



Behtash G. Nezami¹, Bin Tean Teh², Xiaoqi Lin¹ and Ximing J. Yang^{1,*}

- ¹ Department of Pathology, Feinberg School of Medicine, Northwestern University, Chicago, IL 60611, USA; behtash.nezami@northwestern.edu (B.G.N.)
- ² NCCS-VARI Translational Cancer Research Laboratory, National Cancer Centre, Singapore 169610, Singapore
- Correspondence: xyang@northwestern.edu

Abstract: Birt–Hogg–Dubé syndrome (BHDS) is an autosomal dominant disease characterized by skin, lung, and renal manifestations. This syndrome is caused by a germline mutation in the *FLCN* gene, which leads to disruption in multiple downstream pathways. Renal cell carcinomas are one of the serious clinical manifestations of the disease, which usually presents as bilateral and multiple tumors. Morphologically, most of these tumors are classified as hybrid oncocytic tumors. Recent advances in molecular techniques have shed light on the pathogenesis of these renal tumors. In this review, we evaluate and summarize the current knowledge of BHDS, pathologic changes, and its molecular basis with the focus on the renal hybrid oncocytic tumor (HOT), their pathogenesis, and molecular underpinning.

Keywords: Birt-Hogg-Dubé syndrome; RCC; kidney cancer; molecular genetics

1. Introduction

Renal cell carcinomas (RCCs) accounted for 2% of cancers worldwide in 2020, with an incidence of 431,288 per year [1,2]. While most cases of RCC are sporadic, 4–8% are associated with hereditary syndrome involving causative germline mutation [3–5]. One such hereditary syndrome is Birt-Hogg-Dubé syndrome (BHDS, (OMIM #135150), an autosomal dominant disorder characterized by a constellation of clinical features, including fibrofolliculomas, lung cysts associated, primary spontaneous pneumothorax (PSP), and a markedly increased risk of renal cell carcinoma (RCC). BHDS was first suggested in 1975 by German scientists Hornstein and Knickenberg [6]. However this syndrome is named after three Canadian physicians who studied a family with multiple skin fibrofolliculomas, trichodiscomas, and acrochordons on the head and neck and upper torso in 1977 [7]. Initial publications did not describe extracutaneous manifestations, and the association with renal tumors was not established until 1993 almost twenty years later [8]. In 2001, the genetic basis of BHDS was mapped to chromosome 17p11.2, leading to the identification of mutations in the folliculin (FLCN) gene (MIM 607273, formerly called BHD) [9,10]. By 2002, germline mutations in the folliculin gene were identified as the cause of BHDS [11]. These mutations result in a loss of function of the folliculin protein, disrupting multiple cellular metabolic pathways.

BHDS-associated renal cell carcinomas exhibit unusual morphology, and recent research has advanced our understanding of the mechanism underlying BHDS tumorigenesis. This review explores these discoveries, providing a comprehensive overview of BHDS and discusses some unsolved questions as well as our experience. We also examine the clinical implications and potential therapeutic avenues for this group of renal cell carcinomas.



Citation: Nezami, B.G.; Teh, B.T.; Lin, X.; Yang, X.J. Molecular Pathogenesis of Renal Neoplasms in Patients with Birt–Hogg–Dubé Syndrome. *J. Mol. Pathol.* 2024, *5*, 478–496. https://doi.org/10.3390/jmp5040032

Academic Editor: Pasquale Pisapia

Received: 27 August 2024 Revised: 17 September 2024 Accepted: 25 October 2024 Published: 30 October 2024



Copyright: © 2024 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/).

2. BHDS Diagnostic Criteria

The exact prevalence of BHDS remains uncertain, with estimates ranging from 0.5 to 5 per million [12,13]. However, the condition is likely underdiagnosed, and some studies suggest a genetic predisposition as high as 1 in 3000 [14,15].

BHDS exhibits variable expressivity and incomplete penetrance, meaning that clinical presentation of BHDS can differ widely among individuals. Factors influencing expressivity and penetrance are not fully understood but are speculated to include age, race and geographic location, genetic modifiers, environmental factors, and the type of somatic mutations [16,17].

Pulmonary manifestations, such as lung cysts and primary spontaneous pneumothorax due to the rupture of the lung cysts, often precede the development of renal tumors in BHDS patients [18–21]. In some cases, lung cysts and PSP may be the only phenotypic manifestations. Despite this, patients are frequently referred for genetic examination due to the presence of multiple skin tumors.

To address the variability in BHDS manifestation, the European BHD consortium established screening and surveillance guidelines in 2009, incorporating a set of major and minor criteria. The major criterion includes having at least five fibrofolliculomas/trichodiscomas with at least one histologically confirmed and identification of a pathogenic germline variant in the FLCN gene. Minor criteria involve the presence of multiple lung cysts, RCC diagnosed before age 50, multiple or bilateral RCC, hybrid RCC, and having a first-degree relative with BHD. Diagnosis requires meeting either one major criterion or two minor criteria [22]. This criteria was later adopted by European Reference Network for patients with a rare genetic tumor risk syndrome (ERN GENTURIS) in 2024, with some modifications to address the increasing use of genetic testing as a first-line diagnostic test [23]. The risk of renal tumors is higher in BHDS patients with a family history of kidney tumors. However, these guidelines have been mostly replaced by molecular genetic testing to detect FLCN gene mutations. Genetic testing is the definitive diagnostic tool for BHDS. Methods such as single gene polymerase chain reaction (PCR)-based testing, Sanger sequencing, and Multiple Ligation-dependent Probe Amplification as well as advanced sequencing techniques, like targeted sequencing, whole exome sequencing (WES), and whole genome sequencing (WGS), are employed to identify pathogenic or likely pathogenic variants of the FLCN gene.

3. FLCN Gene and BHDS Pathogenesis

3.1. FLCN Gene Structure and Normal Expression

The *FLCN* gene (OMIM# 607273) is composed of 14 exons, of which 11 are coding regions encoding the 579-amino acid, 64-kDa protein folliculin. *FLCN* mRNA is expressed in various healthy tissues, including the skin and its appendages, the distal nephron of the kidney, lung stromal cells, and type 1 pneumocytes, and secretory tissues, such as the epithelial cells of the breast, acinar cells of the pancreas, and serous glands of the parotid. There is no expression of *FLCN* in the colon mucinous glands or epithelium [24,25].

3.2. FLCN Gene Mutations

FLCN mutations are predominantly truncating and include duplications (46.4%), deletions (29%), substitutions (7.1), insertions (0.7%), insertion/deletion (0.3%), long genomic deletions (4%), and splice site deletions (12.5%) [26,27]. These mutations lead to early decay or loss of function of the folliculin protein [11,17,20]. Recurrent hotspot mutations account for approximately half of the cases in most populations. Certain regions of the gene, such as hypermutable C8 tract of exon 11, are more prone to mutations due to DNA polymerase slippage during replication. Specific genotypes, such as hypermutable C8 tract mutations, are associated with a higher frequency of certain manifestations, like fewer renal tumors [17,20]. Several founder variants have been reported in Danish, Chinese, and Swedish families [15,28,29].

Efforts to catalog all *FLCN* variants in online databases have enhanced diagnostic and research capabilities in understanding BHDS [27]. A recent systematic literature review identified 1059 individuals with pathogenic *FLCN* variants across 575 families [30].

The online repository of *FLCN* genes maintained by "the Human Variome Project" at Leiden University in the Netherlands lists approximately 230 unique pathogenic and likely pathogenic mutations (http://www.lovd.nl/FLCN, online accessed 17 August 2024).

Somatic *FLCN* mutations occur in other tumors and may predispose individuals to cancer, although the frequency is very low. In a study by Gad et al., somatic *FLCN* mutations were found in 2 of 46 chromophobe renal cell carcinoma (chRCC) and 1 of 18 renal oncocytoma (RO) cases. Other studies have shown similar findings, suggesting that *FLCN* is not a major driver in other renal tumors [31].

While specific genotype–phenotype correlations in BHDS have not been firmly established [17], certain mutations in the *FLCN* gene have been proposed to be related to phenotypes. For example, the c.1285dupC variant is associated with a higher risk of developing renal cancer, and mutations in exon 9 are linked to an increased number of lung cysts and a tendency for PSP [32]. Furthermore, some studies have suggested ethnic variations in the clinical presentation of BHDS. For instance, it is suggested that skin abnormalities are more common in European population, while the Asian population manifests cutaneous symptoms less frequently; Chinese BHDS patients have a higher prevalence of large intragenic deletions spanning exons 1–3, seem to have increased risk of PSP [15,28,29,33–36].

3.3. Mechanism of Action of FLCN

Loss of *FLCN* function due to mutations leads to dysregulation of key cellular pathways such as AMP-activated protein kinase (AMPK) and mechanistic target of rapamycin (mTOR) pathways. This dysregulation causes several downstream effects that promote tumorigenesis, including increased cell proliferation, impaired cellular energy sensing, disrupted autophagy, and altered cellular differentiation.

AMP-activated protein kinase is a positive regulator of catabolic metabolism and a negative regulator of anabolic processes under low energy conditions [37]. FLCN interacts with FLCN-interacting proteins FNIP1 and FNIP2 [38–40], which are involved in the regulation of AMPK and AMPK-mediated energy sensing [41], by forming a complex with FNIP1, FNIP2, and FLCN through binding of the FNIPs to the C-terminal region of *FLCN*. Nearly all mutations in BHDS patients produce a C-terminally truncated FLCN unable to bind FNIP1 [38] (Figure 1).

