

Proceedings



Polypyrrole Fibre Electrodes for Drug Sensing +

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- Presented at the International Conference on the Challenges, Opportunities, Innovations and Applications in Electronic Textiles (E-Textiles 2019), London, UK, 12 November 2019.

Published: 18 January 2020

Abstract: Polypyrrole (PPy) fibre electrodes and their ability to sense paracetamol (as a model drug) in addition to interferents such as ascorbic acid and dopamine were studied. PPy was electrodeposited onto carbon fibre (CF) through electropolymerisation using cyclic voltammetry in the presence of two different counter anions: potassium nitrate (KNO₃) and sodium dodecyl sulfate (SDS). PPy with SDS as dopant could sense paracetamol with an oxidation peak at 0.55 V vs. Ag/AgCl. The limit of detection of this fibre sensor was found to be 1 μ M with a linear range of 1–100 μ M of paracetamol (R² = 0.985).

Keywords: polypyrrole; fibre electrode; cyclic voltammetry; drug sensing; wearable sensors

1. Introduction

Polypyrrole (PPy) is a biocompatible conductive polymer, with good electrical conductivity, electrochemical switching, and flexibility, which makes it an ideal flexible electrode material [1,2]. Counterions are employed to balance the charge of the PPy film. Anions are incorporated during oxidation to stabilise the radical cation intermediate and are then displaced during reduction, whereas cations merge with the reduced PPy film to balance the negative charge [3,4]. Figure 1 illustrates the structure of PPy in its oxidised form (p-doped), with a counterion used to balance the charge. The aim of this work was to fabricate a fibre electrode with a PPy coating to improve the ability of the fibre electrode to sense paracetamol (as model drug) in addition to interferents such as ascorbic acid and dopamine.



Figure 1. Schematic of polypyrrole (PPy) in the oxidised form with different counterions (**a**) dodecylsulfate; (**b**) nitrate.

2. Materials and Methods

2.1. Reagents

Analytical grade pyrrole (98%), potassium nitrate (KNO₃), sodium dodecyl sulfate (SDS), paracetamol, dopamine hydrochloride, and L-ascorbic acid were purchased from Sigma-Aldrich

(Gillingham, UK). Phosphate buffered saline (PBS) tablets were purchased from Fisher Scientific (Loughborough, UK). Carbon fibre was purchased from Alfa Aesar (Heysham, UK).

2.2. Apparatus

All cyclic voltammetry (CV) was performed using an eDAQ EA161 potentiostat with an eDAQ e-corder 401 supported by EChem V2.1.16 software. A Basi® Ag/AgCl in 3.0 M NaCl was used as a reference electrode and a platinum mesh was used as a counter electrode. Raman spectroscopy was performed using a Renishaw inVia confocal Raman microscope with a 785 nm laser. Scanning electron microscopy (SEM) images were obtained using a JEOL USA JSM-7100 F field emission electron microscope.

2.3. Procedure

PPy was electrodeposited onto carbon fibre (CF) through electropolymerisation using CV in the presence of two different counter anions: KNO₃ and SDS. Deposition occurred over a potential range of –0.2 to +1.0 V (vs. Ag/AgCl) at a scan rate of 50 mV s⁻¹. The concentration of pyrrole was varied (from 0.02 to 0.2 M) together with the number of cycles (from 5 to 15 cycles) to optimize the PPy sensing layer.

The performance of the sensor with respect to paracetamol, interferents such as ascorbic acid, and dopamine (commonly found in blood) was tested [5]. The three-electrode system was used for analysis by CV at a potential range of -0.6 to +0.9 V vs. Ag/AgCl at a scan rate of 50 mV s⁻¹ in $0-500 \mu$ M of analyte solution.

