

Cumulative Effect of the Aqueous and Ethanolic Extracts of *Annona Reticulate* and *Allium Sativum* on Brine Shrimp Lethality Assay [†]

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Abstract: The present study was conducted to test for in vivo Brine Shrimp Lethality Assay (BSLA) of the Aqueous and ethanolic extracts *Annona reticulata* Linn. and *Allium sativum* and correlate cytotoxicity results with known pharmacological activities of the plants. Cytotoxicity was evaluated in terms of LC50 (lethality concentration). Ten nauplii were added into three replicates of each concentration of the plant extract. After 24 h the surviving brine shrimp larvae were counted and LC50 was assessed. Results showed that the presence of alkaloids, tannins, and flavonoids could be accounted for its cytotoxic properties. In the other hand, studies have shown that the leaf extracts of *Alcoholic and aqueous extract of Annona reticulata* and *bulbs of Allium sativum* extracts exhibited cumulative activity when they were combined and compared. Thus, the results on the leaf extracts of *Alcoholic and aqueous extract of Annona reticulata* and *bulbs of Allium sativum* exhibited increase in activity support its use in traditional medicine.

Keywords: brine shrimp lethality assay; *Annona reticulata*; *Allium sativum*; LC50; potent; cytotoxicity

1. Introduction

The crushed leaves of *A. reticulata* are used as poultice on boils, ulcers and abscesses and leaf decoction is used as vermifuge. The tree is not especially attractive. It is erect, with a rounded or spreading crown and trunk 10 to 14 in (25–35 cm) thick. Height ranges from 15 to 35 ft (4.5–10 m). The ill-smelling leaves are deciduous, alternate, oblong or narrow-lanceolate, 4 to 8 in (10–20 cm) long, 3/4 to 2 in (2–5 cm) wide, with conspicuous veins.

Free radicals have been accused of initiating many serious diseases [1–3]. These free radicals drive oxidative stress and transform the pathophysiological condition of the patient by acting on immune system. It has been known that phenolic and flavonoid compounds of the plant extracts are responsible for antioxidant and antibacterial effects [4–6].

Taking all the above concerns into account, we conducted this study to find out more about *A. reticulata* leaves. We studied the antioxidant effects with presence of such phytochemical constituents as equivalent to standards in different extracts, the cytotoxic effect, and hence antitumor effect.

2. Materials and Methods

2.1. Plant Materials

The leaves of *A. reticulata* and *Allium sativum* (bulbs) were collected from regions of Karjat Dist-Raigad, Maharashtra, India in December 2018. Plant materials were authenticated at “The Blatter Herbarium” – St. Xavier’s College, Mumbai.

After identification and authentication of the plant, leaves of the plant were collected for the experimental process. The leaves were shade dried, made into coarse powder and the powdered material was initially defatted with petroleum ether and then subjected to cold maceration process for 72-h using 1:1 mixture of methanol and water as solvent to prepare hydro-alcoholic extract of *Annona reticulata* leave (percentage yield 20.5% *w/w* with respect to dried powder). The extract was filtered and concentrated by rotary evaporator. For the preparation of different fractions method was used [7–9].

The sun dried and powdered leaves (76 g) of *A. reticulata* were successively extracted in a Soxhlet extractor at elevated temperature using 200 mL of distilled n-hexane (40–60) °C which was followed by petroleum ether, methanol, and chloroform. All extracts were filtered individually through filter paper and poured on petri dishes to evaporate the liquid solvents from the extract to get dry extracts. The dry crude extracts were weighed and stored in air-tight container with necessary markings for identification and kept in a refrigerator for future investigations.

2.2. Brine Shrimp Lethality Bioassay

The extracts, fractions and pure isolated compounds were routinely evaluated in a test for lethality to brine shrimp larvae. Toxicities of compounds were tested at 1, 10, 100 and 1000 ppm in 10 mL sea-water solutions with 1% DMSO (*v/v*). Ten, nauplii were used in each test and survivors counted after 24 h. Three replications were used for each concentration. The blank control is conducted with Distilled water. The lethal concentration for 50% mortality after 24 h of exposure, the chronic LC₅₀ was determined using the probit method, as the measure of toxicity of the extract or fractions. LC₅₀ values greater than 1000 ppm for plant extracts were considered inactive.