The mTOR pathway, a major regulator of angiogenesis and cell growth, is rarely mutated but is a downstream effector of frequently mutated oncogenic pathways, including PI3K/Akt and Ras/Raf/Mek/Erk pathways. The Rag GTPases interact with mTORC1 and signal amino acid sufficiency by promoting the translocation of mTORC1 to the lysosomal surface, its site of activation. Structural studies have determined the role of FLCN as a GTPase-activating protein (GAP) for small GTPases, such as Rag GTPases. This GAP activity on the Rags is required for the recruitment of mTORC1 [37,42]. Hyperactivation of mTOR signaling is observed in 80% of human cancers [37]. Under normal conditions, FLCN inhibits the mTOR pathway through AMPK to maintain cellular homeostasis [43]. Downregulation of FLCN reduces the phosphorylation of ribosomal protein S6, an indicator of mTORC1 activity, and disruption in the mTOR signaling pathway results in uncontrolled cell proliferation and growth [37]. However, the role of FLCN/FNIP complex as a positive modulator of mTORC1 activity is controversial, and in certain cell lines, depletion of FLCN impairs mTORC1 activation [42,44].

FLCN loss of function also inhibits mTORC1-dependent phosphorylation of *TFE3/TFEB*, resulting in nuclear localization and activation of these transcription factors, which may play a role in tumorigenesis [44,45]. Along the same line of evidence, *PRDM10* alteration has been shown to reduce *FLCN* gene expression, driving *TFE3*-induced tumor formation via canonical mTOR pathway activation. Interestingly, *PRDM10* germline mutations cause a predisposition for a novel hereditary disorder in families with similar manifestations to BHDS, including fibrofolliculomas and renal cancers, but with reduced pulmonary involvement [46,47] (Figure 1).

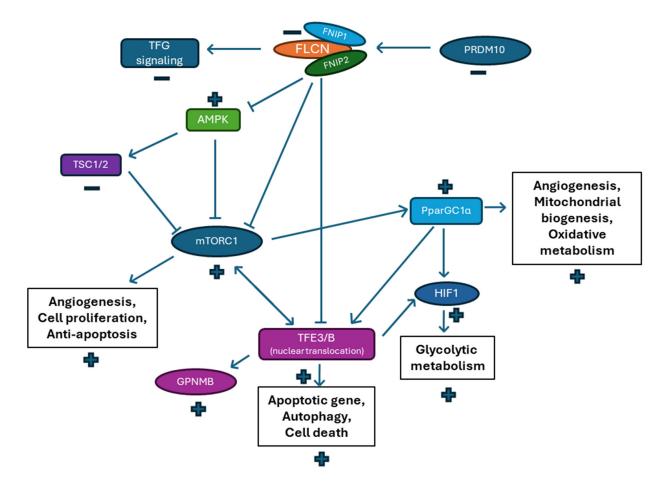


Figure 1. The function of FLCN/FNIP complex mostly occurs through the modulation of mTORC1 and AMPK, two of the key protein kinases. AMPK is an essential player in energetic homeostasis. Loss of FLCN due to *FLCN* gene mutations results in constitutive activation of AMPK. FLCN inhibits the mTOR pathway through AMPK to maintain cellular homeostasis and loss of FLCN leads to mTOR signaling pathway activation and uncontrolled cell proliferation and growth. Subcellular localization of TFE3 and TFEB is controlled by mTORC1. In the absence of FLCN, TFE3 and TFEB translocate to the nucleus and induce the expression of their target genes and induction of HIF pathway. PparGC1 α , which is a transcriptional coactivator involved in mitochondrial biogenesis and oxidative metabolism, is also activated by mTORC1.

Loss of *FLCN* induces the upregulation of PPARG coactivator 1 alpha (PPARGC1a) transcription factor, a potent inducer of mitochondrial biogenesis [48]. This leads to deregulation of the PGC-1 α -TFAM signaling axis and high expression of mitochondria- and oxidative phosphorylation associated genes [49]. Oxidative phosphorylation provides most of the energy in many somatic cells, whereas malignant transformation generally leads to an increased reliance on glycolysis despite the presence of oxygen (aerobic glycolysis), known as the Warburg effect [50]. FLCN inhibits lactate dehydrogenase A (LDHA) and regulates glycolysis, and pathogenic mutations enable LDHA hyperactivity due to the lack of direct inhibition by FLCN [51]. Loss of FLCN function also leads to AMPK-dependent increases in autophagy, HIF1/2 activity. Interestingly, similar phenomenon is reported in clear cell renal cell carcinoma (ccRCC), where reduced *FLCN* gene expression leads to hyperactive LDHA [51]. Finally, *FLCN* knockdown models in cell lines have been shown to cause impaired autophagy, an evolutionary conserved process of controlled degradation and recycling of damaged organelles and macromolecules [52].

There are rare reports of cases suggesting a combination of BHDS with other syndromes such as Multiple inherited neoplasia alleles syndrome (MINAS) (with multiple rare inherited cancer syndrome genes, including combinations of *FLCN* with *NF1*, *TP53*, *MSH2*, *MLH1, XPA, BRCA2*) or hereditary leiomyomatosis involving BHD [53,54], suggesting a cumulative increase in renal tumor risk. Vocke et al. reported four unrelated adults with Smith–Magenis syndrome (SMS, characterized by a distinctive facial appearance, varying degrees of cognitive impairment and distinct behavioral phenotype) [55] and concomitant features of BHDS [56]. Another case involved a young patient with both BHDS and hereditary paraganglioma-pheochromocytoma syndrome, presenting with metastatic ccRCC and no lung lesions with *FLCN* and *SDHB* germline mutations [57].

3.4. Two-Hit Hypothesis

Individuals with BHDS are born with one variant copy of the *FLCN* gene in each cell. The inactivation of both copies of the *FLCN* gene is a critical step in the development of these tumors. Without somatic second hit mutations, *FLCN* likely exists in a haploinsufficient form, potentially leading to impaired function [58]. Missense germline mutations are rare in BHDS, likely because missense *FLCN* mutations still lead to amino acid substitution that have little or no effect on folliculin function [59]. Heterozygosity for *FLCN* mutations seems sufficient to cause skin and lung lesions, while renal tumors require a second hit in the remaining wild-type allele [60].

Somatic mutations, mostly frameshift mutations or loss of heterozygosity (LOH), inactivate the second copy of the gene [17,61–64]. Second hit *FLCN* alterations may occur in the early third decade of life in BHDS patients [64]. However, some studies report low frequency of LOH in 17p as the second hit [63]. Different second hit mutations have been observed in patients with more than one tumor, supporting the notion that BHDS renal tumors occur independently [61].

3.5. mRNA Expression and Nonsense-Mediated Decay (NMD)

FLCN mRNA expression is hardly detectable in BHD-associated renal tumors [25,65,66], suggesting that degradation by NMD is a surveillance mechanism that eliminates mRNAs with premature termination codons (in all but the last exon of a gene) [67]. However, this view is challenged by some studies that did not find a significant differences in the *FLCN* transcript levels, suggesting incomplete NMD in certain mutations (i.e., c.563delT and c.1489-1490delTG) [13,49,68]. This may be due to a truncating mutation in the last 50 nucleotides of the penultimate exon escaping NMD [69]. Non-truncating mutations in *FLCN* do not disrupt the mRNA splicing pattern, supporting the hypothesis that these mutations impair folliculin function by disrupting the stability of the *FLCN* gene product [20,33].

4. Extrarenal Manifestations of BHDS

4.1. Pulmonary Manifestations of BHDS

Lung cysts and associated PSP are the most prevalent features of BHD. Pulmonary cysts, considered a key risk factor for pneumothorax development, occur in 70–100% of patients, typically forming by the age of 40 [3,17,20,22,70–73]. Most patients (76.9%) present with small pulmonary cysts less than 1 cm in diameter. Histologically these cysts are multiple small intraparenchymal structures rimmed by thin fibrous walls and normal pulmonary parenchyma. The vast majority of cysts are in lower lobes of the lungs [74].

Spontaneous pneumothorax is reported in 32–51% of patients with BHD, representing up to a 50-fold increase compared to general populations [20,30,72,75]. Loss of *FLCN* may lead to the imbalance of cell–cell adhesions and cell polarity, contributing to lung cyst development [76]. It is speculated that up to 5–10% of PSP cases are attributable to underlying BHD [77]. PSP may be caused by mutations in multiple genes including *FBN1*, *COL3A1*, *CBS*, *SERPINA1*, and *TSC1/TSC2* genes [78].

BHDS patients with pneumothorax tend to be older (mean age 42yo) and of normal weight (mean BMI 24.7) [74], which deviates from the usual demographic of PSP patients (younger, tall, and low BMI). Therefore, BHDS should be considered in the differential diagnosis of PSP, especially in the presence of a family history, older age, and normal weight. Recurrent PSP are common and tend to persist throughout life.

4.2. Cutaneous Manifestations of BHSD

Cutaneous lesions are one of the most common phenotypic features of BHDS found in 47–85% of BHDS cases, typically appearing by the third or fourth decade of life. These lesions are characterized by multiple small papules, including fibrofolliculomas, trichodiscomas, and acrochordons [3,17,71,79–83]. Fibrofolliculomas and trichodiscomas are benign tumors of the perifollicular connective tissue and mesodermal portion of the hair disk, occurring as yellowish dome-shaped papules. Fibrofolliculoma and trichodiscoma appear similarly in histology, consisting of hamartomatous hair follicles with cords and thin columns of epithelial components in a fenestrated pattern. It has been suggested that fibrofolliculomas and trichodiscomas represent the same lesion. The main difference between the two is that the former has epithelial cell proliferation emanating from the hair follicle, whereas the latter does not [84]. Acrochordons, also known as skin tags, are benign outgrowths of epidermal and dermal tissue, commonly found on the neck, eyelids, upper chest, and axilla. The treatment of cutaneous manifestations, such as fibrofolliculomas, usually involves ablative laser therapy.

