

Review

Adsorption, Surface Viscoelasticity, and Foaming Properties of Silk Fibroin at the Air/Water Interface

Xiuying Qiao ^{1,*}, Reinhard Miller ² , Emanuel Schneck ² and Kang Sun ¹

¹ State Key Laboratory of Metal Matrix Composites, School of Materials Science and Engineering, Shanghai Jiao Tong University, Shanghai 200240, China; ksun@sjtu.edu.cn

² Department of Physics, Technical University of Darmstadt, 64277 Darmstadt, Germany; miller@fkp.tu-darmstadt.de (R.M.); emanuel.schneck@pkm.tu-darmstadt.de (E.S.)

* Correspondence: xyqiao@sjtu.edu.cn

Abstract: Like other proteins, the natural silk fibroin (SF) extracted from domesticated silkworms can adsorb at the air/water interface and stabilize foam due to its amphiphilic character and surface activity. At the interface, the adsorbed SF molecules experience structural reorganization and form water-insoluble viscoelastic films, which protect foam bubbles from coalescence and rupture. The solution conditions, such as protein concentration, pH, and additives, have significant influences on the molecular adsorption, layer thickness, interfacial mechanical strength, and, thus, on the foaming properties of SF. The understanding of the relationship between the interfacial adsorption, surface viscoelasticity, and foaming properties of SF is very important for the design, preparation, and application of SF foams in different fields.

Keywords: silk fibroin; adsorption; interface; foam



Citation: Qiao, X.; Miller, R.; Schneck, E.; Sun, K. Adsorption, Surface Viscoelasticity, and Foaming Properties of Silk Fibroin at the Air/Water Interface. *Colloids Interfaces* **2022**, *6*, 40. <https://doi.org/10.3390/colloids6030040>

Academic Editors: Pierre Bauduin and Seong H. Kim

Received: 5 June 2022

Accepted: 17 July 2022

Published: 19 July 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Foam is a gas-in-liquid dispersion with gas bubbles separated by thin liquid films. These thin liquid films are thermodynamically unstable and will rupture eventually due to drainage and van der Waals attractions, thus leading to foam collapse. Foam can be stabilized by amphiphilic proteins or low-molecular-weight surfactants, which can adsorb at the air/water interface and protect the gas bubbles against coalescence. Compared with surfactants, proteins usually form thicker adsorbed layers and endow the foam with higher stability, although they adsorb more slowly and result in larger bubble size [1]. Generally, the foaming properties can be quantified in terms of foamability and foam stability. Foamability is defined as the capacity of the continuous phase to entrap gas and depends on the rate of diffusion and interfacial adsorption of surface-active molecules [2]. Foam stability is the ability to retain the gas for a certain period of time, and relates to the thickness, mechanical strength, and molecular interactions of the adsorbed layers [3]. The mechanical resistance of the surface films to shear and dilatational deformations counteracts bubble coalescence caused by film drainage and rupture. Therefore, understanding the relationships between the adsorption, surface viscoelasticity, and foaming properties of proteins at the air/water interface is very important for the design, preparation, and stability control of foams stabilized by proteins.

It is commonly agreed that not only the intrinsic molecular properties but also the solution conditions such as protein concentration, pH, and additives remarkably affect the interfacial adsorption, film formation, and thus the foam stability [4,5]. High protein concentrations can lead to the formation of a multilayer of globular proteins and, thus, enhance the foam stability [6]. At pH near the isoelectric point (pI), proteins usually show poor foaming properties because of their strong hydrophobic interactions in the formed surface films [4,7,8]. Acidic pH can help some proteins adsorb much faster and give a higher elastic modulus by dissociating into small subunits with better flexibility [4,9]. Even

at the same pH, different proteins exhibit different foamability due to the difference in adsorption and unfolding rates [10]. The charge of proteins also plays an important role in their adsorption and foaming properties [11,12], and reduced electrostatic repulsion between molecules at the interface can improve the foam stability [2,13]. Salts with different ions can alter the effective surface charge of proteins by interacting with the amino acids, and then change the conformation and molecular interactions of proteins and the formation and stability of surface films [5]. Increasing ion valence or concentration in a certain range can enhance the protein adsorption and the foam stability, due to the decrease in net surface charge, the promotion of protein–protein interactions and the increase in surface dilatational elasticity [5,11,14]. For example, lysozyme in its native state forms a fragile monolayer film, but the addition of NaNO₃ leads to a thick and strongly viscoelastic film, in which lysozyme transforms its secondary structure from helix to β sheet [15]. Similar to salts, surfactants can also accelerate the interfacial adsorption and enhance the foamability of proteins. However, a competitive adsorption exists between proteins and surfactants, which may result in the decrease in the interfacial modulus and foam stability [16–18]. Different surfactants exhibit different competitive adsorption effects on some proteins [19]. The displacement of β -lactoglobulin (β -LG) by the anionic sodium dodecyl sulphate (SDS) can be described by the nucleation of surfactant domains at the foam bubble surfaces without significant domain growth. In contrast, the displacement of β -LG by nonionic Tween 20 surfactants includes both nucleation and growth of surfactant domains, with the rupture of protein networks and the disappearance of shear elasticity [18,20]. Although Tween 20 decreases the surface dilatational modulus of soy globulins because of competitive adsorption, it accelerates their interfacial adsorption and improves the foamability and foam stability [16,17]. The interfacial interactions between proteins and surfactants are driven not only by their surface activity, but also by the abilities of proteins to form networks [16].

Silk fibroin (SF), a natural protein extracted from domesticated silkworms, has attracted great attention in various biomedical fields due to its unique mechanical properties, excellent biocompatibility, and low immunogenicity [21–23]. As a kind of fibrous structural protein with the components of amino acids glycine, alanine, and serine, SF can arrange into different forms of random coil, α -helix, and crystalline β -sheet [23], and can be processed into different forms of film, gel, membrane, powder, and porous sponge as well [24,25]. Due to the existence of highly repetitive amino acid sequences with alternating hydrophobic and hydrophilic blocks along the molecular chains, SF exhibits an amphiphilic character and can adsorb at interfaces between water and hydrophobic media (such as air or oil) to form stable viscoelastic films [26]. Using a combination of transmission electron microscopy and electron diffraction techniques, it was found that SF at the air/water interface exhibits a silk III conformation, an approximately hexagonal packing of protein molecules in a left-handed threefold helical chain conformation [27].

Like other proteins, SF can also form viscoelastic protective films at the air/water interface and stabilize foam. SF foams were successfully utilized to prepare some special and functional materials for creative investigations and biomedical applications. SF foam can produce single crystals of SF in the metastable silk I polymorph after being dried and sonicated [28]. The injectable SF foams fabricated by using low-pressure nitrous oxide gas or freeze-processing techniques allow SF to support tissue regeneration as a porous bioactive degradable filler [29,30]. It was demonstrated that the freeze-dried microporous SF foams are useful for constructing long-term controlled drug release systems [31,32]. Through a new method of compressing the SF foams, the porous SF membranes can be obtained for tissue scaffolding applications [33]. The production of stable SF foams is very important for their applications. As mentioned above, the foamability and foam stability of SF are closely related to its interfacial adsorption and the mechanical resistance of its interfacial films. In this review, we present a summary about the state of the art on the adsorption, surface viscoelasticity, and foaming properties of SF at the air/water interface with the effects of different solution conditions. The techniques of dynamic surface tension measurements as well as surface dilatational and interfacial shear rheology provide a lot of

important information on the molecular interactions during adsorption, the mechanical strength of adsorbed layers, and the long-term foam stability [1]. The insights gained can provide a basis for the design, preparation, and application of biocompatible SF foams.

2. Preparation and Features of Aqueous Silk Fibroin Solution

Aqueous SF solutions are typically prepared from fresh domestic *Bombyx mori* cocoon shells by sericin extraction, cocoon fiber dissolution, and solution dialysis. They are then diluted with the solutions at different solution conditions to the desired concentration (C_{SF}) [34–37]. The pI of SF, at which the zeta potential vanishes, was determined to be 4.2 [7]. This means that SF should be negatively charged in aqueous solution at neutral state. The average hydrodynamic diameter (D_{hyd}) of SF shows a minimum at pH 7 and a maximum at pH 4 (Figure 1). When the solution pH changes from neutral to acidic values, the neutralization of side chain carboxyl groups via protonation [38] leads to the decrease in electrostatic repulsion and solubility, thus promoting the formation of large SF aggregates. At pH 4 close to the pI, SF exhibits the lowest solubility and the largest aggregates [35].