The brine shrimp lethality assay (BSLA) is a simple and inexpensive bioassay used for testing the efficacy of phytochemical present in the plant extracts. The present study determined that the extent of lethality was directly proportional to the concentration of the extract. After 24 h of observation all the shrimp were survived in the control. Even though, maximum mortalities were observed upto a concentration of 1000 µg/mL and least mortality at 1 µg/mL concentrations. It was observed that in higher concentration of treatment extracts, the shrimps were start dying only after 8 h and after 24 h all the shrimps died. The lethality concentration (LC₅₀) was calculated by using probit analysis (Table 1). The LC₅₀ (median lethal concentration) values were calculated by using the regression line obtained by plotting the concentration against the death percentage on a probit scale.

2.3. Significance of Brine Shrimp Lethality Assay of the Plant

The evolution of the toxic action of plant extracts is indispensable to consider a treatment safe; it enables the definition of the intrinsic toxicity of the plant, and the effects of acute overdose [10], a cheap and general bioassay that appears capable of detecting a spectrum of bioactivity present in crude extract is the brine shrimp lethality test. The lethality of the test sample in a simple zoological organism like the brine shrimp (*Artemia salina*) has been utilised by many researchers and has proven to be a useful tool in screening various chemical compounds found in various bioactivities. In this study, it was observed combined aqueous and alcoholic fractions of *Annona reticulata* and *Allium sativum* extract exhibited the highest brine shrimp cytotoxic activity.

The combined aqueous and alcoholic fractions of leaves of *Annona reticulata* and bulbs extracts of *Allium sativum* exhibited a concentration-dependent cytotoxic activity in brine shrimp and is considered containing active or potent components, brine shrimp lethality assay is inadequate in determining the mechanism of action of the bioactive substances in the plant, but it is useful in

providing a preliminary screen that can be supported by a more specific bioassay, once the active compound has been isolated [11–14].

Table 1. % Mortality of shrimp nauplii after treating with Alcoholic and aqueous extract of *Annona reticulata* and *Allium sativum*.

Plant Methanolic Extracts	Concentration (ppm or µg/mL)	Number of Surviving Nauplii (after 24 h)			Total Number of Nauplii Survivors	% Mortality	LC ₅₀ (µg/mL)	Graph
		T1	T2	T3				
Control (Distilled water)	1	10	10	9	29	96%	372.846	Figure 1
	10	10	9	10	29	96%		
	100	8	10	10	28	93%		
	1000	10	10	10	30	100%		
Standard (Vincristine sulphate)	1	0	0	0	0	100%	0.00	----
	10	0	0	0	0	100%		
	100	0	0	0	0	100%		
	1000	0	0	0	0	100%		
<i>Annona reticulata</i> (Alcoholic)	1	10	10	10	30	0%	24.162	Figure 2
	10	6	8	7	22	73%		
	100	3	2	3	8	27%		
	1000	1	0	0	1	3.3%		
<i>Annona reticulata</i> (Aqueous)	1	10	10	10	30	0%	18.923	Figure 3
	10	8	6	6	20	66.6%		
	100	3	2	5	10	33.3%		
	1000	0	1	0	1	3.3%		
<i>Allium sativum</i> (Alcoholic)	1	5	7	7	19	37%	10.840	Figure 4
	10	4	4	3	11	63%		
	100	0	0	0	0	100%		
	1000	0	0	0	0	100%		
<i>Allium sativum</i> (Aqueous)	1	6	4	5	21	70%	8.180	Figure 5
	10	2	3	3	4	13.3%		
	100	1	1	0	2	6.6%		
	1000	0	0	0	0	100%		
<i>Annona reticulata</i> and <i>Allium sativum</i> (1:1) Alcoholic extracts	1	3	5	4	12	40%	129.257	Figure 6
	10	4	5	5	17	56.6%		
	100	3	2	2	4	13.3%		
	1000	1	1	0	2	0.03%		
<i>Annona reticulata</i> and <i>Allium sativum</i> (1:1) Aqueous extracts	1	3	3	5	12	40%	93.482	Figure 7
	10	5	4	4	14	46.6%		
	100	1	1	3	5	16.6%		
	1000	0	0	0	0	100%		

Cytotoxicity of Control

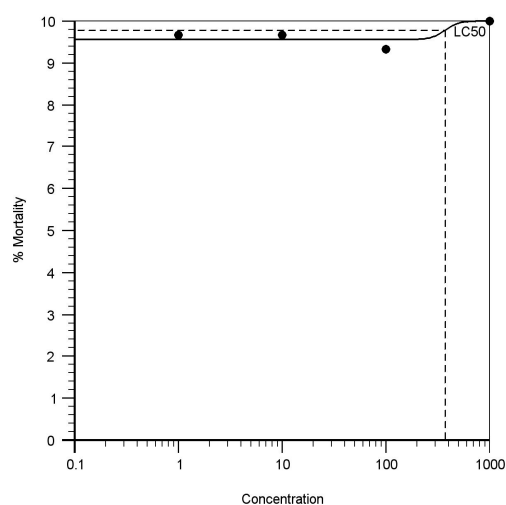


Figure 1. Cytotoxicity of Control.