4.3. Other Manifestations

Extensive screening studies of families with BHDS continue to report conflicting results about its associations with different lesions across the body. Conflicting reports about the association of BHDS with colon cancer or colonic polyposis and dysplastic lesions have been reported so far. However, a more recent study on 256 BHDS patients by Sattler et al. showed that 50% of BHDS patients had colorectal polyps, including tubulovillous adenomas with high-grade dysplasia and benign gastric polyps, and a moderately increased rate of colorectal cancer (5.1%) [85]. It has also been suggested that different mutations of the *FLCN* gene might confer varying risks for colorectal polyps and neoplasia in BHDS patients [86].

Other less common manifestations reported in BHDS include oral fibroma, parotid oncocytoma, lipomas, inverted papilloma of the nose, fibrosarcoma of the leg, basal cell carcinoma, squamous cell carcinoma, and lymphoma [83,87–93].

5. Renal Pathology in BHDS

5.1. Prevalence and Behavior

The lifetime risk (20–30%) for development of renal tumors in BHDS patients is much lower than lung involvement or skin involvement (approximately 90%). However, because of the bilateral and multifocal nature, the presence of renal tumors is one of the most serious manifestations in BHD patients. Renal involvement in BHDS manifests as renal cysts and renal tumors. Renal cysts occur in 25–30% of BHDS cases [33,73–75]. the prevalence of renal tumors in BHDS patients ranges from 27–34%, representing a sevenfold increase in risk compared to general population, with the median age of onset being 46–52 years, and no sex predilection [20,72,79,80,94,95]. However, the earliest onset of RCC in a BHDS patient has been reported at age 14 [81]. Renal tumors are often multiple and bilateral, commonly exhibiting a spectrum of histological patterns in the same patient. It was originally reported that the most common forms of renal tumors in BHDS patients are hybrid oncocytic tumors (HOT) (50–67%), chRCC (23–50%), and RO (3%) [79,94–97]. ccRCC and papillary RCC are also reported in BHDS but with much less frequency [24,79,94]. Based on our own evaluation of more than 20 HOTs, they are essentially the same tumor although they may display different histologic patterns, while other tumors such as ccRCC or papillary RCC may be coincidence, rather than related to FLCN mutations. More studies are needed to address this question. If second hit FLCN mutations are identified in the ccRCC and pRCC, in the absence of the genetic aberrations known to drive these tumors, it would be assumed they are caused by loss of FLCN.

Another interesting kidney-related phenomenon seen in approximately 58% of BHDS patients are oncocytosis, characterized by clusters or cysts of small epithelial oncocytic cells in non-neoplastic tubules, which may contribute to the development of renal tumors [80]. Although oncocytosis is associated with chRCC, most cases are now thought to be related to BHD, driven from principal cells as opposed to intercalated cells in RO and chRCC [98–100]. Interestingly, oncocytosis is present in 50% of HOTs, studies on the cell of origin show that these cells are different from those comprising RO or chRCC, with diffused L1CAM expression and absence of LINC01187 [100].

Metastasis in renal cell carcinoma is rare in BHDS patients, with instances of mortality, but none have been proven to arise from HOT, indicating a more indolent nature [17]. Houweling et al. reported metastasis in 5 out of 14 patients, 3 of which described as renal cell carcinoma with eosinophilic cytoplasm and characteristics of both ccRCC and chRCC [95]. Benusiglio et al. reported metastasis in 4 out of 32 (one of them was hybrid), and Pavlovich et al. reported 2 mortality due to metastasis out of 14 patients with follow up data, 1 histologically designated as ccRCC and 1 as predominantly clear cells with areas of tubular papillary and chromophobe histology [79,95,97].

5.2. HOT Morphology

Hybrid tumors are the most prevalent renal tumors in BHDS patients, notable for their unique morphology and tumorigenesis. The term "hybrid oncocytic tumor (HOT)" has been suggested for hereditary cases seen in Birt–Hogg–Dubé syndrome by the Genitourinary Pathology Society (GUPS) descriptions for renal tumors [101]. These tumors are estimated to have a slow growth rate of 0.1 cm per year [102]. Grossly, HOTs are circumscribed, discrete yellow to tan masses, ranging 0.7 to 5.5 cm, although masses as large as 20 cm have also been reported [81].

Morphologically, the main differential diagnoses include chromophore RCC and RO. HOT often presents with multiple and bilateral lesions, which can be seen in RO but are rare in chRCC. chRCC usually forms nested, alveolar or sheet like patterns and is cytologically distinguished by granular pale cells with prominent cell borders, a finely reticular cytoplasm, perinuclear halos, and wrinkled hyperchromatic nuclei, although it may show deeply eosinophilic features in the oncocytic variant of chRCC [103]. RO, on the other hand, is a benign renal epithelial neoplasm characterized by large round eosinophilic cells with uniformly round hyperchromatic nuclei; smooth nuclear borders very and low nuclear pleomorphism, typically forming small solid nests in a loose connective tissue (edematous-looking) stroma [104,105].

Microscopically, HOT exhibits solid to nested and alveolar/tubular architecture consisting of a checkerboard patterned mixture of RO-like cells with polygonal cells with light eosinophilic cytoplasm and minimal to no koilocytic atypia, with focal edematous stroma, and the second cell population with clear cytoplasm resembling chRCC and round monomorphic nuclei, inconspicuous nucleoli of WHO/International Society of Urological Pathology nucleolar grade 2 and perinuclear halos [64,94,106,107] (Figure 2). The key to correct diagnosis is the two-cell population present in an intermixed pattern, along with multi-locality and bilaterality as well as other syndromic clues. Therefore, a hybrid tumor should not imply that the tumor is a combination of chromophobe cells and renal oncocytoma cells. In our experience with more than 20 cases of HOTs, we have also observed several unique architectural patterns, such as alveolar, solid, slit-like, and microcystic.

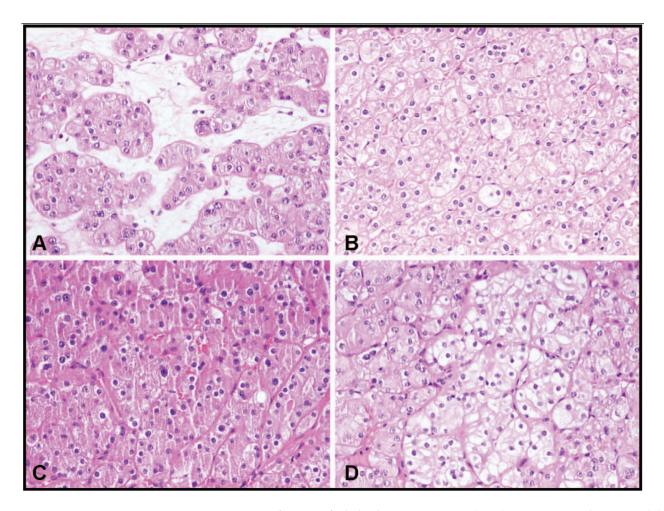


Figure 2. Microscopic features of a hybrid oncocytic tumor (HOT). Most commonly HOTs exhibit a solid nested architecture consisting of a mixture of polygonal cells with light eosinophilic cytoplasm and minimal to no koilocytic atypia resembling renal oncocytoma; and a second cell population with clear cytoplasm resembling chromophobe renal cell carcinoma (chRCC) with perinuclear halos. A predominantly eosinophilic cell nested population may resemble renal oncocytoma (**A**). HOT can resemble chRCC in some areas of the tumor (**B**) and can show a morphology that is not characteristic for renal oncocytoma or chRCC (**C**). An admixture of the two cell populations is characteristic of HOT (**D**). Magnifications 400×. (Reproduced with permission, Adley, B. et al., Arch Pathol Lab Med, 2006) [108].

5.3. Immunohistochemical (IHC) Staining Profile of HOT

The immunohistochemical profile of HOT is ambiguous between RO and chRCC, featuring heterogeneous immunophenotypical cell populations. HOT shows focal positivity for KRT7 and CKIT, and strong PAX8, while being negative for vimentin, and CA9 negative [94,106] (Figure 3). KRT7 is usually diffusely and strongly positive in chRCC, while negative in RO. In HOT, RO-like cells are negative for KRT7 while chRCC-like component is focally positive for KRT7 and diffusely positive for colloidal iron. CKIT is not helpful in distinguishing between these entities. S100A1 has also been reported positive in HOT, which further argues against chRCC. GPNMB (Marker of mTOR pathway activation) IHC stain is strong in the cytoplasm in HOT, but its sensitivity and specificity are not well defined [64,109].

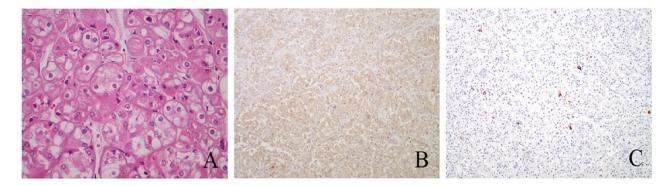


Figure 3. The commonly used immunohistochemical stains for the diagnosis of hybrid oncocytic tumors (**A**), besides positive PAX8 and negative CA9, are weak or negative CD117 (**B**) and patchy positivity for KRT7 (**C**). (Reproduced with permission, Li, J. et al., J Cancer Res Clin Oncol 2022) [110].