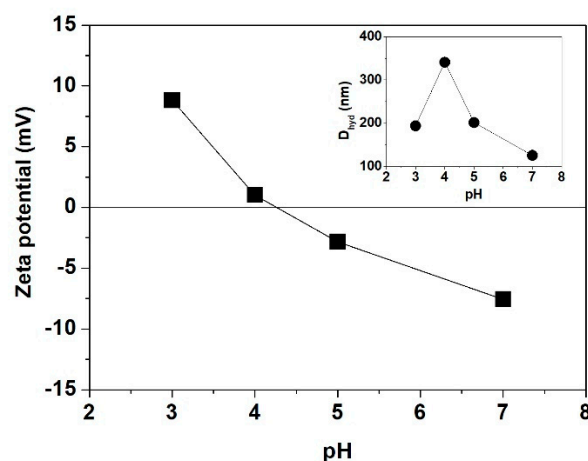


Figure 1. Dependence of the Zeta potential of SF on pH. The inset plot shows the hydrodynamic diameter (D_{hyd}) of SF aggregates at different pH (taken from [35]).

3. The Adsorption of Silk Fibroin at the Air/Water Interface

3.1. Ellipsometry

The thickness of the adsorbed layers formed at the air/water interface can be determined by ellipsometry. The adsorption of proteins usually experiences several steps, including diffusion to the interface, surface adsorption, structural reorganization, and spreading at the surface [39]. During adsorption, the layer thickness of SF gradually increases and then becomes steady when saturation is reached. Clearly, the amphiphilic SF adsorbs to the air/water interface and forms surface layers. The thickness of SF layers is much higher than that of soy protein isolates [40], but slightly lower than that of soy protein fibrils (SPF) [41]. Variations in the equilibrium thickness (δ_e) (Figure 2) arise from a subtle balance between molecular size, shape, surface charge, and surface hydrophobicity [41].

Irrespective of the pH, δ_e is larger at higher C_{SF} . High C_{SF} helps to promote the packing density, but at small or saturated adsorption the effect of C_{SF} weakens [34]. In addition, δ_e is higher at pH 4 than at pH 3 and pH 7, due to the lack of electrostatic repulsion near the pI and the resulting denser accumulation of SF molecules at the interface [35]. δ_e depends not only on the electrostatic repulsion but also on the protein conformation and flexibility, especially at high C_{SF} . At pH 7, the smaller aggregate size and higher molecular flexibility make SF reorganize and pack at the interface more efficiently than at pH 3. When salt is added, δ_e becomes thicker and increases with the salt concentration (C_{salt}) and the ion valence from NaCl to NdCl₃ [36]. SF in aqueous solution is negatively charged at neutral pH, and the counter-ion screening effect of salt significantly reduces the electrostatic repulsion between SF molecules, thus causing a gradual molecular aggregation

in the bulk and at the interface. In addition, the increasing ion valence further enhances the molecular interactions within the layers and, thus, the thickness of adsorbed layers. The incorporated surfactants remarkably enhance the surface activity of SF: δ_e increases much more strongly and attains equilibrium at much lower C_{SF} . Moreover, δ_e exhibits different dependence on the surfactant concentration (C_{Surf}) for different surfactant types within the studied range, increasing with C_{Surf} for $C_{16}TAB$, but exhibiting a non-monotonic behavior for Tween 20 and SDS [37]. Due to electrostatic attraction, the cationic $C_{16}TAB$ and negatively charged SF can form complexes which co-adsorb in a very dense way and maximize the adsorbed amount. However, due to electrostatic repulsion, the anionic SDS can merely fill up SF-free defects and increase δ_e only slightly. The adsorbed layers formed by surfactants are usually thinner than those formed by proteins. Therefore, when C_{Surf} exceeds a critical value, SDS and Tween 20 make the interfacial layers become thinner due to increasing competition in the adsorption layer, the growth of surfactant domains and the breakage of protein networks. The faster decline in δ_e for Tween 20-mixed solutions should be attributed to the higher surface activity and stronger competition of Tween 20 (its critical micelle concentration (~ 0.05 mM [42]) is much lower than that of SDS (~ 8 mM [43]).

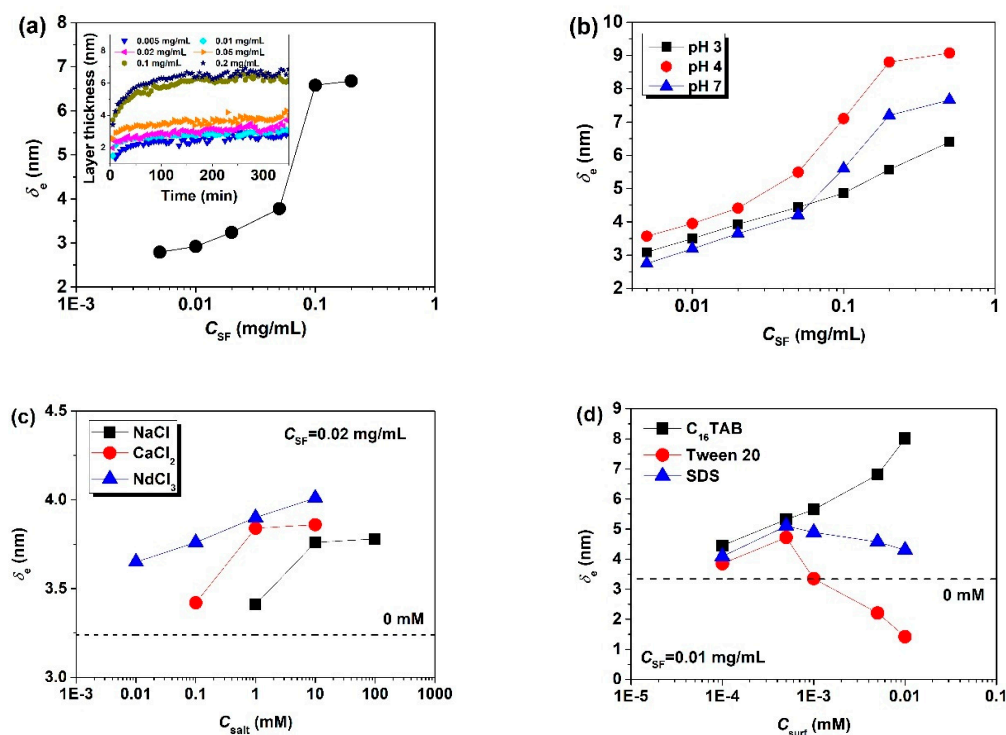


Figure 2. Influences of solution conditions on the equilibrium thickness (δ_e) of the adsorbed layers formed by SF at the air/water interface: (a) influence of C_{SF} ; (b) influence of pH; (c) influence of salts; (d) influence of surfactants (taken from [34–37]).

3.2. Surface Tension

The dynamics of adsorption of SF at the air/water interface can be investigated by measurements of the surface tension (σ) as a function of the adsorption time (t_{ad}) under different solution conditions (Figure 3). When $C_{SF} \leq 0.002$ mg/mL, there is no obvious adsorption of SF and σ shows a plateau value close to that of pure water [34]. At sufficiently low C_{SF} and short t_{ad} , changes in the adsorbed amounts usually do not directly lead to changes in σ , and this time range of negligible σ changes is called induction period (t_{ind}) [44]. When C_{SF} exceeds 0.002 mg/mL, σ begins to decrease immediately until the equilibrium surface tension (σ_{eq}) is reached [34–36]. The initial rapid decrease in σ is related to the adsorption of SF, and the following slow change of σ is more related to the structure formation of adsorption layers. The time for σ to reach equilibrium is around 6 h for SF [34], much longer than that for β -LG [5], gliadin nanoparticles (GNPs) [8], or amaranth

protein isolates [4] at comparable bulk concentrations, indicating that SF needs more time to complete the structural reorganization into gelled β -sheets during the formation of water-insoluble films [45]. SF shows a lower σ_{eq} and a much shorter t_{ind} [35,36] than ovalbumin (OVA) with its more compact and rigid structure [10,46], but a higher σ_{eq} than the highly hydrophobic bovine serum albumin (BSA) and the more deformable wheat protein fractions [46]. Obviously molecular size, structure, and hydrophobicity of proteins significantly influence their adsorption characteristics at interfaces. A lower degree of ordering, higher hydrophobicity, and stronger molecular flexibility can increase the surface activity of proteins and accelerate their adsorption.