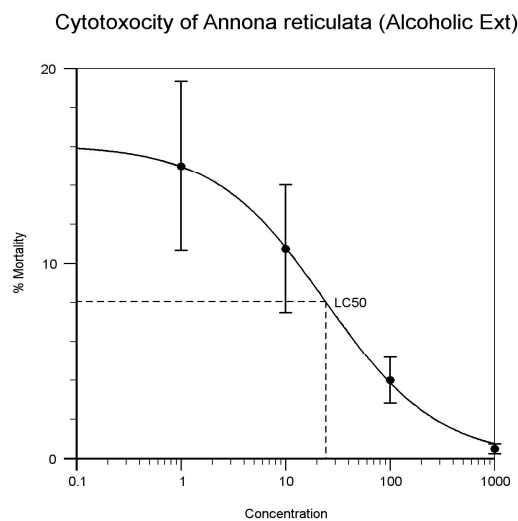


Figure 2. Cytotoxicity of *Annona reticulata*. (Alcoholic Extract).

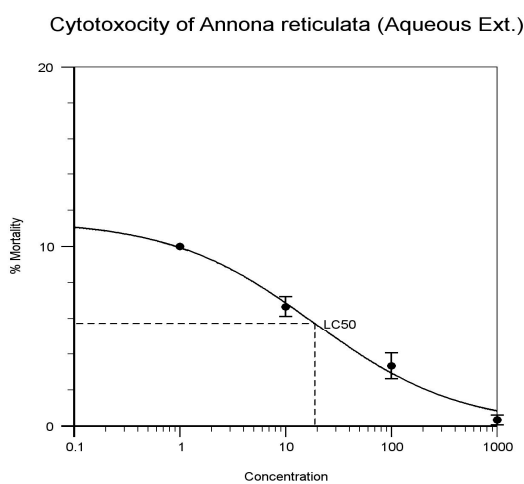


Figure 3. *Annona reticulata*. (Aqueous Extract).

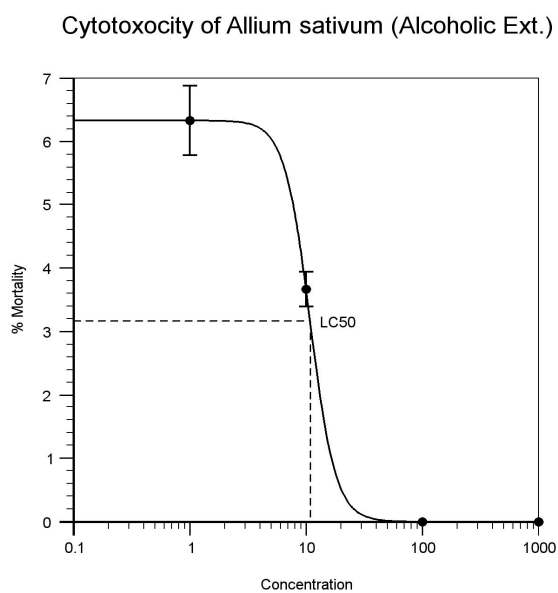


Figure 4. *Allium sativum*. (Alcoholic extract).

Cytotoxicity of *Allium sativum* (Aqueous Ext.)

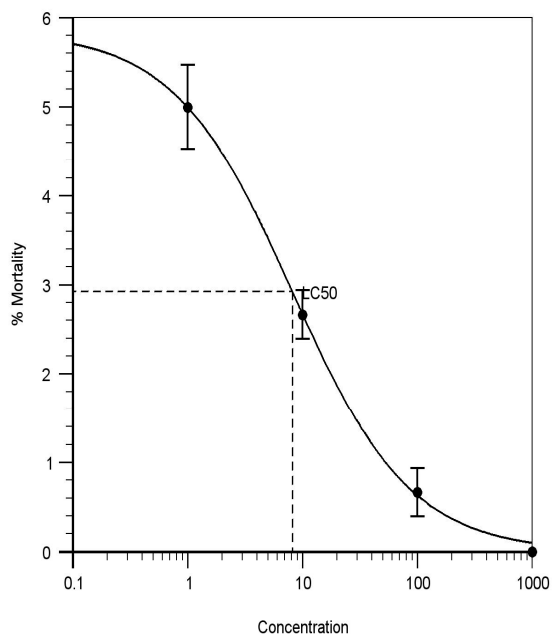


Figure 5. *Allium sativum*. (Aqueous extract).