5.4. Chromosomal Abnormalities and Mutation Patterns of HOT

HOT exhibits a unique molecular profile distinct from other RCC types. BHDassociated HOTs do not have many recurrent mutations, such as mutations in classic RCC driver genes, such as VHL, BAP1, FH, MET, PTEN, TERT, TP53, ERCC2, and SDHA-D [111,112]. However, they exhibit unique copy number alterations more like RO than chRCC. Comparative gene expression profiling analysis showed that BHDS-related renal tumors have distinct gene expression in BHD-related tumors, with mitochondrial DNA harboring higher copy numbers and fewer variants compared to sporadic chRCC [64]. chRCC is typified by multiple monosomies (1, 2, 6, 7, 10, 13, 17, and 21) and the absence of polysomies [64,112,113]. Sporadic chRCC frequently *TP53* and/or *PTEN* alterations without *FLCN* alteration. RO chromosomal commonly harbors chromosome 1 and Y losses, as well as rearrangement of 11q13, which is the locus of the *CCND1* gene (cyclin D1) [114].

BHD-derived tumors generally exhibit fewer chromosomal abnormalities than sporadic chRCC [49]. Losses are observed in chromosomes 2p, 5p, 8p, 9p, and 19p, which is more similar to RO than chRCC [115]. Single nucleotide polymorphism array studies have shown copious numbers of LOH in BHD-associated renal tumors (classified as chRCC and HOT), mostly due to uniparental disomy, with a similar pattern between two types of tumors [63].

5.5. Two Cell Population in HOT

The origin of the tumor cells in HOT has long intrigued researchers. Recent studies using genomics and single cell sequencing techniques have shown that both chRCC and RO originate from intercalated cells, which have been proven to be different from HOT [64,100,116,117]. RNA transcript data from HOTs suggest an intermediate expression profile between RO and ChRCC [64]. Single cell sequencing data have revealed heterogeneous cell populations with mutually exclusive expression of certain genes [118]. Approximately 50% of the tumor epithelia express FOXI1 and LINC01187, markers of intercalated cells, which are homogeneously expressed in chRCC and RO [64,100,117,119]. The second populations of tumor cells, which are FOXI1-negative, show overexpression of L1CAM and are lineage-specific markers labeling and collecting duct principal cells in the benign kidney. Dual RNA in situ hybridization (RNA-ISH) of LINC1187 and L1CAM demonstrated mutually exclusive expression within HOT epithelia, suggesting that HOT is not a hybrid tumor because of admixture of 'chRCC and RO'-like areas [100]. Instead, HOT exhibits transcriptomic intratumor heterogeneity and displays morphologic and immunophenotypic heterogeneity due to an admixture of neoplastic cells arising from two distinct cells, the IC cells and PC cells [64,117]. This finding suggests that HOT tumors arise from a progenitor cell that is in a state of flux between IC and PC and capable of differentiating into both [100]. During embryologic development, NOTCH signaling regulates the differentiation of the ureteric bud into either L1CAM-positive principal cells or

FOXI1-positive intercalated cells [118]. It is proposed that NOTCH signaling may play a role in the observed heterogeneity of HOT tumors. By validating markers specific for these distinct epithelial populations within the HOT, L1CAM is suggested as the first line diagnostic marker to screen for HOT [100,117,119].

5.6. Clinical Implications and Management

The clinical management of HOTs in BHDS patients requires a tailored approach due to their unique characteristics and the potential for multiple and bilateral tumors and the high chance of recurrence. Given the high risk of renal tumors in BHDS patients, regular surveillance with imaging studies is recommended. Current guidelines suggest initiating surveillance in early adulthood, typically around the age of 20–25 years [22]. Imaging modalities, such as MRI or CT scans, are used to monitor the kidneys for tumor development allowing for timely intervention. Surgical approaches in BHD-associated kidney cancer aim to maximize renal function preservation while preventing metastatic disease. Complex partial nephrectomy is considered the approach of choice for familial kidney cancer syndromes like BHD. The classical protocol includes surveillance of masses of less than 3 cm emphasizing aggressive nephron-sparing techniques for smaller masses while reserving more extensive surgery for larger or more aggressive tumors [102,120,121].

5.7. Therapeutic Approaches

Advances in understanding the molecular pathways involved in BHDS have opened new avenues for targeted therapies. The involvement of the AMPK/mTOR pathway in BHDS pathogenesis suggests that mTOR inhibitors, such as everolimus and sirolimus (routinely used in the treatment of metastatic RCC), may be effective in treating BHDassociated tumors [122,123]. Rapamycin has shown efficacy in halting renal cyst and tumor growth in animal models. Early clinical trials with mTOR inhibitors, such as everolimus, have shown promise, but further research is needed to establish their efficacy and safety in BHDS patients. One study by Nakamura et al. showed that the administration of this drug resulted in longer progression-free survival compared to previously utilized sorafenib and sunitinib [111]. Additionally, understanding the role of autophagy in BHDS tumorigenesis could lead to novel therapeutic strategies targeting this pathway. Recent studies suggest that MET inhibitors, such as cabozantinib and crizotinib, may provide promising therapeutic approaches for BHDS-associated kidney cancer, given the high MET expression in various histological types of BHDS-associated tumors [118].

5.8. Preclinical Models

Developing accurate preclinical models of BHDS is critical for studying the disease and testing potential therapies. Animal models, such as mice with targeted inactivation of the *FLCN* gene, have provided valuable insights into the pathogenesis of BHD-associated tumors. Several *FLCN* knockout mouse models have been created, including skin-specific, lung-specific, muscle-specific, and kidney-specific knockouts. Whole-body *FLCN* knockout and FNIP1/2 double knockout mice are embryonic lethal, indicating a defect in nutrient uptake and transport in the *FLCN*-null embryo [122]. Whole-body *FNIP1* knockout mice, however, show B-cell developmental defects and muscle and cardiac hypertrophy but no kidney phenotype, whereas *FNIP2* knockout mice show no phenotype [124]. Kidneytargeted *FLCN* knockout or *FNIP1/FNIP2* double inactivated mice develop enlarged polycystic kidneys and die at 3 weeks of age due to renal failure [41,125,126]. These models have shown that loss of *FLCN* leads to the development of renal cysts and tumors resembling those seen in human BHDS patients [59,127].

Additionally, these models have been instrumental in studying the role of folliculin in various cellular pathways and in identifying potential therapeutic targets. For example, kidney-specific knockout models disrupting the *FLCN* gene in proximal tubules have demonstrated upregulation of mTOR and TGF- β signaling pathways, contributing to renal

tumorigenesis. Treatment with mTOR inhibitors, such as rapamycin or sirolimus, has been shown to suppress tumor growth in these models [128–130].

Other models, such as cell lines derived from BHD-associated tumors, also contribute to understanding the molecular mechanisms underlying BHDS and facilitate the testing of new drugs [131]. RCC cell line models deficient in the *FLCN* protein have shown increased sensitivity to Olaparib treatment, suggesting that *FLCN* deficiency may impair BRCA1-A complex-associated DNA repair ability, thereby making PARP inhibitors potentially more effective in these tumors [132].

6. Discussion

A hybrid oncocytic tumor is a uniquely interesting neoplasm, distinguished by its unique morphology and distinct molecular and tumorigenesis characteristics. In recent years, we have learned that HOT exhibits intrinsic heterogeneity and comprises two distinct cell types. This tumor class is markedly different from sporadic hybrid oncocytic/chromophobe tumors (HOCT), which have been proposed as a heterogeneous group with features intermediate between but distinct from chromophobe RCC and renal oncocytoma [113,133–135]. While sporadic HOCT has been extensively studied, the findings have not been cohesive enough to categorize them as a unified entity. Sporadic HOCTs are characterized by multiple numerical chromosomal aberrations, including both monosomies and polysomies of chromosomes 1, 2, 6, 9, 10, 13, 17, 21, and 22. These tumors lack mutations in key genes such as VHL, KIT, PDGFRA, and FLCN [113]. Clinically, these tumors are mostly indolent, the median age of the HOCT patients at the diagnosis is in seventh and eighth decades, are male predominant, and more often present as solitary masses [113,133,136]. These findings put HOCT in a different category compared to BHDS-related HOT. Unlike HOT, the two components of sporadic HOCT are not as intimately intermixed and are typically regarded as separate regions within the tumor, each displaying different morphologies. These findings suggest either two independent pathogenic pathways or an early pathogenic divergence from RO-like and chRCC-like components. Clinically, HOCTs are generally indolent with no evidence of disease recurrence, necrosis, or sarcomatoid change. However, cases of locally advanced disease and metastasis have been reported, indicating potential variability in their clinical behavior.

Transcriptomic analyses of BHD-associated HOT revealed intratumor heterogeneity comprising distinct cell clusters expressing L1CAM and FOXI1, representing two cell populations of intercalated cells (IC) and principal cells (PC). This puts HOT into a very interesting and unique group of tumors, in terms of tumorigenesis. Different models of tumor formation have been proposed. The clonal evolution model suggests that tumors arise from a single cell that acquires a growth advantage. In contrast, the big bang model, proposed in 2015, posits that tumors develop through a single expansion, continuously accumulating mutations not subjected to selective pressure [137]. The neutral evolution model suggests that tumors arise from cells undergoing sequential genomic insults or dedifferentiating into precursor cells with progenitor-like features that can later transform into tumors [138]. Most tumor models assume that mutations accumulate over time, leading to tumor cell dedifferentiation and loss of specialization, but HOT does not seem to perfectly match this model. Recent studies by Wang et al. have demonstrated that HOT originates from progenitor cells capable of differentiating into both IC and PC cells [100], perhaps fitting better in the neutral model and the progenitor-derived hypothesis. This model posits that tumors either lose differentiation into a primitive state or stem/progenitor cells serve as the origin, which contrasts with the course of tumor evolution seen in most other cancers with multiple hit progression. Interestingly, other rare tumors, such as combined hepatocellular-cholangiocarcinoma (cHCC-CCAs), display features of both cholangiocarcinoma and hepatocellular carcinoma in an intermixed pattern. Genomic data suggest a monoclonal origin for both HCC and CCA tumor components in mixed tumors, suggesting a progenitor cell-derived origin. cHCC-CCAs is still a controversial entity, but

its genomic data suggest a monoclonal origin for both HCC and CCA tumor components in mixed tumors [139].

