J.; Cantor, A.; Kvols, L. Selective hepatic artery embolization for treatment of patients with metastatic carcinoid and pancreatic endocrine tumors. *Cancer Control.* **2006**, *13*, 72–78. [[CrossRef](#)]
191. Osborne, D.; Zervos, E.; Strosberg, J.; Boe, B.; Malafa, M.; Rosemurgy, A.; Yeatman, T.; Carey, L.; Duhaime, L.; Kvols, L. Improved outcome with cytoreduction versus embolization for symptomatic hepatic metastases of carcinoid and neuroendocrine tumors. *Ann. Surg. Oncol.* **2006**, *13*, 572–581. [[CrossRef](#)] [[PubMed](#)]
192. Gupta, S.; Yao, J.; Ahrar, K.; Wallace, M.; Morello, F.; Madoff, D.; Murthy, R.; Hicks, M.; Ajani, J. Hepatic artery embolization and chemoembolization for treatment of patients with metastatic carcinoid tumors: The M.D. Anderson experience. *Cancer J.* **2003**, *9*, 261–267. [[CrossRef](#)]
193. Brown, K.; Koh, B.; Brody, L.; Getrajdman, G.; Susman, J.; Fong, Y.; Blumgart, L. Particle embolization of hepatic neuroendocrine metastases for control of pain and hormonal symptoms. *J. Vasc. Interv. Radiol.* **1999**, *10*, 397–403. [[CrossRef](#)] [[PubMed](#)]
194. de Mestier, L.; Lepage, C.; Baudin, E.; Coriat, R.; Courbon, F.; Couvelard, A.; Cao, C.D.; Frampas, E.; Gaujoux, S.; Gincul, R.; et al. Digestive Neuroendocrine Neoplasms (NEN): French Intergroup clinical practice guidelines for diagnosis, treatment and follow-up (SNFGE, GTE, RENATEN, TENPATH, FFCD, GERCOR, UNICANCER, SFCD, SFED, SFRO, SFR). *Dig. Liver Dis.* **2020**, *52*, 473–492. [[CrossRef](#)]
195. Clouse, M.; Perry, L.; Stuart, K.; Stokes, K. Hepatic Arterial Chemoembolization for Metastatic Neuroendocrine Tumors. *Digestion* **1994**, *55*, 92–97. [[CrossRef](#)] [[PubMed](#)]
196. Frilling, A.; Clift, A.; Braat, A.; Alsafi, A.; Wasan, H.; Al-Nahhas, A.; Thomas, R.; Drymousis, P.; Habib, N.; Tait, P. Radioembolisation with 90Y microspheres for neuroendocrine liver metastases: An institutional case series, systematic review and meta-analysis. *HPB* **2019**, *21*, 773–783. [[CrossRef](#)]
197. Braat, A.; Kappadath, S.; Ahmadzadehfar, H.; Stothers, C.; Frilling, A.; Deroose, C.; Flamen, P.; Brown, D.; Sze, D.; Mahvash, A.; et al. Radioembolization with 90Y Resin Microspheres of Neuroendocrine Liver Metastases: International Multicenter Study on Efficacy and Toxicity. *Cardiovasc. Intervent. Radiol.* **2019**, *42*, 413–425. [[CrossRef](#)] [[PubMed](#)]
198. Fan, K.; Wild, A.; Halappa, V.; Kumar, R.; Ellsworth, S.; Ziegler, M.; Garg, T.; Rosati, L.; Su, Z.; Hacker-Prietz, A.; et al. Neuroendocrine tumor liver metastases treated with yttrium-90 radioembolization. *Contemp. Clin. Trials.* **2016**, *50*, 143–149. [[CrossRef](#)]
199. Peker, A.; Çiçek, O.; Soydal, Ç.; Küçük, N.; Bilgiç, S. Radioembolization with yttrium-90 resin microspheres for neuroendocrine tumor liver metastases. *Diagn. Interv. Radiol.* **2015**, *21*, 54–59. [[CrossRef](#)]
200. Memon, K.; Lewandowski, R.; Mulcahy, M.; Riaz, A.; Ryu, R.; Sato, K.; Gupta, R.; Nikolaidis, P.; Miller, F.; Yaghmai, V.; et al. Radioembolization for neuroendocrine liver metastases: Safety, imaging, and long-term outcomes. *Int. J. Radiat. Oncol. Biol. Phys.* **2012**, *83*, 887–894. [[CrossRef](#)]
201. Barbier, C.; Garske-Román, U.; Sandström, M.; Nyman, R.; Granberg, D. Selective internal radiation therapy in patients with progressive neuroendocrine liver metastases. *Eur. J. Nucl. Med. Mol. Imag.* **2016**, *43*, 1425–1431. [[CrossRef](#)]
202. Paprottka, P.; Hoffmann, R.; Haug, A.; Sommer, W.; Raebler, F.; Trumm, C.; Schmidt, G.; Ashoori, N.; Reiser, M.; Jakobs, T. Radioembolization of symptomatic, unresectable neuroendocrine hepatic metastases using yttrium-90 microspheres. *Cardiovasc. Intervent. Radiol.* **2012**, *35*, 334–342. [[CrossRef](#)] [[PubMed](#)]