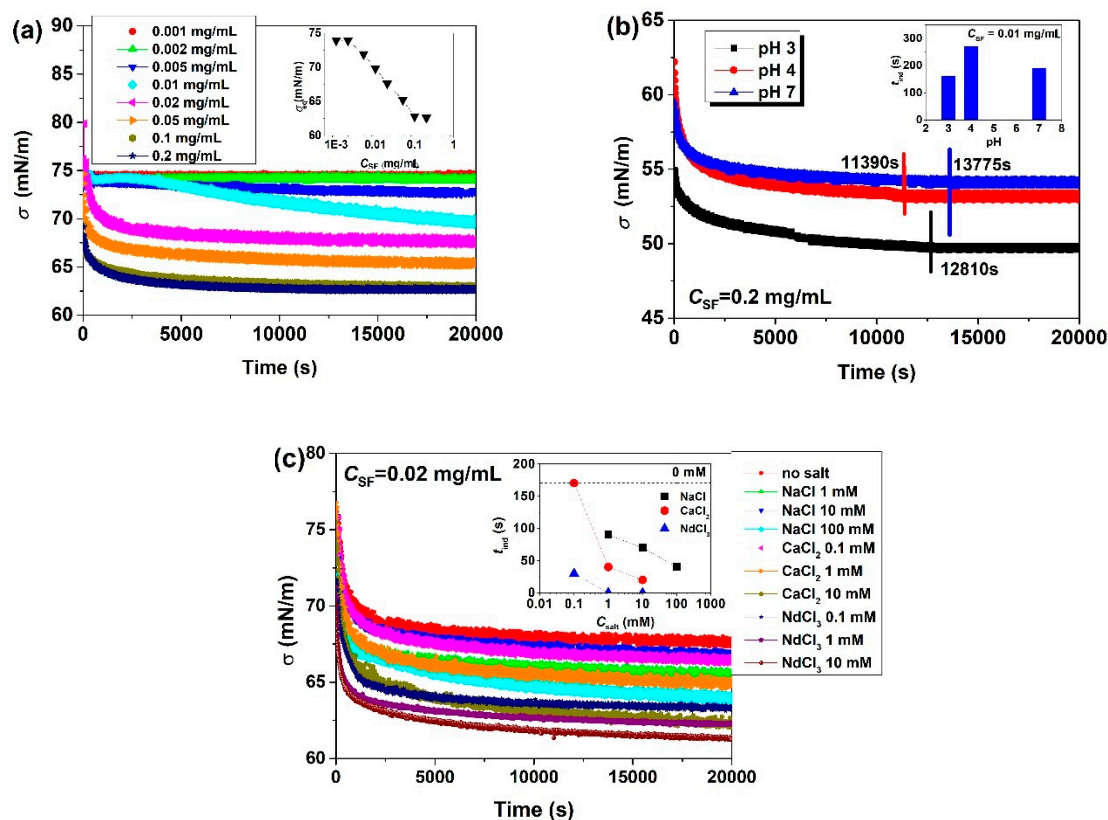


Figure 3. Influences of solution conditions on the surface tension (σ) of SF solutions at the air/water interface: (a) influences of C_{SF} ; (b) influences of pH; (c) influences of salts. The inset plots are the effects of C_{SF} , pH and C_{salt} on the equilibrium surface tension (σ_{eq}) and induction time (t_{ind}) (taken from [34–36]).

Increasing C_{SF} leads to lower σ_{eq} and faster establishment of equilibrium, but σ_{eq} shows no further decrease at $C_{SF} \geq 0.1$ mg/mL when adsorption saturation is reached [34]. Therefore, the surface can only accommodate a limited number of SF molecules. Increasing C_{SF} also causes a decrease in t_{ind} . The initial adsorption of SF at the interface is controlled by diffusion and thus by the aggregate size, while the consecutive steps are controlled by intermolecular interactions at the interface. With that, both molecular flexibility and surface charge are of great importance for this later adsorption stage. σ and t_{ind} show the same ranking of pH 4 > pH 7 > pH 3 [35]. At pH 4, the large aggregate size decelerates the diffusion, thus resulting in a longer t_{ind} and slower decrease in σ . At pH 7, although the size of SF aggregates is smaller than at pH 3, the electrostatic barrier interferes with the interfacial adsorption [47,48] and then causes a slower decrease in σ as compared to pH 3. The addition of salt effectively reduces the electrostatic repulsion between SF molecules and, thus, the energy barrier to SF adsorption. As a consequence, both σ and t_{ind} decrease with increasing C_{salt} and ion valence [36]. Obviously, the surface activity of SF is remarkably

promoted in the presence of salt, and salt addition not only accelerates the diffusion but also enhances the interfacial adsorption of SF molecules.

4. The Surface Viscoelasticity of Silk Fibroin Adsorption Layers at the Air/Water Interface

The mechanical surface properties of adsorbed SF layers at the aqueous solution/air interface are subject to compression/expansion as well as to shear deformations. Both types of surface perturbations provide important information on the mechanical behavior of the SF adsorption layers in the process of foam formation and against foam destabilization. A general introduction into the surface viscoelasticity of adsorbed layers was given in Ref. [49].

4.1. Surface Dilatational Rheology

E' of SF (Figure 4) is much larger than that of β -LG [14], whey globular proteins [50] and soy globulins [51], revealing that the adsorbed SF layers formed by densely packed β -sheets [52] possess stronger mechanical strength than those of other proteins. The dependence of E' on C_{SF} is insensitive to frequency, and E' exhibits a maximum at $C_{SF} = 0.02\sim 0.05$ mg/mL [34]. Additionally, human serum albumin (HSA) [53], β -casein (BCS) [53], and alkaline protease [54] exhibit such non-monotonic-behavior-featuring maxima. High C_{SF} enhances surface adsorption and increases E' . Nevertheless, when C_{SF} exceeds a critical value, interfacial SF molecules reach a jamming state and can no longer reorganize well into ordered elastic networks [55], thus leading to a decrease in E' [34]. At high C_{SF} , E' is larger at pH 7 and decreases with C_{SF} at pH 3 and especially at pH 4 [35]. It seems that neutral pH is advantageous for forming strong interfacial networks [56]. At pH 7, due to electrostatic repulsion, SF molecules adsorb gradually and have more space for the conformational rearrangements required to form strong networks. In contrast, at pH 4, due to the strong hydrophobic interactions, a jamming state is rapidly reached, and the formation of a rigid network does not occur. In the presence of salt, E' remarkably increases with C_{salt} at $C_{SF} \leq 0.01$ mg/mL, but begins to decrease with C_{salt} and ion valence for $C_{SF} \geq 0.02$ mg/mL [36]. Obviously, the added salts strengthen the molecular interactions and increase E' by screening electrostatic repulsion [57]. However, the reduction in electrostatic repulsion concurrently enhances the hydrophobic interactions and causes possible SF aggregation, similar to the results observed at pH near pI. Therefore, with the increase in the ionic strength, the enhanced SF accumulation at the air/water interface leads to thicker adsorbed layers and higher E' at low C_{SF} , but to weaker networks and lower E' at high C_{SF} because of the imperfect molecular reorganization at the crowded interface [55]. To conclude, at high C_{SF} , low ionic strength is beneficial for the SF molecules to form stronger interfacial networks.

4.2. Surface Shear Rheology

Through surface shear rheology measurements, the time dependence of the viscoelastic parameters of storage modulus (G') and loss modulus (G'') can be obtained and utilized to analyze the formation and stability of interfacial layers (Figure 5). The values of G' of SF layers increases first rapidly and then gradually with the adsorption time and cannot attain a steady state even after 24 h [34]. As for SF, the structural reorganization and network formation are remarkably slower than the molecular diffusion and adsorption, thus making G' increase slowly and reach equilibrium with more difficulty at a later stage. However, in the presence of salts and surfactants, the surface activity of SF is significantly enhanced, and G' reaches a plateau rapidly, especially in the presence of salts within 3 h [36,37]. As mentioned above, the presence of salt remarkably decreases the electrostatic repulsion and accelerates the molecular diffusion, adsorption, and structural reorganization of SF at the interface [36]. Due to their low molecular weight, the added surfactant molecules can diffuse, adsorb, and reorganize much faster than the SF molecules at the interface, thus greatly shortening the time to establish adsorption equilibrium [37]. Compared with the proteins of bovine serum albumin (BSA) and bovine gamma globulin (BGG), SF needs

much more time to form stable surface networks and shows almost one order of magnitude higher G' values [16]. Hence, the viscoelastic networks formed by fibrous SF are much stronger than those formed by globular BSA and BGG, and the surface layers of SF can be expected to be more resistant against external perturbations and contribute to a higher foam stability. For all solution conditions, the magnitude of G' is significantly larger than that of G'' , implying that the SF molecules can form elastic surface gels at the interface by physical crosslinks [34,36,37]. With the increase in C_{SF} , the rapid increase in G' at low C_{SF} changes to a slow increase at high C_{SF} , which indicates that high C_{SF} helps to increase both the surface coverage and the mechanical strength of the adsorbed layers [34]. Similar to the equilibrium of E' , the equilibrium value of G' (G_e') gradually decreases with increasing C_{salt} [36]. At high C_{SF} , although added salt enhances the hydrophobic interactions and interfacial adsorption of SF, the more crowded adsorbed SF molecules are unable to establish highly ordered and tight networks, thus resulting in a decrease in G_e' . The competitive adsorption of surfactants also interferes with the structural reorganization and interfacial gel formation of SF [37]. The rapid arrangement of surfactants at the interface, termed as the “orogenic” displacement mechanism [16], will preempt the formation of robust SF networks. The cationic $C_{16}TAB$ remarkably reduces the surface charge and promotes the aggregation of SF, such that it cannot form tightly packed networks, leading to an initial fast decrease in G_e' . However, with the charge neutralization and upon approaching the pI, the cross-linked SF networks become stronger [58] and prevent the adsorption of $C_{16}TAB$, which, therefore, has only a weak impact [37]. When C_{surf} is above 0.001 mM, the decrease in G_e' in the presence of the nonionic Tween 20 exceeds that of $C_{16}TAB$ [37]. The anionic SDS shows a weak ability to replace SF, and the possibly formed SF-SDS complexes make the mixture more hydrophilic and less surface active [59].