Along the same line of evidence, considering the tumorigenesis and molecular pathways involved in BHDS tumorigenesis, it seems improbable that sporadic chRCC, with its known chromosomal losses, gains, and mutational patterns, would occur alongside HOT in BHDS patients. It is possible that at least some of chRCC cases of BHDS patients in the literature, without confirmation by CGH and molecular confirmatory evidence, may represent a spectrum of HOT, with higher percentage of one cell type as opposed to a balanced distribution. The literature has yet to definitively address this question, although recent RNA-seq data seems to point to this direction, as Jikuya et al. showed that BHDassociated renal tumors display different expression profiles from sporadic chRCC. chRCC does not typically present as multifocal [64]. We recommend that unless chRCC in BHDS is proven to exhibit the classic genetic and unequivocally exhibit classic features of sporadic chRCC without any disputed characteristics, the diagnosis of chRCC should be made with extreme care, as misdiagnosis could lead to unnecessary loss of kidney tissue in BHDS patients, who are prone to developing frequent tumors over time. In a similar fashion, particularly in biopsies, extreme care should be taken not to underdiagnose a potential hybrid tumor as RO. The impact of BHDS extends beyond the affected individuals, with significant implications for genetic counseling and family screening.

Future studies are needed to confirm recent discoveries on cells of origin of HOT and expand the findings presented here. *FLCN* mutations lead to a complex cascade of downstream effects via multiple cellular pathways. The exact mechanisms of these events are not fully understood. Specific mutational signatures (e.g., SBS1, SBS2, and SBS13) differ between chromophobe and BHDS tumors, highlighting the complexity of their genetic landscape [64]. This suggests that epigenetic changes in BHDS may play a role in the heterogeneity of tumors particularly because these tumors do not exhibit extensive mutational and chromosomal changes. By integrating clinical, genetic, and molecular insights, we can improve the diagnosis, management, and treatment of this rare but impactful syndrome.

In conclusion, HOTs in BHDS represent a unique subset of renal tumors with distinct histological, immunohistochemical and molecular features as well as biological behavior. The diagnostic criteria remain to be refined. Accurate diagnosis of these renal tumors can provide the best clinical management strategy and genetic monitoring for the family members who may have BHDS with increased risk of developing renal tumors and other clinical manifestations. Advances in understanding the pathways disrupted by *FLCN* mutations may facilitate the development of targeted therapies, improving outcomes for BHDS patients.

Author Contributions: Writing—original draft preparation, B.G.N.; writing—review and editing, B.G.N., X.L., B.T.T. and X.J.Y. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Conflicts of Interest: The authors declare no conflicts of interest.

References

- Padala, S.A.; Barsouk, A.; Thandra, K.C.; Saginala, K.; Mohammed, A.; Vakiti, A.; Rawla, P.; Barsouk, A. Epidemiology of Renal Cell Carcinoma. World J. Oncol. 2020, 11, 79–87. [CrossRef] [PubMed]
- Sung, H.; Ferlay, J.; Siegel, R.L.; Laversanne, M.; Soerjomataram, I.; Jemal, A.; Bray, F. Global Cancer Statistics 2020: GLOBOCAN Estimates of Incidence and Mortality Worldwide for 36 Cancers in 185 Countries. CA Cancer J. Clin. 2021, 71, 209–249. [CrossRef] [PubMed]
- 3. Adeniran, A.J.; Shuch, B.; Humphrey, P.A. Hereditary Renal Cell Carcinoma Syndromes: Clinical, Pathologic, and Genetic Features. *Am. J. Surg. Pathol.* **2015**, *39*, e1–e18. [CrossRef]
- 4. Barrisford, G.W.; Singer, E.A.; Rosner, I.L.; Linehan, W.M.; Bratslavsky, G. Familial renal cancer: Molecular genetics and surgical management. *Int. J. Surg. Oncol.* 2011, 2011, 658767. [CrossRef]
- Gaur, S.; Turkbey, B.; Choyke, P. Hereditary Renal Tumor Syndromes: Update on Diagnosis and Management. Semin. Ultrasound CT MR 2017, 38, 59–71. [CrossRef]
- 6. Hornstein, O.P.; Knickenberg, M. Perifollicular fibromatosis cutis with polyps of the colon--a cutaneo-intestinal syndrome sui generis. *Arch. Dermatol. Res.* **1975**, 253, 161–175. [CrossRef]
- Birt, A.R.; Hogg, G.R.; Dube, W.J. Hereditary multiple fibrofolliculomas with trichodiscomas and acrochordons. *Arch. Dermatol.* 1977, 113, 1674–1677. [CrossRef] [PubMed]
- 8. Roth, J.S.; Rabinowitz, A.D.; Benson, M.; Grossman, M.E. Bilateral renal cell carcinoma in the Birt-Hogg-Dube syndrome. *J. Am. Acad. Dermatol.* **1993**, *29*, 1055–1056. [CrossRef]
- 9. Khoo, S.K.; Bradley, M.; Wong, F.K.; Hedblad, M.A.; Nordenskjold, M.; Teh, B.T. Birt-Hogg-Dube syndrome: Mapping of a novel hereditary neoplasia gene to chromosome 17p12–q11.2. *Oncogene* 2001, 20, 5239–5242. [CrossRef]
- Schmidt, L.S.; Warren, M.B.; Nickerson, M.L.; Weirich, G.; Matrosova, V.; Toro, J.R.; Turner, M.L.; Duray, P.; Merino, M.; Hewitt, S.; et al. Birt-Hogg-Dube syndrome, a genodermatosis associated with spontaneous pneumothorax and kidney neoplasia, maps to chromosome 17p11.2. *Am. J. Hum. Genet.* 2001, *69*, 876–882. [CrossRef]
- 11. Nickerson, M.L.; Warren, M.B.; Toro, J.R.; Matrosova, V.; Glenn, G.; Turner, M.L.; Duray, P.; Merino, M.; Choyke, P.; Pavlovich, C.P.; et al. Mutations in a novel gene lead to kidney tumors, lung wall defects, and benign tumors of the hair follicle in patients with the Birt-Hogg-Dube syndrome. *Cancer Cell* **2002**, *2*, 157–164. [CrossRef]
- 12. Muller, M.E.; Daccord, C.; Taffe, P.; Lazor, R. Prevalence of Birt-Hogg-Dube Syndrome Determined Through Epidemiological Data on Spontaneous Pneumothorax and Bayes Theorem. *Front. Med.* **2021**, *8*, 631168. [CrossRef]
- 13. Naf, E.; Laubscher, D.; Hopfer, H.; Streit, M.; Matyas, G. Birt-Hogg-Dube syndrome: Novel FLCN frameshift deletion in daughter and father with renal cell carcinomas. *Fam. Cancer* **2016**, *15*, 127–132. [CrossRef] [PubMed]
- Savatt, J.M.; Shimelis, H.; Moreno-De-Luca, A.; Strande, N.T.; Oetjens, M.T.; Ledbetter, D.H.; Martin, C.L.; Myers, S.M.; Finucane, B.M. Frequency of truncating FLCN variants and Birt-Hogg-Dube-associated phenotypes in a health care system population. *Genet. Med.* 2022, 24, 1857–1866. [CrossRef] [PubMed]
- 15. Lagerstedt-Robinson, K.; Baranowska Korberg, I.; Tsiaprazis, S.; Bjorck, E.; Tham, E.; Poluha, A.; Hellstrom Pigg, M.; Paulsson-Karlsson, Y.; Nordenskjold, M.; Johansson-Soller, M.; et al. A retrospective two centre study of Birt-Hogg-Dube syndrome reveals a pathogenic founder mutation in FLCN in the Swedish population. *PLoS ONE* **2022**, *17*, e0264056. [CrossRef] [PubMed]
- 16. Shah, R.R.; Lambert, W.C.; Schwartz, R.A. Birt-Hogg-Dube syndrome: Another mTOR phenomenon. *Clin. Dermatol.* **2022**, *40*, 700–705. [CrossRef] [PubMed]
- Toro, J.R.; Wei, M.H.; Glenn, G.M.; Weinreich, M.; Toure, O.; Vocke, C.; Turner, M.; Choyke, P.; Merino, M.J.; Pinto, P.A.; et al. BHD mutations, clinical and molecular genetic investigations of Birt-Hogg-Dube syndrome: A new series of 50 families and a review of published reports. *J. Med. Genet.* 2008, 45, 321–331. [CrossRef]
- 18. Graham, R.B.; Nolasco, M.; Peterlin, B.; Garcia, C.K. Nonsense mutations in folliculin presenting as isolated familial spontaneous pneumothorax in adults. *Am. J. Respir. Crit. Care Med.* **2005**, *172*, 39–44. [CrossRef]
- 19. Painter, J.N.; Tapanainen, H.; Somer, M.; Tukiainen, P.; Aittomaki, K. A 4-bp deletion in the Birt-Hogg-Dube gene (FLCN) causes dominantly inherited spontaneous pneumothorax. *Am. J. Hum. Genet.* **2005**, *76*, 522–527. [CrossRef]
- 20. Schmidt, L.S.; Nickerson, M.L.; Warren, M.B.; Glenn, G.M.; Toro, J.R.; Merino, M.J.; Turner, M.L.; Choyke, P.L.; Sharma, N.; Peterson, J.; et al. Germline BHD-mutation spectrum and phenotype analysis of a large cohort of families with Birt-Hogg-Dube syndrome. *Am. J. Hum. Genet.* 2005, *76*, 1023–1033. [CrossRef]
- 21. Adley, B.P.; Schafernak, K.T.; Yeldandi, A.V.; Yang, X.J.; Nayar, R. Cytologic and histologic findings in multiple renal hybrid oncocytic tumors in a patient with Birt-Hogg-Dube syndrome: A case report. *Acta Cytol.* **2006**, *50*, 584–588. [CrossRef] [PubMed]
- Menko, F.H.; van Steensel, M.A.; Giraud, S.; Friis-Hansen, L.; Richard, S.; Ungari, S.; Nordenskjold, M.; Hansen, T.V.; Solly, J.; Maher, E.R.; et al. Birt-Hogg-Dube syndrome: Diagnosis and management. *Lancet Oncol.* 2009, 10, 1199–1206. [CrossRef] [PubMed]
- Geilswijk, M.; Genuardi, M.; Woodward, E.R.; Nightingale, K.; Huber, J.; Madsen, M.G.; Liekelema-van der Heij, D.; Lisseman, I.; Marle-Ballange, J.; McCarthy, C.; et al. ERN GENTURIS clinical practice guidelines for the diagnosis, surveillance and management of people with Birt-Hogg-Dube syndrome. *Eur. J. Hum. Genet.* 2024, 1–9. [CrossRef]