203. Devcic, Z.; Rosenberg, J.; Braat, A.; Techasith, T.; Banerjee, A.; Sze, D.; Lam, M. The Efficacy of Hepatic 90Y Resin Radioembolization for Metastatic Neuroendocrine Tumors: A Meta-Analysis. *J. Nucl. Med.* **2014**, *55*, 1404–1410. [CrossRef]
204. Jia, Z.; Wang, W. Yttrium-90 radioembolization for unresectable metastatic neuroendocrine liver tumor: A systematic review. *Eur. J. Radiol.* **2018**, *100*, 23–29. [CrossRef]
205. Ceelen, F.; Theisen, D.; De Albéniz, X.; Auernhammer, C.; Haug, A.; D'Anastasi, M.; Paprottka, P.; Rist, C.; Reiser, M.; Sommer, W. Towards new response criteria in neuroendocrine tumors: Which changes in MRI parameters are associated with longer progression-free survival after radioembolization of liver metastases? *J. Magn. Reson. Imag.* **2015**, *41*, 361–368. [CrossRef]
206. Sommer, W.; Ceelen, F.; García-Albéniz, X.; Paprottka, P.; Auernhammer, C.; Armbruster, M.; Nikolaou, K.; Haug, A.; Reiser, M.; Theisen, D. Defining predictors for long progression-free survival after radioembolisation of hepatic metastases of neuroendocrine origin. *Eur. Radiol.* **2013**, *23*, 3094–3103. [CrossRef]
207. Filippi, L.; Scopinaro, F.; Pelle, G.; Cianni, R.; Salvatori, R.; Schillaci, O.; Bagni, O. Molecular response assessed by 68Ga-DOTANOC and survival after 90Y microsphere therapy in patients with liver metastases from neuroendocrine tumours. *Eur. J. Nucl. Med. Mol. Imag.* **2016**, *43*, 432–440. [CrossRef]
208. Fendrich, V.; Langer, P.; Celik, I.; Bartsch, D.; Zielke, A.; Ramaswamy, A.; Rothmund, M. An aggressive surgical approach leads to long-term survival in patients with pancreatic endocrine tumors. *Ann. Surg.* **2006**, *244*, 845–851. [CrossRef] [PubMed]
209. Pavel, M.; Baudin, E.; Couvelard, A.; Krenning, E.; Öberg, K.; Steinmüller, T.; Anlauf, M.; Wiedenmann, B.; Salazar, R. ENETS Consensus Guidelines for the management of patients with liver and other distant metastases from neuroendocrine neoplasms of foregut, midgut, hindgut, and unknown primary. *Neuroendocrinology* **2012**, *95*, 157–176. [CrossRef] [PubMed]
210. Frilling, A.; Modlin, I.; Kidd, M.; Russell, C.; Breitenstein, S.; Salem, R.; Kwekkeboom, D.; Lau, W.; Klerys, C.; Vilgrain, V.; et al. Recommendations for management of patients with neuroendocrine liver metastases. *Lancet. Oncol.* **2014**, *15*, e8–e21. [CrossRef]
211. Bocchini, M.; Nicolini, F.; Severi, S.; Bongiovanni, A.; Ibrahim, T.; Simonetti, G.; Grassi, I.; Mazza, M. Biomarkers for Pancreatic Neuroendocrine Neoplasms (PanNENs) Management—An Updated Review. *Front. Oncol.* **2020**, *10*. [CrossRef] [PubMed]
212. Fehrenbach, U.; Xin, S.; Hartenstein, A.; Auer, T.; Dräger, F.; Froböse, K.; Jann, H.; Mogl, M.; Amthauer, H.; Geisel, D.; et al. Automatized Hepatic Tumor Volume Analysis of Neuroendocrine Liver Metastases by Gd-EOB MRI-A Deep-Learning Model to Support Multidisciplinary Cancer Conference Decision-Making. *Cancers* **2021**, *13*, 2726. [CrossRef]
213. Haider, M.; Jiang, B.; Parker, J.; Bullock, A.; Goehler, A.; Tsai, L. Use of MRI and Ga-68 DOTATATE for the detection of neuroendocrine liver metastases. *Abdom. Radiol.* **2022**, *47*, 586–595. [CrossRef] [PubMed]
214. Morin, C.; Drolet, S.; Daigle, C.; Deshaies, I.; Ouellet, J.; Ball, C.; Dixon, E.; Marceau, J.; Ouellet, J. Additional value of gadoxetic acid-enhanced MRI to conventional extracellular gadolinium-enhanced MRI for the surgical management of colorectal and neuroendocrine liver metastases. *HPB* **2020**, *22*, 710–715. [CrossRef] [PubMed]
215. Granata, V.; Fusco, R.; De Muzio, F.; Cutolo, C.; Setola, S.V.; Dell, F.; Grassi, F.; Belli, A.; Silvestro, L.; Ottaiano, A.; et al. Radiomics and machine learning analysis based on magnetic resonance imaging in the assessment of liver mucinous colorectal metastases. *Radiol. Med.* **2022**, *127*, 763–772. [CrossRef] [PubMed]