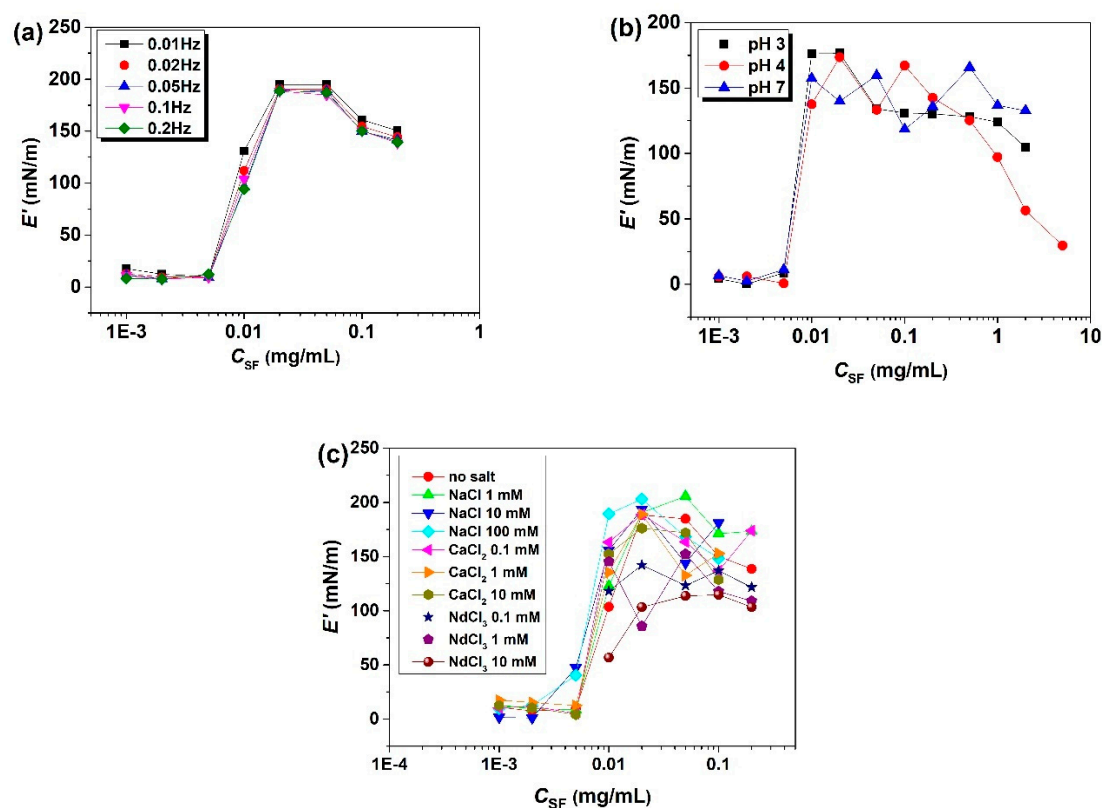


Figure 4. Influences of the solution conditions on the surface dilatational elastic modulus (E') of the SF layers formed at the air/water interface after reaching adsorption equilibrium: (a) influences of C_{SF} ; (b) influences of pH at 0.1 Hz; (c) influences of salts at 0.1 Hz (taken from [34–36]).

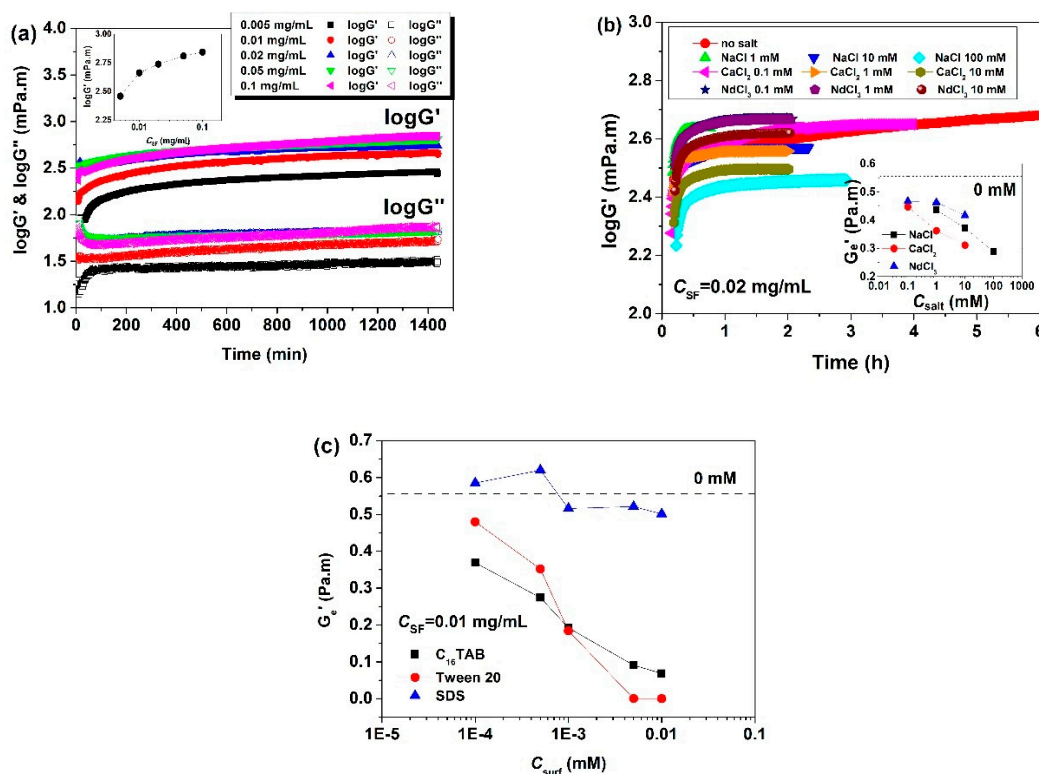


Figure 5. Influences of the solution conditions on the (equilibrium) surface shear moduli of adsorbed SF layers at the air/water interface: (a) influences of C_{SF} ; (b) influences of salts; (c) influences of surfactants. G' in the inset plot of (a) is obtained after 24 h, and the dashed horizontal line in the inset plots of (b) and (c) indicates the value for a pure SF solution without any added surfactant (taken from [34,36,37]).

4.3. Nonlinear Fracture of SF Surface Layers

Strain (γ) sweeps are useful for evaluating the nonlinear fracture of SF layers formed at the air/water interface (Figure 6). During the strain sweeps, interfacial SF layers exhibit a linear behaviour with a constant G' at low γ , and then a nonlinear behaviour is observed with a continuous decrease in G' with increasing γ due to the structural fracture [34]. The degree of structure fracture (η_f) can be calculated via $\eta_f = (G'_0 - G'_\gamma)/G'_0 \times 100\%$, where G'_γ represents the elastic modulus at a certain γ and G'_0 represents the initial constant elastic modulus at low γ [32]. High C_{SF} enhances the elasticity of interfacial layers and their abilities to resist against nonlinear breakage, and η_f decreases with increasing C_{SF} . However, this enhancement effect declines with the increase in γ , and all SF layers are fully broken at $\gamma \geq 15\%$ [34]. In the presence of salts, η_f increases with γ and decreases with C_{salt} , and the adsorbed SF layers are still partially intact even at $\gamma = 25\%$ [36]. Obviously, the added salt indeed improves the interfacial toughness of SF layers, although it decreases the interfacial strength of SF layers in terms of E' . The increase in interfacial toughness may be attributed to the binding of ions to the functional groups of SF and the resultant enhanced molecular interconnection. However, high-valent ions cause an increase in the brittleness of SF layers, and η_f is lower for NaCl than for CaCl₂ and NdCl₃ [36]. As mentioned above, interfaces covered by surfactant molecules are more expandable than those covered by protein molecules, and the incorporated surfactants endow the SF layers with better toughness and markedly decrease η_f . Nevertheless, at high C_{surf} , the excessive interfacial adsorption of surfactant molecules reduces the bond density of SF networks, resulting in a decreasing η_f with C_{surf} [37]. Different from Tween 20 and SDS, C₁₆TAB promotes SF aggregation by charge neutralization, and large SF aggregates cannot pack tightly to form strong interfacial SF networks, which makes η_f exceed that of pure SF when $C_{surf} \geq 0.001$ mM [37]. Among the three surfactants, SDS shows the lowest η_f and

inconspicuous sacrifice of strength, and it appears to be a promising additive to improve the foaming properties of SF solutions.