- 24. Palmirotta, R.; Savonarola, A.; Ludovici, G.; Donati, P.; Cavaliere, F.; ML, D.E.M.; Ferroni, P.; Guadagni, F. Association between Birt Hogg Dube syndrome and cancer predisposition. *Anticancer Res.* **2010**, *30*, 751–757.
- Warren, M.B.; Torres-Cabala, C.A.; Turner, M.L.; Merino, M.J.; Matrosova, V.Y.; Nickerson, M.L.; Ma, W.; Linehan, W.M.; Zbar, B.; Schmidt, L.S. Expression of Birt-Hogg-Dube gene mRNA in normal and neoplastic human tissues. *Mod. Pathol.* 2004, 17, 998–1011. [CrossRef]
- 26. Namba, Y.; Ebana, H.; Okamoto, S.; Kobayashi, E.; Kurihara, M.; Sekimoto, Y.; Tsuboshima, K.; Okura, M.K.; Mitsuishi, Y.; Takahashi, K.; et al. Clinical and genetic features of 334 Asian patients with Birt-Hogg-Dube syndrome (BHDS) who presented with pulmonary cysts with or without a history of pneumothorax, with special reference to BHDS-associated pneumothorax. *PLoS ONE* 2023, *18*, e0289175. [CrossRef]
- Lim, D.H.; Rehal, P.K.; Nahorski, M.S.; Macdonald, F.; Claessens, T.; Van Geel, M.; Gijezen, L.; Gille, J.J.; Giraud, S.; Richard, S.; et al. A new locus-specific database (LSDB) for mutations in the folliculin (FLCN) gene. *Hum. Mutat.* 2010, *31*, E1043–E1051. [CrossRef]
- Rossing, M.; Albrechtsen, A.; Skytte, A.B.; Jensen, U.B.; Ousager, L.B.; Gerdes, A.M.; Nielsen, F.C.; Hansen, T.V. Genetic screening of the FLCN gene identify six novel variants and a Danish founder mutation. *J. Hum. Genet.* 2017, 62, 151–157. [CrossRef] [PubMed]
- 29. Wang, Y.; Cai, M.; Jiang, X.; Lv, G.; Hu, D.; Zhang, G.; Liu, J.; Wei, W.; Xiao, J.; Shen, B.; et al. Exons 1-3 deletion in FLCN is associated with increased risk of pneumothorax in Chinese patients with Birt-Hogg-Dube syndrome. *Orphanet J. Rare Dis.* **2023**, *18*, 115. [CrossRef]
- Matsumoto, K.; Lim, D.; Pharoah, P.D.; Maher, E.R.; Marciniak, S.J. A systematic review assessing the existence of pneumothoraxonly variants of FLCN. Implications for lifelong surveillance of renal tumours. *Eur. J. Hum. Genet.* 2021, 29, 1595–1600. [CrossRef]
- Gad, S.; Lefevre, S.H.; Khoo, S.K.; Giraud, S.; Vieillefond, A.; Vasiliu, V.; Ferlicot, S.; Molinie, V.; Denoux, Y.; Thiounn, N.; et al. Mutations in BHD and TP53 genes, but not in HNF1beta gene, in a large series of sporadic chromophobe renal cell carcinoma. *Br. J. Cancer* 2007, *96*, 336–340. [CrossRef] [PubMed]
- 32. Toro, J.R.; Pautler, S.E.; Stewart, L.; Glenn, G.M.; Weinreich, M.; Toure, O.; Wei, M.H.; Schmidt, L.S.; Davis, L.; Zbar, B.; et al. Lung cysts, spontaneous pneumothorax, and genetic associations in 89 families with Birt-Hogg-Dube syndrome. *Am. J. Respir. Crit. Care Med.* **2007**, *175*, 1044–1053. [CrossRef] [PubMed]
- 33. Liu, K.; Xu, W.; Tian, X.; Xiao, M.; Zhao, X.; Zhang, Q.; Qu, T.; Song, J.; Liu, Y.; Xu, K.F.; et al. Genotypic characteristics of Chinese patients with BHD syndrome and functional analysis of FLCN variants. *Orphanet J. Rare Dis.* **2019**, *14*, 223. [CrossRef]
- Kuroda, N.; Furuya, M.; Nagashima, Y.; Gotohda, H.; Kawakami, F.; Moritani, S.; Ota, S.; Hora, M.; Michal, M.; Hes, O.; et al. Review of renal tumors associated with Birt-Hogg-Dube syndrome with focus on clinical and pathobiological aspects. *Pol. J. Pathol.* 2014, 65, 93–99. [CrossRef] [PubMed]
- 35. Zhou, W.; Liu, K.; Xu, K.F.; Liu, Y.; Tian, X. Clinical and Genetic Comparison of Birt-Hogg-Dube Syndrome (Hornstein-Knickenberg Syndrome) in Chinese: A Systemic Review of Reported Cases. *Int. J. Gen. Med.* **2022**, *15*, 5111–5121. [CrossRef]
- Lee, J.H.; Jeon, M.J.; Song, J.S.; Chae, E.J.; Choi, J.H.; Kim, G.H.; Song, J.W. Birt-Hogg-Dube syndrome in Korean: Clinicoradiologic features and long term follow-up. *Korean J. Intern. Med.* 2019, 34, 830–840. [CrossRef]
- 37. Ramirez Reyes, J.M.J.; Cuesta, R.; Pause, A. Folliculin: A Regulator of Transcription Through AMPK and mTOR Signaling Pathways. *Front. Cell Dev. Biol.* 2021, 9, 667311. [CrossRef]
- 38. Baba, M.; Hong, S.B.; Sharma, N.; Warren, M.B.; Nickerson, M.L.; Iwamatsu, A.; Esposito, D.; Gillette, W.K.; Hopkins, R.F., 3rd; Hartley, J.L.; et al. Folliculin encoded by the BHD gene interacts with a binding protein, FNIP1, and AMPK, and is involved in AMPK and mTOR signaling. *Proc. Natl. Acad. Sci. USA* 2006, 103, 15552–15557. [CrossRef]
- 39. Hasumi, H.; Baba, M.; Hong, S.B.; Hasumi, Y.; Huang, Y.; Yao, M.; Valera, V.A.; Linehan, W.M.; Schmidt, L.S. Identification and characterization of a novel folliculin-interacting protein FNIP2. *Gene* 2008, *415*, 60–67. [CrossRef]
- Takagi, Y.; Kobayashi, T.; Shiono, M.; Wang, L.; Piao, X.; Sun, G.; Zhang, D.; Abe, M.; Hagiwara, Y.; Takahashi, K.; et al. Interaction of folliculin (Birt-Hogg-Dube gene product) with a novel Fnip1-like (FnipL/Fnip2) protein. *Oncogene* 2008, 27, 5339–5347. [CrossRef]
- Baba, M.; Furihata, M.; Hong, S.B.; Tessarollo, L.; Haines, D.C.; Southon, E.; Patel, V.; Igarashi, P.; Alvord, W.G.; Leighty, R.; et al. Kidney-targeted Birt-Hogg-Dube gene inactivation in a mouse model: Erk1/2 and Akt-mTOR activation, cell hyperproliferation, and polycystic kidneys. J. Natl. Cancer Inst. 2008, 100, 140–154. [CrossRef] [PubMed]
- 42. Tsun, Z.Y.; Bar-Peled, L.; Chantranupong, L.; Zoncu, R.; Wang, T.; Kim, C.; Spooner, E.; Sabatini, D.M. The folliculin tumor suppressor is a GAP for the RagC/D GTPases that signal amino acid levels to mTORC1. *Mol. Cell* **2013**, *52*, 495–505. [CrossRef]
- Gupta, S.; Kang, H.C.; Ganeshan, D.; Morani, A.; Gautam, R.; Choyke, P.L.; Kundra, V. The ABCs of BHD: An In-Depth Review of Birt-Hogg-Dube Syndrome. *AJR Am. J. Roentgenol.* 2017, 209, 1291–1296. [CrossRef] [PubMed]
- 44. Petit, C.S.; Roczniak-Ferguson, A.; Ferguson, S.M. Recruitment of folliculin to lysosomes supports the amino acid-dependent activation of Rag GTPases. J. Cell Biol. 2013, 202, 1107–1122. [CrossRef]