Cytotoxicity of *A. reticulata* and *A. sativum* Alc.

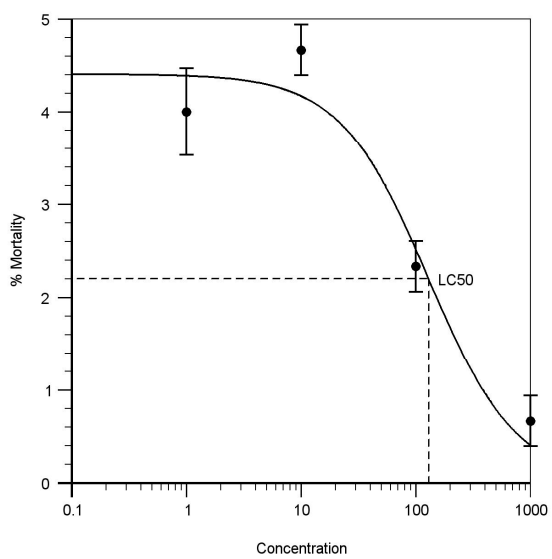


Figure 6. *Annona reticulata* and *Allium sativum* (1:1). Alcoholic extracts.

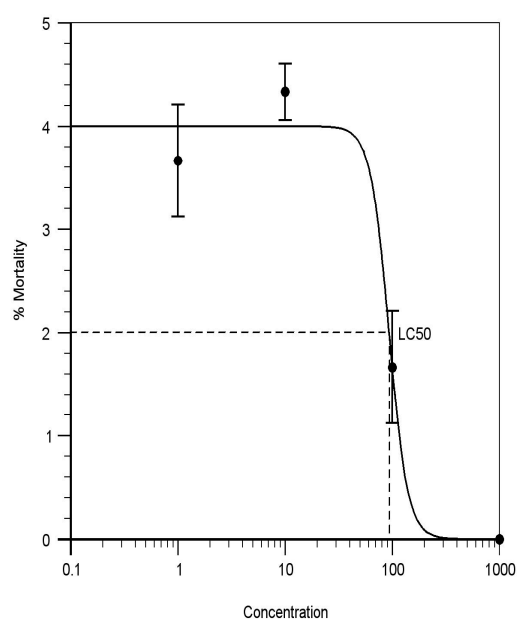
Cytotoxicity of *A. reticulate* and *A. sativum* Aq.

Figure 7. *Annona reticulate* and *Allium sativum* (1:1). Aqueous extracts.

3. Result and Discussion

The result on the lethality of Alcoholic and aqueous extract of *Annona reticulate* on brine shrimps is in agreement with other studies where its LC_{50} values are 24.162 $\mu\text{g}/\text{mL}$ and 18.923 $\mu\text{g}/\text{mL}$ (as indicated in Figures 2 and 3). Alcoholic and aqueous extract bulbs of *Allium sativum* recorded LC_{50} values of 10.840 and 8.180 mg/mL (as indicated in Figures 4 and 5) against brine shrimps. The presence of alkaloids, tannins, and flavonoids could be accounted for its cytotoxic properties. In the other hand, studies have shown that the leaf extracts of Alcoholic and aqueous extract of *Annona reticulata* and bulbs of *Allium sativum* extracts exhibited cumulative activity when they were combined and compared. Thus, the leaf extracts of Alcoholic and aqueous extract of *Annona reticulata* and bulbs of *Allium sativum* exhibited increase in activity support its use in traditional medicine.