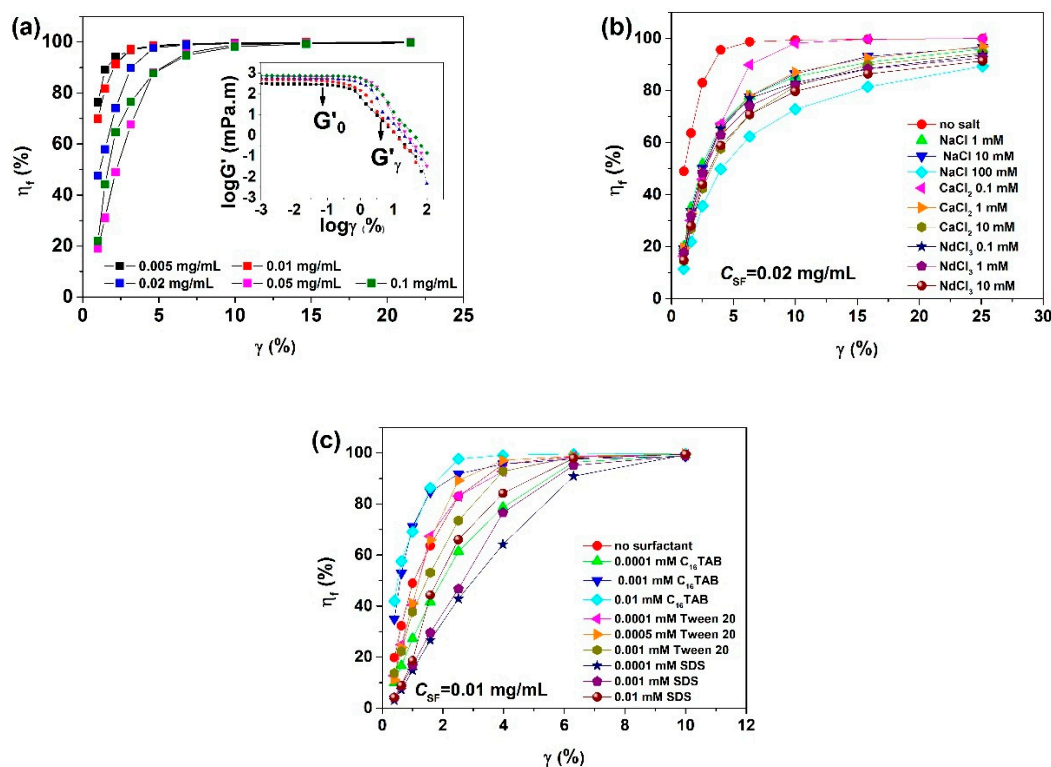


Figure 6. Change of the degree of structure fracture (η_f) with strain (γ) for equilibrium adsorbed SF layers formed at the air/water interface under different solution conditions: (a) influences of C_{SF} ; (b) influences of salts; (c) influences of surfactants. The inset in (a) is the double-logarithmic plot of the interfacial shear storage modulus (G') versus shear strain (γ) (taken from [34,36,37]).

5. The Foaming Properties of Aqueous Silk Fibroin Solutions

5.1. Foamability

SF foams can be easily prepared by the bubbling method through injection of air into different SF solutions through a porous glass frit. The minimum protein concentration required to generate foam with aqueous SF solutions is 0.1 mg/mL [34], 100 times lower than that for rigid globular ovalbumin and glycinin (above 10 mg/mL) [10]. This indicates the superior foamability of SF with its high flexibility and fast molecular adsorption. Foams are thermodynamically unstable systems. It is clear that SF can be a stabilizer for foams although the final “stable” volume fraction is not very large. The surface activity promotes SF to generate the foam, and the adsorbed SF layers with a high G' help maintain the foam structure through mechanical protection against bubble coalescence and rupture. The foamability of SF was quantified in terms of the foaming rate (K_f) by dividing the foam height by the time at a constant air flow (Figure 7). K_f is around 140 mm/min when $C_{SF} < 0.2$ mg/mL, and then decreases to approximately 120 mm/min for $C_{SF} = 1$ mg/mL [34]. Moreover, K_f is lower at pH 4 than at pH 3 and pH 7 [35]. Just as mentioned above, the SF molecules need more time to reach equilibrium when forming surface layers at high C_{SF} , and the larger aggregate size at pH 4 may slow down the diffusion and adsorption of SF molecules at the interface and then the formation of the foam. The incorporated surfactant obviously accelerates the foaming of SF solutions [37], because small surfactant molecules can diffuse and adsorb to the interface much faster than the big protein molecules [3]. However, the cationic C₁₆TAB induces SF aggregation, slows down the SF adsorption, disturbs the SF network formation, and then decreases the foamability. As the concentration of C₁₆TAB

reaches 0.005 mM, the foam decays while being generated, and cannot completely fill up the glass cylinder [37].

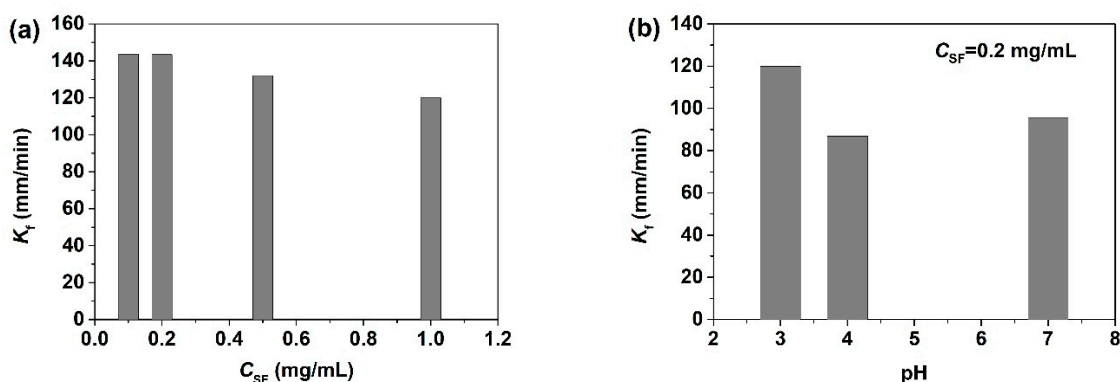


Figure 7. Change of the foaming rate of SF solutions under different solution conditions: (a) the influence of C_{SF} , (b) the influence of pH (taken from [34,35]).

5.2. Foam Stability

After the air injection was stopped, the SF foam started to collapse until reaching a steady state without any obvious changes in volume (Figures 8–10). During collapse, greatly different from GNPs [8] and β -LG fibrils [4], SF foam does not have a flat surface and even collapses from inside, which makes it difficult to quantify the accurate volume of SF foam at different times. In addition, some elastic fiber meshes with filamentous structures appear and adhere to the cylinder wall due to the favorable interactions between SF and glass [34]. This phenomenon implies that even when the gas diffuses out, some SF layers do not break and still retain their strong interconnected network structures with weak mobility. Nevertheless, with the increasing addition of C_{16} TAB and Tween 20, the SF fiber meshes gradually decrease, and even visually disappear when C_{surf} is above 0.005 and 0.01 mM, respectively [37]. The fracture of interfacial SF networks should be attributed to the SF aggregation caused by the electric screening effect of C_{16} TAB and the displacement of SF by growing Tween 20 domains.

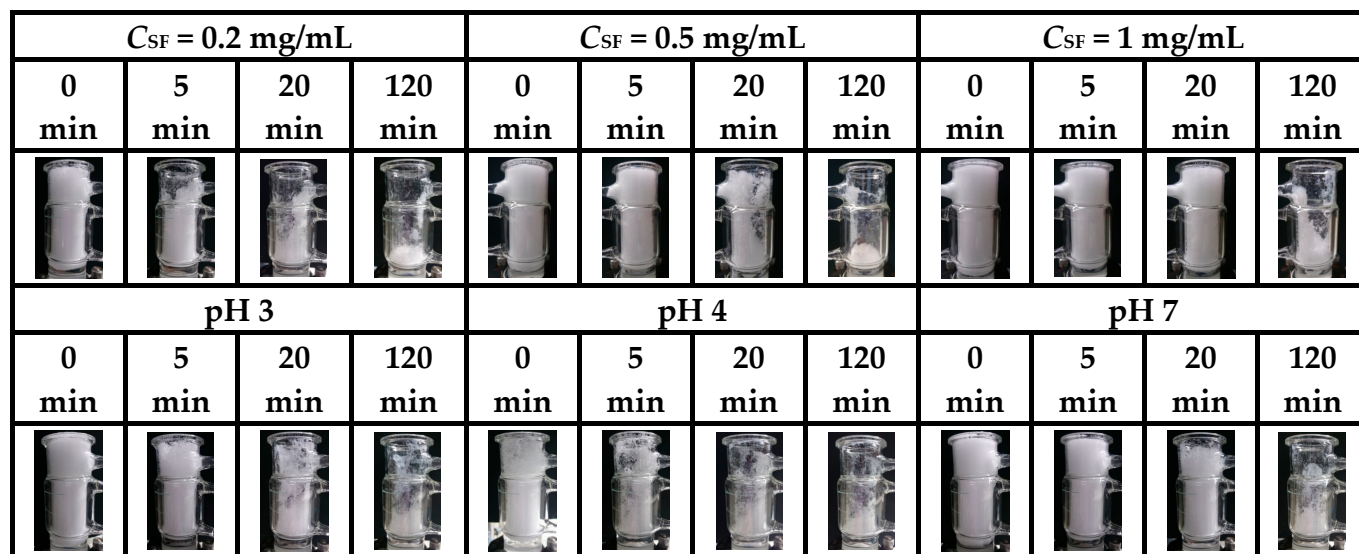


Figure 8. Photographs of foams fabricated by SF aqueous solutions with different C_{SF} and by 0.2 mg/mL SF buffer solutions at different pH taken at different times immediately after foam preparation (taken from [34,35]).