- 45. Hong, S.B.; Oh, H.; Valera, V.A.; Baba, M.; Schmidt, L.S.; Linehan, W.M. Inactivation of the FLCN tumor suppressor gene induces TFE3 transcriptional activity by increasing its nuclear localization. *PLoS ONE* **2010**, *5*, e15793. [CrossRef]
- Schmidt, L.S.; Vocke, C.D.; Ricketts, C.J.; Blake, Z.; Choo, K.K.; Nielsen, D.; Gautam, R.; Crooks, D.R.; Reynolds, K.L.; Krolus, J.L.; et al. PRDM10 RCC: A Birt-Hogg-Dube-like Syndrome Associated With Lipoma and Highly Penetrant, Aggressive Renal Tumors Morphologically Resembling Type 2 Papillary Renal Cell Carcinoma. *Urology* 2023, *179*, 58–70. [CrossRef] [PubMed]
- 47. van de Beek, I.; Glykofridis, I.E.; Oosterwijk, J.C.; van den Akker, P.C.; Diercks, G.F.H.; Bolling, M.C.; Waisfisz, Q.; Mensenkamp, A.R.; Balk, J.A.; Zwart, R.; et al. PRDM10 directs FLCN expression in a novel disorder overlapping with Birt-Hogg-Dube syndrome and familial lipomatosis. *Hum. Mol. Genet.* 2023, *32*, 1223–1235. [CrossRef]
- Yan, M.; Gingras, M.C.; Dunlop, E.A.; Nouet, Y.; Dupuy, F.; Jalali, Z.; Possik, E.; Coull, B.J.; Kharitidi, D.; Dydensborg, A.B.; et al. The tumor suppressor folliculin regulates AMPK-dependent metabolic transformation. *J. Clin. Investig.* 2014, 124, 2640–2650. [CrossRef]
- 49. Klomp, J.A.; Petillo, D.; Niemi, N.M.; Dykema, K.J.; Chen, J.; Yang, X.J.; Saaf, A.; Zickert, P.; Aly, M.; Bergerheim, U.; et al. Birt-Hogg-Dube renal tumors are genetically distinct from other renal neoplasias and are associated with up-regulation of mitochondrial gene expression. *BMC Med. Genom.* **2010**, *3*, 59. [CrossRef]
- 50. Warburg, O.; Wind, F.; Negelein, E. The Metabolism of Tumors in the Body. J. Gen. Physiol. 1927, 8, 519–530. [CrossRef]
- Woodford, M.R.; Baker-Williams, A.J.; Sager, R.A.; Backe, S.J.; Blanden, A.R.; Hashmi, F.; Kancherla, P.; Gori, A.; Loiselle, D.R.; Castelli, M.; et al. The tumor suppressor folliculin inhibits lactate dehydrogenase A and regulates the Warburg effect. *Nat. Struct. Mol. Biol.* 2021, 28, 662–670. [CrossRef] [PubMed]
- Dunlop, E.A.; Seifan, S.; Claessens, T.; Behrends, C.; Kamps, M.A.; Rozycka, E.; Kemp, A.J.; Nookala, R.K.; Blenis, J.; Coull, B.J.; et al. FLCN, a novel autophagy component, interacts with GABARAP and is regulated by ULK1 phosphorylation. *Autophagy* 2014, 10, 1749–1760. [CrossRef] [PubMed]
- Pan, H.H.; Ruan, D.D.; Wu, M.; Chen, T.; Lu, T.; Gan, Y.M.; Wang, C.; Liao, L.S.; Lin, X.F.; Chen, X.; et al. Clinical phenotype and genetic function analysis of a rare family with hereditary leiomyomatosis and renal cell carcinoma complicated with Birt-Hogg-Dube syndrome. J. Med. Genet. 2023, 60, 1210–1214. [CrossRef]
- Whitworth, J.; Skytte, A.B.; Sunde, L.; Lim, D.H.; Arends, M.J.; Happerfield, L.; Frayling, I.M.; van Minkelen, R.; Woodward, E.R.; Tischkowitz, M.D.; et al. Multilocus Inherited Neoplasia Alleles Syndrome: A Case Series and Review. *JAMA Oncol.* 2016, 2, 373–379. [CrossRef] [PubMed]
- 55. Elsea, S.H.; Girirajan, S. Smith-Magenis syndrome. Eur. J. Hum. Genet. 2008, 16, 412–421. [CrossRef]
- Vocke, C.D.; Fleming, L.R.; Piskorski, A.M.; Amin, A.; Phornphutkul, C.; de la Monte, S.; Vilboux, T.; Duncan, F.; Pellegrino, J.; Braddock, B.; et al. A diagnosis of Birt-Hogg-Dube syndrome in individuals with Smith-Magenis syndrome: Recommendation for cancer screening. *Am. J. Med. Genet. A* 2023, 191, 490–497. [CrossRef]
- Boland, J.; Shahbazi, D.; Stevenson, R.; Shahbazi, S. Concurrent Birt-Hogg-Dube Syndrome and Hereditary Paraganglioma-Pheochromocytoma Syndrome Presenting as Metastatic Renal Cell Carcinoma in a 25-Year-Old Man: A Case Report. *Perm. J.* 2020, 24, 1–6. [CrossRef]
- Hoshika, Y.; Takahashi, F.; Togo, S.; Hashimoto, M.; Nara, T.; Kobayashi, T.; Nurwidya, F.; Kataoka, H.; Kurihara, M.; Kobayashi, E.; et al. Haploinsufficiency of the folliculin gene leads to impaired functions of lung fibroblasts in patients with Birt-Hogg-Dube syndrome. *Physiol. Rep.* 2016, 4, e13025. [CrossRef]
- Hasumi, H.; Hasumi, Y.; Baba, M.; Nishi, H.; Furuya, M.; Vocke, C.D.; Lang, M.; Irie, N.; Esumi, C.; Merino, M.J.; et al. H255Y and K508R missense mutations in tumour suppressor folliculin (FLCN) promote kidney cell proliferation. *Hum. Mol. Genet.* 2017, 26, 354–366. [CrossRef]
- van Steensel, M.A.; Verstraeten, V.L.; Frank, J.; Kelleners-Smeets, N.W.; Poblete-Gutierrez, P.; Marcus-Soekarman, D.; Bladergroen, R.S.; Steijlen, P.M.; van Geel, M. Novel mutations in the BHD gene and absence of loss of heterozygosity in fibrofolliculomas of Birt-Hogg-Dube patients. J. Investig. Dermatol. 2007, 127, 588–593. [CrossRef]
- 61. Vocke, C.D.; Yang, Y.; Pavlovich, C.P.; Schmidt, L.S.; Nickerson, M.L.; Torres-Cabala, C.A.; Merino, M.J.; Walther, M.M.; Zbar, B.; Linehan, W.M. High frequency of somatic frameshift BHD gene mutations in Birt-Hogg-Dube-associated renal tumors. *J. Natl. Cancer Inst.* **2005**, *97*, 931–935. [CrossRef] [PubMed]
- 62. Schmidt, L.S.; Linehan, W.M. Molecular genetics and clinical features of Birt-Hogg-Dube syndrome. *Nat. Rev. Urol.* 2015, 12, 558–569. [CrossRef]
- Iribe, Y.; Yao, M.; Tanaka, R.; Kuroda, N.; Nagashima, Y.; Nakatani, Y.; Furuya, M. Genome-Wide Uniparental Disomy and Copy Number Variations in Renal Cell Carcinomas Associated with Birt-Hogg-Dube Syndrome. *Am. J. Pathol.* 2016, 186, 337–346. [CrossRef] [PubMed]
- 64. Jikuya, R.; Johnson, T.A.; Maejima, K.; An, J.; Ju, Y.S.; Lee, H.; Ha, K.; Song, W.; Kim, Y.; Okawa, Y.; et al. Comparative analyses define differences between BHD-associated renal tumour and sporadic chromophobe renal cell carcinoma. *EBioMedicine* **2023**, *92*, 104596. [CrossRef] [PubMed]
- 65. Liu, L.; Yang, K.; Wang, X.; Shi, Z.; Yang, Y.; Yuan, Y.; Guo, T.; Xiao, X.; Luo, H. Detection of Folliculin Gene Mutations in Two Chinese Families with Birt-Hogg-Dube Syndrome. *BioMed Res. Int.* **2017**, *2017*, 8751384. [CrossRef] [PubMed]