3. Results and Discussion

3.1. Characterisation

Figure 2 shows the Raman spectrum of an uncoated carbon fibre, which illustrates peaks at 1577 and 1377 cm⁻¹ corresponding to the G and D bands of carbon, respectively [6]. The Raman spectrum of a PPy.SDS coated CF confirms the deposition of PPy film onto the fibre electrode surface. The peak at 1572 cm⁻¹ corresponds to C=C stretching that is associated with the existence of polarons. The spectrum also displays the skeletal band at 1477 cm⁻¹ and an antisymmetrical C–N stretching at 1384 cm⁻¹. The peaks at 928 cm⁻¹ and 1062 cm⁻¹ relate to the bipolaron structure and those at 962 cm⁻¹ and 1043 cm⁻¹ to the polaron structure of PPy.



Figure 2. Raman spectrum of uncoated carbon fibre (red line) and PPy.SDS coated carbon fibre (blue line).



Figure 3. Scanning electron microscope images of (a) uncoated carbon fibre; (b) PPy.SDS coated carbon fibre.

3.2. Surface Area of the Fibre Electrode

Cyclic voltammetry post-polymerization (Figure 4a) show oxidation peak potentials at –0.75 and 0.10 V (vs. Ag/AgCl) for the PPy.SDS and PPy.KNO₃ films on CF, respectively. The surface area of the sensing fibre was calculated using the Randles–Sevcik equation; cyclic voltammetry was performed in 10 mM K₃Fe(CN)₆ and 0.1 M KCl as supporting electrolyte at different scan rates. Table 1 shows how the PPy.SDS film on CF gave a five-fold increase in electrochemical surface area, whereas coating with PPy.KNO₃ resulted in a four-fold increase. Figure 4b displays the CVs of uncoated and PPy coated CF in potassium ferricyanide solution.



Figure 4. (a) Post-polymerisation cyclic voltammetry of PPy.SDS and PPy.KNO₃ on carbon fibre; (b) Cyclic voltammograms obtained for the fibre electrodes in 10 mM K₃Fe(CN)₆ and 0.1 M KCl solution at a scan rate of 50 mVs⁻¹.

Table 1. Surface area of uncoated carbon fibre and PPy-coated carbon fibre.

Scan Rate (mV s ⁻¹)	Surface Area (cm²)		
	CF	PPy.SDS on CF	PPy.KNO3 on CF
50	0.18	0.92	0.71

3.3. Drug Sensing

The oxidation of paracetamol of the CF electrode was not evidenced in cyclic voltammetry. Figure 5a shows the sensing ability of PPy.SDS on CF in various concentrations of paracetamol at a scan rate of 50 mVs⁻¹. The oxidation peak of paracetamol was found at 0.55 V (vs. Ag/AgCl). Furthermore, the PPy.SDS on CF could sense dopamine and ascorbic acid at oxidation peaks of 0.35 and 0.25 V (vs. Ag/AgCl), respectively (results not shown). PPy.KNO₃ was not a stable electrode,

as evidenced by changing signals during analysis. A calibration curve for paracetamol sensing with PPy.SDS on CF is shown in Figure 5b using the peak current value plotted against concentration. The limit of detection of this fibre sensor was found to be 1 μ M with a linear range of 1–100 μ M of paracetamol (R² = 0.985).



Figure 5. (a) PPy.SDS on CF sensing various concentration of paracetamol in 0.1 M PBS solution at a scan rate of 50 mVs⁻¹; (b) calibration curve for PPy.SDS on CF with a concentration range of 0–500 μ M paracetamol in 0.1 M PBS solution.

4. Conclusions

The electropolymerisation of PPy onto a carbon fibre electrode improves the sensor's ability to detect paracetamol, ascorbic acid, and dopamine. The PPy.SDS films provided insight into how a large counterion positively affects the sensing capabilities of the film, as the use of SDS shows a good response and improved the stability of the film compared to PPy.KNO₃ (which was not a stable sensor). Further work is underway to sense a broader range of drug molecules and to improve the fibre sensor response with further surface modification.

Acknowledgments: The authors thank the Royal Thai Government Scholarship for providing funding for this work as well as the EPSRC grant, EP/M022749/1.

Conflicts of Interest: The authors declare no conflict of interest and the funders had no role in the design of the study; in the collection, analyses, or interpretation of data; in the writing of the manuscript, or in the decision to publish the results.

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