216. Granata, V.; Fusco, R.; De Muzio, F.; Cutolo, C.; Raso, M.; Gabelloni, M.; Avallone, A.; Ottaiano, A.; Tatangelo, F.; Brunese, M.; et al. Radiomics and Machine Learning Analysis Based on Magnetic Resonance Imaging in the Assessment of Colorectal Liver Metastases Growth Pattern. *Diagnostics* **2022**, *12*, 1115. [CrossRef] [PubMed]
217. Sun, J.; Li, H.; Gao, J.; Li, J.; Li, M.; Zhou, Z.; Peng, Y. Performance evaluation of a deep learning image reconstruction (DLIR) algorithm in “double low” chest CTA in children: A feasibility study. *Radiol. Med.* **2021**, *126*, 1181–1188. [CrossRef] [PubMed]
218. Granata, V.; Fusco, R.; Avallone, A.; De Stefano, A.; Ottaiano, A.; Sbordone, C.; Brunese, L.; Izzo, F.; Petrillo, A. Radiomics-Derived Data by Contrast Enhanced Magnetic Resonance in RAS Mutations Detection in Colorectal Liver Metastases. *Cancers* **2021**, *13*, 453. [CrossRef]
219. Sandor, A.; Modlin, I. A retrospective analysis of 1570 appendiceal carcinoids. *Am. J. Gastroenterol.* **1998**, *93*, 422–428. [CrossRef] [PubMed]
220. Modlin, I.; Kidd, M.; Latich, I.; Zikusoka, M.; Shapiro, M. Current status of gastrointestinal carcinoids. *Gastroenterology* **2005**, *128*, 1717–1751. [CrossRef]
221. Turaga, K.; Kvols, L. Recent progress in the understanding, diagnosis, and treatment of gastroenteropancreatic neuroendocrine tumors. *Cancer J. Clin.* **2011**, *61*, 113–132. [CrossRef]
222. Chang, S.; Choi, D.; Soon, J.; Won, J.; Park, M.; Sang, W.; Da, K.; Jang, K. Neuroendocrine neoplasms of the gastrointestinal tract: Classification, pathologic basis, and imaging features. *Radiographics* **2007**, *27*, 1667–1679. [CrossRef]
223. Heller, M.; Shah, A. Imaging of neuroendocrine tumors. *Radiol. Clin. North Am.* **2011**, *49*, 529–548. [CrossRef] [PubMed]
224. Paski, S.; Semrad, C. Small bowel tumors, Gastrointest. Endosc. *Clin. N. Am.* **2009**, *19*, 461–479. [CrossRef]
225. Paulsen, S.; Huprich, J.; Fletcher, J.; Booya, F.; Young, B.; Fidler, J.; Johnson, C.; Barlow, J. CT enterography as a diagnostic tool in evaluating small bowel disorders: Review of clinical experience with over 700 cases. *Radiographics* **2006**, *26*, 641–657. [CrossRef]
226. Pickhardt, P. Differential Diagnosis of Polypoid Lesions Seen at CT Colonography (Virtual Colonoscopy)1. *Radiographics* **2004**, *24*, 1535–1556. [CrossRef]
227. Dasari, A.; Shen, C.; Halperin, D.; Zhao, B.; Zhou, S.; Xu, Y.; Shih, T.; Yao, J. Trends in the Incidence, Prevalence, and Survival Outcomes in Patients with Neuroendocrine Tumors in the United States. *JAMA Oncol.* **2017**, *3*, 1335–1342. [CrossRef] [PubMed]
228. Hallet, J.; Law, C.; Cukier, M.; Saskin, R.; Liu, N.; Singh, S. Exploring the rising incidence of neuroendocrine tumors: A population-based analysis of epidemiology, metastatic presentation, and outcomes. *Cancer* **2015**, *121*, 589–597. [CrossRef]

229. Meyerhardt, J.; Mangu, P.; Flynn, P.; Korde, L.; Loprinzi, C.; Minsky, B.; Petrelli, N.; Ryan, K.; Schrag, D.; Wong, S.; et al. Follow-up care, surveillance protocol, and secondary prevention measures for survivors of colorectal cancer: American Society of Clinical Oncology clinical practice guideline endorsement. *J. Clin. Oncol.* **2013**, *31*, 4465–4470. [[CrossRef](#)]
230. Pellegrino, F.; Carnevale, A.; Bisi, R.; Cavedagna, D.; Reverberi, R.; Uccelli, L.; Leprotti, S.; Giganti, M. Best Practices on Radiology Department Workflow: Tips from the Impact of the COVID-19 Lockdown on an Italian University Hospital. *Healthcare* **2022**, *10*, 1771. [[CrossRef](#)]

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.