C_{salt}	NaCl				CaCl ₂ (mM)				NdCl ₃ (mM)			
	0 min	5 min	20 min	120 min	0 min	5 min	20 min	120 min	0 min	5 min	20 min	120 min
0 mM												
0.1 mM												
1 mM												
10 mM												
100 mM												

Figure 9. Digital photographs of foams fabricated from 0.2 mg/mL SF solutions in presence of different salt types and concentrations taken at various times after the foaming process was terminated (taken from [36]).

C_{surf}	C ₁₆ TAB				Tween 20				SDS			
	0 min	5 min	20 min	120 min	0 min	5 min	20 min	120 min	0 min	5 min	20 min	120 min
0 mM												
0.0001 mM												
0.0005 mM												
0.001 mM												
0.005 mM												
0.01 mM												

Figure 10. Digital photographs of foams generated from 0.2 mg/mL SF solutions in absence and presence of various types and amounts of surfactants taken at different times immediately after foam preparation (taken from [37]).

Increasing C_{SF} helps to stabilize the foam, with slower foam collapse and larger stable foam fraction [34]. The foamability strongly depends on the diffusion and adsorption of

proteins to the interface, while the foam stability strongly depends on the elasticity of the surface films. At higher C_{SF} , although K_f is slightly lower, the intermolecular interactions become stronger and more tightly packed networks form at the interface, which significantly enhance the mechanical strength of the surface films and, thus, the foam stability. The foams prepared at pH 3 and pH 7 exhibit a denser structure and a larger stable volume fraction than those prepared at pH 4 [35], similar to sodium caseinate (Na-cas) showing higher foam stability at neutral pH than at pI [60]. At pH 4, which is near the pI, the lower solubility and enhanced aggregation prevent SF from forming fine, homogeneous, and stable bubbles, while at pH 7, higher solubility, thicker layers, and stronger interfacial elasticity endow SF foams with the best stability with the slowest foam decay and the largest stable volume fraction [35]. This conclusion is proven by the fact that the SF layers show the smallest η_f at pH 7 and the largest η_f at pH 4 under the same γ . The possible increase in the crystallinity of SF at acidic pH may decrease the flexibility of interfacial films [45] and then further decrease the foam stability of SF at pH 3.

In the presence of salt, the foam stability of SF solutions increases due to the screening of surface charge and the promotion of protein–protein interactions (except for NdCl_3 , which is generally found to impede SF foam stability) [5], just as was reported for the effects of Ca^{2+} on β -LG [14] and of Na^+ on glycinin [51]. However, with increasing C_{SF} and ion valence, the distance between the protein-stabilized bubble surfaces is reduced, thus facilitating bubble coalescence and decreasing the foam stability with the loosening and rapid collapse of foam during decay [36]. Moreover, as mentioned before, salt-induced protein aggregation can also interfere with the interfacial network formation and, thereby, further destabilize the foam. E' of SF layers becomes larger in the presence of NaCl and CaCl_2 but smaller in the presence of NdCl_3 , and the strengthening of interfacial elasticity certainly contributes to protect SF foam against bubble coalescence and film rupture [36]. Similarly, the presence of surfactants also improves the foam stability of SF, except for C_{16}TAB at $C_{\text{surf}} \geq 0.0005$ mM. The foam height of SF at the same standing time decreases with increasing C_{surf} for C_{16}TAB , but exhibits a maximum at $C_{\text{surf}} = 0.0005$ mM for Tween 20 and increases with C_{surf} for SDS [37]. In general, surfactants tend to form thin, mobile, and elastic films at the interface, while proteins tend to form thick, immobile, and rigid films. It seems that the expandable surfactant layers can transfer the external stress from the interfacial SF networks to improve the foam stability, especially for SDS [37]. C_{16}TAB leads to a strong SF aggregation due to the charge neutralization effect, and it is difficult for these large SF aggregates to form tight networks, thus resulting in a weak foam stability at high C_{surf} . SDS exhibits a less pronounced SF displacement than Tween 20, and increasing SDS addition does not bring obvious negative effects to the foam stability.

6. Conclusions and Perspectives

Similar to other proteins, the amphiphilic SF can form protective films at the air/water interface to suppress bubble coalescence and stabilize aqueous foams, thus making SF become a novel natural foam stabilizer. The solution conditions, such as protein concentration, pH, and concentrations of added salts or surfactants, have significant influences on the adsorption, surface rheology, and foaming properties of SF solutions.

Higher C_{SF} leads to thicker adsorbed layers, accelerates the interfacial adsorption and increases the viscoelastic moduli at the interface. However, the increase in the dilatational elastic modulus is limited due to the finite surface coverage by SF molecules, and it even decreases when C_{SF} exceeds a critical value. Faster adsorption and foaming are found for pH 3, larger SF aggregates, thicker adsorbed layers, and slower foaming are found for pH 4, and denser and more stable foams can be generated at pH 7. At pH 7, smaller aggregate size and higher molecular flexibility help SF form more ordered and stable viscoelastic networks at the interface, which are beneficial for the interfacial films to resist deformation breakage. Both salts and surfactants can substantially increase the surface activity of SF, but the type and concentration of additives cause great differences in the molecular adsorption, layer thickness, interfacial mechanical strength, and, thus, the

foaming properties of SF solutions. The incorporated salt reduces the electrostatic repulsion between SF molecules, thus accelerating the diffusion, adsorption and reorganization of SF molecules and increasing the thickness and toughness of interfacial layers. Low ionic strength is beneficial for foam stability, while high ionic strength reduces the foam stability by impeding the formation of rigid interfacial networks due to possible SF aggregation. The surfactants improve the toughness of interfacial networks and, thus, the foam stability of SF solutions, especially the anionic surfactant SDS. However, at high C_{surf} , the enhanced competition adsorption of surfactant leads to the destruction of interfacial SF networks, thus resulting in a decrease in the foam stability of SF. The negative effect of nonionic Tween 20 and anionic SDS comes from the displacement of surfactant domains, while that of the cationic C_{16} TAB comes from the SF aggregation caused by the electric screening effect of surfactant.

The understanding of the relationship between the interfacial adsorption, surface viscoelasticity, and foaming properties of SF is very important for the design, preparation, and application of SF foams in different fields. Smaller molecular size and higher surface activity fasten the molecular diffusion and interfacial adsorption, and the greater mechanical strength of adsorbed layers helps to stabilize the foam by resisting against external deformation breakage and bubble rupture and coalescence. Until now, the molecular diffusion and adsorption and the interfacial structure, morphology, and viscoelasticity of SF at the air/water interface have been less reported. The microstructural rearrangements and interfacial morphology changes of SF layers under different solution conditions need to be further explored with the help of advanced characterization techniques. The influences of temperature and other additives on the adsorption, surface rheology, and foaming properties of SF need to be further studied in future work. In addition, the molecular interactions between SF and additives and their mechanisms also need to be deeply investigated and described. There may be a global optimum in the foamability with regard to all the parameters including SF concentration, salt type/concentration, pH, and surfactant type/concentration.

Author Contributions: The manuscript was written through the contributions of all authors. X.Q.: Conceptualization, Methodology, Investigation, Writing—original draft, Writing—review and editing. R.M.: Conceptualization, Methodology, Writing—review and editing. E.S.: Project administration, Resources, Writing—review and editing. K.S.: Writing—review and editing. All authors have read and agreed to the published version of the manuscript.

Funding: This research was supported by the Alexander von Humboldt Foundation (grant number: CHN/1150450).

Institutional Review Board Statement: Not available.

Informed Consent Statement: Not available.

Data Availability Statement: Not available.