4. Conclusions

The leaf extracts of *Alcoholic and aqueous extract of Annona reticulata and bulbs of Allium sativum* exhibited cytotoxic activity against the brine shrimp and considered as containing active or potent components. This is because their LC_{50} values are less than 1000 ppm or $\mu\text{g}/\text{mL}$. *Annona reticulate* and *Allium sativum* (1:1) Alcoholic extracts shows LC_{50} values 129.257 $\mu\text{g}/\text{mL}$ and *Annona reticulate* and *allium sativum* (1:1) Aqueous extracts shows LC_{50} values 93.482 $\mu\text{g}/\text{mL}$ (as indicated in Figures 6 and 7). The *Alcoholic and aqueous extract of Annona reticulata and bulbs of Allium sativum* when combined in (1:1) proportions, the higher LC_{50} values shows better cytotoxicity than single extracts. The ethnopharmacological activities of these plant species are due to the different bioactive compounds present in these plants. Although, BSLA is inadequate in determining the mechanism of action of the bioactive substances in the plant, it is very useful by providing a preliminary screen that can be supported by a more specific bioassay, once the active compound has been isolated. Thus, some useful drugs of therapeutic importance may develop out of the research work.

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References

1. Malorni, W.; Rivabene, R.; Lucia, B.M.; Ferrara, R.; Mazzone, A.M.; Cauda, R.; Paganelli, R. The role of oxidative imbalance in progression to AIDS effect of the thiol supplier N-acetyl cysteine. *AIDS Res. Hum. Retrovir.* **1998**, *14*, 1589–1596.
2. Robert, A.; Meunier, B. Is alkylation the main mechanism of action of the antimalarial drug artemisinin? *Chem. Soc. Rev. Artic.* **1998**, *27*, 273–274.
3. Shah, P.M. The need for new therapeutic agents: What is in the pipeline? *Clin. Microbiol. Infect.* **2005**, *11*, 36–42.
4. Da-Silva, J.F.M.; De-Souza, M.C.; Matta, S.R.; De-Andrade, M.R.; Vidal, F.V.N. Correlation analysis between phenolic levels of Brazilian propolis extracts and their antimicrobial and antioxidant activities. *Food Chem.* **2006**, *99*, 431–435.
5. Majhenic, L.; Skerget, M.; Knez, Z. Antioxidant and antimicrobial activity of guarana seed extracts. *Food Chem.* **2007**, *104*, 1258–1268.
6. Pereira, J.A.; Oliveira, I.; Sousa, A.; Valentao, P.; Andrade, P.B. Walnut (*Juglans regia* L.) leaves: Phenolic compounds, antibacterial activity and antioxidant potential of different cultivars. *Food Chem. Toxicol.* **2007**, *45*, 2287–2295.
7. Rout Soumya, P.; Kar Durga, M.; Mohapatra Santosh, B.; Swain Sharada, P. Anti-hyperglycemic effect *Annona reticulata* L. Leaves on experimental diabetic rat model. *Asian J. Pharm. Clin. Res.* **2013**, *6*, 56–60.
8. Rout, S.P.; Kar, D.M. Identification of chemical compounds present in different fractions of *Annona reticulata* L. Leaf by using GC-MS. *Nat. Prod. Res.* **2014**, *28*, 1786–1788.
9. Rout, S.P.; Kar, D.M.; Maharana, L. Anti-Hyperglycemic Effect of different fractions of *Annona Reticulata* leaf. *Asian J. Pharm. Clin. Res.* **2016**, *9*, 256–262. doi:10.22159/ajpcr.2016.v9s2.13710.
10. Padjama, R.; Arun, P.C.; Prashanth, D.; Deepak, M.; Amit, A.; Anajna, M. Brine shrimp lethality bioassay of Indian medicinal plants. *Fitoterapia* **2002**, *73*, 508–510.
11. Abhilasha, S.; Kuntal, K. Analysis of phytochemical constituents and pharmacological properties of *Abrus precatorius* L. *Int. J. Pharm. Biol. Sci.* **2013**, *4*, 91–96.
12. Adelowotan, O.; Aibinu, I.; Aednipekun, E.; Odugbemi, T. The in vitro antimicrobial activity of *Abrus Precatorius* (L) fabaceae extract on some clinical pathogens. *Niger. Postgrad. Med. J.* **2008**, *15*, 32–37.
13. Prashith Kekuda, T.R.; Vinayaka, K.S.; Soumya, K.V.; Ashwini, S.K.; Kiran, R. Antibacterial and antifungal activity of methanolic extract of *Abrus pulchellus* wall and *Abrus precatorius* Linn—a comparative study. *Int. J. Toxicol. Pharm. Res.* **2010**, *2*, 26–29.
14. Adedapo, A.A.; Omoloye, O.A.; Ohore, O.G. Studies on the toxicity of an aqueous extract of the leaves of *Abrus precatorius* in rats. *Onderstepoort J. Vet. Res.* **2007**, *74*, 31–36.



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