Acknowledgments: Reproduced from Ref. [35] with permission from the Royal Society of Chemistry. Reproduced from Refs. [34,36,37] with permission from Elsevier.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Narsimhan, G.; Xiang, N. Role of proteins on formation, drainage, and stability of liquid food foams. *Annu. Rev. Food Sci. Technol.* **2018**, *9*, 45–63. [[CrossRef](#)] [[PubMed](#)]
2. Lech, F.J.; Steltenpool, P.; Meinders, M.B.J.; Sforza, S.; Gruppen, H.; Wierenga, P.A. Identifying changes in chemical, interfacial and foam properties of β -lactoglobulin-sodium dodecyl sulphate mixtures. *Colloid Surf. A Physicochem. Eng. Asp.* **2014**, *462*, 34–44. [[CrossRef](#)]
3. Zhang, H.H.; Xu, G.Y.; Liu, T.; Xu, L.; Zhou, Y.W. Foam and interfacial properties of Tween 20-bovine serum albumin systems. *Colloid Surf. A Physicochem. Eng. Asp.* **2013**, *416*, 23–31. [[CrossRef](#)]
4. Peng, D.F.; Yang, J.C.; Li, J.; Tang, C.; Li, B. Foams stabilized by β -Lactoglobulin amyloid fibrils: Effect of pH. *J. Agric. Food Chem.* **2017**, *65*, 10658–10665. [[CrossRef](#)]

5. Dombrowski, J.; Gschwendtner, M.; Saalfeld, D.; Kulozik, U. Salt-dependent interaction behavior of β -Lactoglobulin molecules in relation, to their surface and foaming properties. *Colloid Surf. A Physicochem. Eng. Asp.* **2018**, *558*, 455–462. [[CrossRef](#)]
6. Damodaran, S. Functional properties. In *Food Proteins: Properties and Characterization*; VCH Publications: New York, NY, USA, 1996; pp. 167–234.
7. Wilde, P.J. Interfaces: Their role in foam and emulsion behaviour. *Curr. Opin. Colloid Interface Sci.* **2000**, *5*, 176–181. [[CrossRef](#)]
8. Brüchner-Gühmann, M.; Heiden-Hecht, T.; Sözer, N.; Drusch, S. Foaming characteristics of oat protein and modification by partial hydrolysis. *Eur. Food Res. Technol.* **2018**, *244*, 2095–2106. [[CrossRef](#)]
9. Martin, A.H.; Bos, M.A.; van Vliet, T. Interfacial rheology properties and conformational aspects of soy glycinin at the air/water interface. *Food Hydrocoll.* **2002**, *16*, 63–71. [[CrossRef](#)]
10. Martin, A.H.; Grolle, K.; Bos, M.A.; Stuart, M.A.C.; van Vliet, T. Network forming properties of various proteins adsorbed at the air/water interface in relation to foam stability. *J. Colloid Interf. Sci.* **2002**, *254*, 175–183. [[CrossRef](#)]
11. Dombrowski, J.; Gschwendtner, M.; Kulozik, U. Evaluation of structural characteristics determining surface and foaming properties of β -lactoglobulin aggregates. *Colloids Surf. A Physicochem. Eng. Asp.* **2017**, *516*, 286–295. [[CrossRef](#)]
12. Dombrowski, J.; Johler, F.; Warncke, M.; Kulozik, U. Correlation between bulk characteristics of aggregated β -lactoglobulin and its surface and foaming properties. *Food Hydrocoll.* **2016**, *61*, 318–328. [[CrossRef](#)]
13. Engelhardt, K.; Lexis, M.; Gochev, G.; Konnerth, C.; Miller, R.; Willenbacher, N.; Peukert, W.; Braunschweig, B. pH effects on the molecular structure of β -lactoglobulin modified air-water interfaces and its impact on foam rheology. *Langmuir* **2013**, *29*, 11646–11655. [[CrossRef](#)] [[PubMed](#)]
14. Braunschweig, B.; Schulze-Zachau, F.; Nagel, E.; Engelhardt, K.; Stoyanov, S.; Gochev, G.; Khristov, K.; Mileva, E.; Exerowa, D.; Miller, R.; et al. Specific effects of Ca^{2+} ions and molecular structure of β -lactoglobulin interfacial layers that drive macroscopic foam stability. *Soft Matter* **2016**, *12*, 5995–6004. [[CrossRef](#)] [[PubMed](#)]
15. Jaganathan, M.; Dhathathreyan, A.; Selvaraju, C.; Miller, R. Jones-Ray effect on the organization of lysozyme in the presence of NaNO_3 at an air/water interface: Is it a cause or consequence? *RSC Adv.* **2015**, *5*, 100638–100645. [[CrossRef](#)]
16. Ruíz-Henestrosa, V.P.; Sánchez, C.C.; Patino, J.M.R. Adsorption and foaming characteristics of soy globulins and Tween 20 mixed systems. *Ind. Eng. Chem. Res.* **2008**, *47*, 2876–2885. [[CrossRef](#)]
17. Ruíz-Henestrosa, V.P.; Sánchez, C.C.; Patino, J.M.R. Formulation engineering can improve the interfacial and foaming properties of soy globulins. *J. Agric. Food Chem.* **2007**, *55*, 6339–6348. [[CrossRef](#)]
18. Gunning, P.A.; Mackie, A.R.; Gunning, A.P.; Woodward, N.C.; Wilde, P.J.; Morris, V.J. Effect of surfactant type on surfactant-protein interactions at the air-water interface. *Biomacromolecules* **2004**, *5*, 984–991. [[CrossRef](#)]
19. Alahverdijeva, V.S.; Grigoriev, D.O.; Fainerman, V.B.; Aksenenko, E.V.; Miller, R.; Möhwald, H. Competitive adsorption from mixed hen egg-white lysozyme/surfactant solutions at the air-water interface studied by tensiometry, ellipsometry, and surface dilational rheology. *J. Phys. Chem. B* **2008**, *112*, 2136–2143. [[CrossRef](#)]
20. Petkov, J.T.; Gurkov, T.D.; Campbell, B.E.; Borwankar, R.P. Dilatational and shear elasticity of gel-like protein layers on air/water interface. *Langmuir* **2000**, *16*, 3703–3711. [[CrossRef](#)]
21. Zhang, Q.; Wang, N.; Hu, R.Q.; Pi, Y.P.; Feng, J.Q.; Wang, H.; Zhuang, Y.; Xu, W.L.; Yang, H.J. Wet spinning of betilla striata polysaccharide/silk fibroin hybrid fibers. *Mater. Lett.* **2015**, *161*, 576–579. [[CrossRef](#)]
22. Teimouri, A.; Azadi, M.; Emadi, R.; Lari, J.; Chermahini, A.N. Preparation, characterization, degradation and biocompatibility of different silk fibroin based composite scaffolds prepared by freeze-drying method for tissue engineering application. *Polym. Degrad. Stab.* **2015**, *121*, 18–29. [[CrossRef](#)]
23. Sangkert, S.; Meesane, J.; Kamonmattayakul, S.; Chai, W.L. Modified silk fibroin scaffolds with collagen/decellularized pulp for bone tissue engineering in cleft palate: Morphological structures and biofunctionalities. *Mater. Sci. Eng. C* **2016**, *58*, 1138–1149. [[CrossRef](#)] [[PubMed](#)]
24. Elia, R.; Michelson, C.D.; Perera, A.L.; Brunner, T.F.; Harsono, M.; Leisk, G.G.; Kugel, G.; Kaplan, D.L. Electrodeposited silk coatings for bone implants. *J. Biomed. Mater. Res. B Appl. Biomater.* **2014**, *103*, 1602–1609. [[CrossRef](#)]
25. Zhong, T.Y.; Jiang, Z.J.; Wang, P.; Bie, S.Y.; Zhang, F.; Zuo, B.Q. Silk fibroin/copolymer composite hydrogels for the controlled and sustained release of hydrophobic/hydrophilic drugs. *Int. J. Pharm.* **2015**, *494*, 264–270. [[CrossRef](#)] [[PubMed](#)]
26. Yang, Y.H.; Dicko, C.; Bain, C.D.; Gong, Z.G.; Jacobs, R.M.J.; Shao, Z.Z.; Terry, A.E.; Vollrath, F. Behavior of silk protein at the air-water interface. *Soft Matter* **2012**, *8*, 9705–9712. [[CrossRef](#)]
27. Valluzzi, R.; Gido, S.P.; Muller, W.; Kaplan, D.L. Orientation of silk III at the air-water interface. *Int. J. Biol. Macromol.* **1999**, *24*, 237–242. [[CrossRef](#)]
28. He, S.J.; Valluzzi, R.; Gido, S.P. Silk Structure in Bombyx Mori Silk Foams. *Int. J. Biol. Macromol.* **1999**, *24*, 187–195. [[CrossRef](#)]
29. Maniglio, D.; Bonani, W.; Migliaresi, C.; Motta, A. Silk fibroin porous scaffolds by N_2O foaming. *J. Biomat. Sci. Polym. Ed.* **2018**, *29*, 491–506. [[CrossRef](#)]
30. Bellas, E.; Lo, T.J.; Fournier, E.P.; Brown, J.E.; Abbott, R.D.; Gil, E.S.; Marra, K.G.; Rubin, J.P.; Leisk, G.G.; Kaplan, D.L. Injectable silk foams for soft tissue regeneration. *Adv. Healthc. Mater.* **2015**, *4*, 452–459. [[CrossRef](#)]
31. Qin, K.; Pereira, R.F.P.; Coradin, T.; Bermudez, V.; Fernandes, F.M. Biomimetic silk macroporous materials for drug delivery obtained via ice-templating. *ACS Appl. Bio Mater.* **2022**, *5*, 2556–2566. [[CrossRef](#)]
32. Chambre, L.; Parker, R.N.; Allardyce, B.J.; Valente, F.; Rajkhowa, R.; Dilley, R.J.; Wang, X.; Kaplan, D.L. Tunable biodegradable silk-based memory foams with controlled release of antibiotics. *ACS Appl. Bio Mater.* **2020**, *3*, 2466–2472. [[CrossRef](#)] [[PubMed](#)]

33. Nogueira, G.M.; Weska, R.F.; Vieira, W.C., Jr.; Polakiewicz, B.; Rodas, A.C.D.; Higa, O.Z.; Beppu, M.M. A new method to prepare porous silk fibroin membranes suitable for tissue scaffolding applications. *J. Appl. Polym. Sci.* **2009**, *114*, 617–623. [[CrossRef](#)]
34. Qiao, X.Y.; Miller, R.; Schneck, E.; Sun, K. Foaming properties and the dynamics of adsorption and surface rheology of silk fibroin at the air/water interface. *Colloid Surf. A Physicochem. Eng. Asp.* **2020**, *591*, 124553. [[CrossRef](#)]
35. Qiao, X.Y.; Miller, R.; Schneck, E.; Sun, K. Influence of pH on the surface and foaming properties of aqueous silk fibroin solutions. *Soft Matter* **2020**, *16*, 3695–3704. [[CrossRef](#)]
36. Qiao, X.Y.; Miller, R.; Schneck, E.; Sun, K. Influence of salt addition on the surface and foaming properties of silk fibroin. *Colloid Surf. A Physicochem. Eng. Asp.* **2021**, *609*, 125621. [[CrossRef](#)]
37. Qiao, X.Y.; Miller, R.; Schneck, E.; Sun, K. Influence of surfactant charge and concentration on the surface and foaming properties of biocompatible silk fibroin. *Mater. Chem. Phys.* **2022**, *281*, 125920. [[CrossRef](#)]
38. Matsumoto, A.; Chen, J.S.; Collette, A.L.; Kim, U.K.; Altman, G.H.; Cebe, P.; Kaplan, D.L. Mechanisms of silk fibroin sol-gel transitions. *J. Phys. Chem. B* **2006**, *110*, 21630–21638. [[CrossRef](#)] [[PubMed](#)]
39. Mitropoulos, V.; Mütze, A.; Fischer, P. Mechanical properties of protein adsorption layers at the air/water and oil/water interface: A comparison in light of the thermodynamical stability of proteins. *Adv. Colloid Interface Sci.* **2014**, *206*, 195–206. [[CrossRef](#)]
40. Piazza, L.; Auser, N.D.; Gigli, J.; Windhab, E.J.; Fischer, P. Interfacial rheology of soy proteins-high methoxyl pectin films. *Food Hydrocoll.* **2009**, *23*, 2125–2131. [[CrossRef](#)]
41. Wan, Z.L.; Yang, X.Q.; Sagis, L.M.C. Contribution of long fibrils and peptides to surface and foaming behavior of soy protein fibril system. *Langmuir* **2016**, *32*, 8092–8101. [[CrossRef](#)]
42. Mahmood, M.E.; Al-Koofee, D.A.F. Effect of temperature changes on critical micelle concentration for tween series surfactant. *Glob. J. Sci. Front. Res. Chem.* **2013**, *13*, 1–7.
43. Fuguet, E.; Ràfols, C.; Rosés, M.; Bosch, E. Critical micelle concentration of surfactants in aqueous buffered and unbuffered systems. *Anal. Chim. Acta* **2005**, *548*, 95–100. [[CrossRef](#)]
44. Miller, R.; Fainerman, V.B.; Aksenenko, E.V.; Leser, M.E.; Michel, M. Dynamic surface tension and adsorption kinetics of β -casein at the solution/air interface. *Langmuir* **2004**, *20*, 771–777. [[CrossRef](#)] [[PubMed](#)]
45. Johnston, E.R.; Miyagi, Y.; Chuah, J.A.; Numata, K.; Serban, M.A. Interplay between silk fibroin's structure and its adhesive properties. *ACS Biomater. Sci. Eng.* **2018**, *4*, 2815–2824. [[CrossRef](#)] [[PubMed](#)]
46. Hill, K.; Horváth-Szancics, E.; Hajós, G.; Kiss, É. Surface and interfacial properties of water-soluble wheat proteins. *Colloid Surf. A Physicochem. Eng. Asp.* **2008**, *319*, 180–187. [[CrossRef](#)]
47. Song, K.B.; Damodaran, S. Influence of electrostatic forces on the adsorption of succinylated beta-lactoglobulin at the air-water interface. *Langmuir* **1991**, *7*, 2737–2742. [[CrossRef](#)]
48. Ulaganathan, V.; Retzlaff, I.; Won, J.Y.; Gochev, G.; Gehin-Delval, C.; Leser, M.; Noskov, B.A.; Miller, R. β -Lactoglobulin adsorption layers at the water/air surface: 1. Adsorption kinetics and surface pressure isotherm: Effect of pH and ionic strength. *Colloid Surf. A Physicochem. Eng. Asp.* **2017**, *519*, 153–160. [[CrossRef](#)]
49. Miller, R.; Ferri, J.K.; Javadi, A.; Krägel, J.; Mucic, N.; Wüstneck, R. Rheology of interfacial layers. *Colloid Polym. Sci.* **2010**, *288*, 937–950. [[CrossRef](#)]
50. Mahmoudi, N.; Gaillard, C.; Boué, F.; Axelos, M.A.V.; Riaublanc, A. Self-similar assemblies of globular whey proteins at the air-water interface: Effect of the structure. *J. Colloid Interf. Sci.* **2010**, *345*, 54–63. [[CrossRef](#)]
51. Ruíz-Henestrosa, V.P.; Sánchez, C.C.; Escobar, M.D.M.Y.; Jiménez, J.J.P.; Rodríguez, F.M.; Patino, J.M.R. Interfacial and foaming characteristics of soy globulins as a function of pH and ionic strength. *Colloid Surf. A Physicochem. Eng. Asp.* **2007**, *309*, 202–215. [[CrossRef](#)]
52. Bai, S.; Liu, S.; Zhang, C.; Xu, W.; Lu, Q.; Han, H.; Zhu, H. Controllable transition of silk fibroin nanostructures: An insight into in vitro silk self-assembly process. *Acta Biomater.* **2013**, *9*, 7806–7813. [[CrossRef](#)] [[PubMed](#)]
53. Fainerman, V.B.; Kovalchuk, V.I.; Aksenenko, E.V.; Zinkovych, I.I.; Makievski, A.V.; Nikolenko, M.V.; Miller, R. Dilational viscoelasticity of proteins solutions in dynamic conditions. *Langmuir* **2018**, *34*, 6678–6686. [[CrossRef](#)] [[PubMed](#)]
54. Zhang, J.; Li, Y.Y.; Wang, J.; Zhang, Y. Interfacial behavior of alkaline protease at the air-water and oil-water interfaces. *Appl. Surf. Sci.* **2018**, *433*, 1128–1136. [[CrossRef](#)]
55. Rühls, P.A.; Scheuble, N.; Windhab, E.J.; Fischer, P. Protein adsorption and interfacial rheology interfering in dilational experiment. *Eur. Phys. J. Spec. Top.* **2013**, *222*, 47–60. [[CrossRef](#)]
56. Lech, F.J.; Delahaije, R.J.; Meinders, M.B.J.; Gruppen, H.; Wierenga, P.A. Identification of critical concentrations determining foam ability and stability of β -lactoglobulin. *Food Hydrocoll.* **2016**, *57*, 46–54. [[CrossRef](#)]
57. Lexis, M.; Willenbacher, N. Relating foam and interfacial rheological properties of β -lactoglobulin solutions. *Soft Matter* **2014**, *10*, 9626–9636. [[CrossRef](#)]
58. Roth, S.; Murray, B.S.; Dickinson, E. Interfacial shear rheology of aged and heat-treated β -lactoglobulin films: Displacement by nonionic surfactant. *J. Agric. Food Chem.* **2000**, *48*, 1491–1497. [[CrossRef](#)]
59. Engelhardt, K.; Weichsel, U.; Kraft, E.; Segets, D.; Peukert, W.; Braunschweig, B. Mixed layers of β -lactoglobulin and SDS at air-water interfaces with tunable intermolecular interactions. *J. Phys. Chem. B* **2014**, *118*, 4098–4105. [[CrossRef](#)]
60. Marinova, K.G.; Basheva, E.S.; Nenova, B.; Temelska, M.; Mirarefi, A.Y.; Campbell, B.; Ivanov, I.B. Physico-chemical factors controlling the foamability and foam stability of milk proteins: Sodium caseinate and whey protein concentrates. *Food Hydrocoll.* **2009**, *23*, 1864–1876. [[CrossRef](#)]