- 66. Murakami, T.; Sano, F.; Huang, Y.; Komiya, A.; Baba, M.; Osada, Y.; Nagashima, Y.; Kondo, K.; Nakaigawa, N.; Miura, T.; et al. Identification and characterization of Birt-Hogg-Dube associated renal carcinoma. *J. Pathol.* **2007**, *211*, 524–531. [CrossRef]
- 67. Maquat, L.E. Nonsense-mediated mRNA decay: Splicing, translation and mRNP dynamics. *Nat. Rev. Mol. Cell Biol.* **2004**, *5*, 89–99. [CrossRef]
- 68. Park, Y.J.; Lee, S.K.; Kang, S.H.; Jang, S.J.; Moon, D.S.; Park, G. A Novel FLCN c.1489_1490delTG Mutation that Escapes the Nonsense-Mediated Decay System. *Ann. Clin. Lab. Sci.* **2016**, *46*, 562–565.
- 69. White, J.; Mazzeu, J.F.; Hoischen, A.; Jhangiani, S.N.; Gambin, T.; Alcino, M.C.; Penney, S.; Saraiva, J.M.; Hove, H.; Skovby, F.; et al. DVL1 frameshift mutations clustering in the penultimate exon cause autosomal-dominant Robinow syndrome. *Am. J. Hum. Genet.* **2015**, *96*, 612–622. [CrossRef]
- 70. Hao, S.; Long, F.; Sun, F.; Liu, T.; Li, D.; Jiang, S. Birt-Hogg-Dube syndrome: A literature review and case study of a Chinese woman presenting a novel FLCN mutation. *BMC Pulm. Med.* **2017**, *17*, 43. [CrossRef]
- 71. Maher, E.R. Genetics of familial renal cancers. Nephron Exp. Nephrol. 2011, 118, e21–e26. [CrossRef] [PubMed]
- 72. Zbar, B.; Alvord, W.G.; Glenn, G.; Turner, M.; Pavlovich, C.P.; Schmidt, L.; Walther, M.; Choyke, P.; Weirich, G.; Hewitt, S.M.; et al. Risk of renal and colonic neoplasms and spontaneous pneumothorax in the Birt-Hogg-Dube syndrome. *Cancer Epidemiol. Biomarkers Prev.* **2002**, *11*, 393–400. [PubMed]
- 73. Skolnik, K.; Tsai, W.H.; Dornan, K.; Perrier, R.; Burrowes, P.W.; Davidson, W.J. Birt-Hogg-Dube syndrome: A large single family cohort. *Respir. Res.* **2016**, *17*, 22. [CrossRef] [PubMed]
- 74. Torricelli, E.; Occhipinti, M.; Cavigli, E.; Tancredi, G.; Rosi, E.; Rossi, C.; Bonaguro, M.; Candita, L.; Papi, L.; Novelli, L.; et al. The Relevance of Family History Taking in the Detection and Management of Birt-Hogg-Dube Syndrome. *Respiration* 2019, 98, 125–132. [CrossRef]
- 75. Leter, E.M.; Koopmans, A.K.; Gille, J.J.; van Os, T.A.; Vittoz, G.G.; David, E.F.; Jaspars, E.H.; Postmus, P.E.; van Moorselaar, R.J.; Craanen, M.E.; et al. Birt-Hogg-Dube syndrome: Clinical and genetic studies of 20 families. *J. Investig. Dermatol.* 2008, 128, 45–49. [CrossRef]
- 76. Schmidt, L.S.; Linehan, W.M. FLCN: The causative gene for Birt-Hogg-Dube syndrome. Gene 2018, 640, 28–42. [CrossRef]
- 77. Scott, R.M.; Henske, E.P.; Raby, B.; Boone, P.M.; Rusk, R.A.; Marciniak, S.J. Familial pneumothorax: Towards precision medicine. *Thorax* 2018, 73, 270–276. [CrossRef]
- 78. Zhang, X.; Ma, D.; Zou, W.; Ding, Y.; Zhu, C.; Min, H.; Zhang, B.; Wang, W.; Chen, B.; Ye, M.; et al. A rapid NGS strategy for comprehensive molecular diagnosis of Birt-Hogg-Dube syndrome in patients with primary spontaneous pneumothorax. *Respir. Res.* 2016, 17, 64. [CrossRef]
- Benusiglio, P.R.; Giraud, S.; Deveaux, S.; Mejean, A.; Correas, J.M.; Joly, D.; Timsit, M.O.; Ferlicot, S.; Verkarre, V.; Abadie, C.; et al. Renal cell tumour characteristics in patients with the Birt-Hogg-Dube cancer susceptibility syndrome: A retrospective, multicentre study. *Orphanet J. Rare Dis.* 2014, 9, 163. [CrossRef]
- Pavlovich, C.P.; Walther, M.M.; Eyler, R.A.; Hewitt, S.M.; Zbar, B.; Linehan, W.M.; Merino, M.J. Renal tumors in the Birt-Hogg-Dube syndrome. *Am. J. Surg. Pathol.* 2002, 26, 1542–1552. [CrossRef]
- Schneider, M.; Dinkelborg, K.; Xiao, X.; Chan-Smutko, G.; Hruska, K.; Huang, D.; Sagar, P.; Harisinghani, M.; Iliopoulos, O. Early onset renal cell carcinoma in an adolescent girl with germline FLCN exon 5 deletion. *Fam. Cancer* 2018, *17*, 135–139. [CrossRef] [PubMed]
- Kunogi, M.; Kurihara, M.; Ikegami, T.S.; Kobayashi, T.; Shindo, N.; Kumasaka, T.; Gunji, Y.; Kikkawa, M.; Iwakami, S.; Hino, O.; et al. Clinical and genetic spectrum of Birt-Hogg-Dube syndrome patients in whom pneumothorax and/or multiple lung cysts are the presenting feature. J. Med. Genet. 2010, 47, 281–287. [CrossRef] [PubMed]
- Maffe, A.; Toschi, B.; Circo, G.; Giachino, D.; Giglio, S.; Rizzo, A.; Carloni, A.; Poletti, V.; Tomassetti, S.; Ginardi, C.; et al. Constitutional FLCN mutations in patients with suspected Birt-Hogg-Dube syndrome ascertained for non-cutaneous manifestations. *Clin. Genet.* 2011, *79*, 345–354. [CrossRef] [PubMed]
- Ackerman, A.B.; Reddy, V.B.; Soyer, H.P. Fibrofolliculoma/Trichodiscoma. In *Neoplasms With Follicular Differentiation*; Ackerman, A.B., Reddy, V.B., Soyer, H.P., Eds.; Ardor Scribendi: Philadelphia, PA, USA, 2001; pp. 221–244.
- 85. Sattler, E.C.; Syunyaeva, Z.; Reithmair, M.; Dempke, W.; Steinlein, O.K. Colorectal cancer risk in families with Birt-Hogg-Dube syndrome increased. *Eur. J. Cancer* 2021, *151*, 168–174. [CrossRef]
- 86. Jirka, G.W.; Lefler, D.S.; Russo, J.; Bashir, B. Colon adenocarcinoma and Birt-Hogg-Dube syndrome in a young patient: Case report and exploration of pathologic implications. *Cancer Biol. Ther.* **2023**, *24*, 2184153. [CrossRef]
- 87. Toro, J.R.; Glenn, G.; Duray, P.; Darling, T.; Weirich, G.; Zbar, B.; Linehan, M.; Turner, M.L. Birt-Hogg-Dube syndrome: A novel marker of kidney neoplasia. *Arch. Dermatol.* **1999**, *135*, 1195–1202. [CrossRef]
- 88. Nadershahi, N.A.; Wescott, W.B.; Egbert, B. Birt-Hogg-Dube syndrome: A review and presentation of the first case with oral lesions. *Oral Surg. Oral Med. Oral Pathol. Oral Radiol. Endodontol.* **1997**, *83*, 496–500. [CrossRef]
- Vincent, A.; Farley, M.; Chan, E.; James, W.D. Birt-Hogg-Dube syndrome: Two patients with neural tissue tumors. J. Am. Acad. Dermatol. 2003, 49, 717–719. [CrossRef]
- 90. Welsch, M.J.; Krunic, A.; Medenica, M.M. Birt-Hogg-Dube Syndrome. Int. J. Dermatol. 2005, 44, 668–673. [CrossRef]
- 91. Yoshida, K.; Miyagawa, M.; Kido, T.; Ide, K.; Sano, Y.; Sugawara, Y.; Takahata, H.; Monden, N.; Furuya, M.; Mochizuki, T. Parotid Oncocytoma as a Manifestation of Birt-Hogg-Dube Syndrome. *Case Rep. Radiol.* **2018**, 2018, 6265175. [CrossRef]

- 92. Xin, J.; Goffinet, A.; Machusko, S.; Shoela, R. Parotid Acinic Cell Carcinoma as a Presentation of Birt-Hogg-Dube Syndrome. *Cureus* 2023, *15*, e36074. [CrossRef] [PubMed]
- 93. Chen, E.L.; Siu, J.; Bastawrous, S.; Truong, C.D.; Wu, L. Oncocytic carcinoma of the parotid gland as a manifestation of Birt-Hogg-Dube syndrome. *Radiol. Case Rep.* **2023**, *18*, 1536–1543. [CrossRef] [PubMed]
- 94. Kennedy, J.M.; Wang, X.; Plouffe, K.R.; Dhanasekaran, S.M.; Hafez, K.; Palapattu, G.S.; Else, T.; Weizer, A.Z.; Morgan, T.M.; Spratt, D.E.; et al. Clinical and morphologic review of 60 hereditary renal tumors from 30 hereditary renal cell carcinoma syndrome patients: Lessons from a contemporary single institution series. *Med. Oncol.* 2019, 36, 74. [CrossRef]
- Houweling, A.C.; Gijezen, L.M.; Jonker, M.A.; van Doorn, M.B.; Oldenburg, R.A.; van Spaendonck-Zwarts, K.Y.; Leter, E.M.; van Os, T.A.; van Grieken, N.C.; Jaspars, E.H.; et al. Renal cancer and pneumothorax risk in Birt-Hogg-Dube syndrome; an analysis of 115 FLCN mutation carriers from 35 BHD families. *Br. J. Cancer* 2011, 105, 1912–1919. [CrossRef] [PubMed]
- 96. Hasumi, H.; Baba, M.; Hasumi, Y.; Furuya, M.; Yao, M. Birt-Hogg-Dube syndrome: Clinical and molecular aspects of recently identified kidney cancer syndrome. *Int. J. Urol.* 2016, 23, 204–210. [CrossRef] [PubMed]
- 97. Pavlovich, C.P.; Grubb, R.L., 3rd; Hurley, K.; Glenn, G.M.; Toro, J.; Schmidt, L.S.; Torres-Cabala, C.; Merino, M.J.; Zbar, B.; Choyke, P.; et al. Evaluation and management of renal tumors in the Birt-Hogg-Dube syndrome. J. Urol. 2005, 173, 1482–1486. [CrossRef]
- 98. Tickoo, S.K.; Reuter, V.E.; Amin, M.B.; Srigley, J.R.; Epstein, J.I.; Min, K.W.; Rubin, M.A.; Ro, J.Y. Renal oncocytosis: A morphologic study of fourteen cases. *Am. J. Surg. Pathol.* **1999**, 23, 1094–1101. [CrossRef]
- Gobbo, S.; Eble, J.N.; Delahunt, B.; Grignon, D.J.; Samaratunga, H.; Martignoni, G.; Zhang, S.; Wang, M.; Brunelli, M.; Cossu-Rocca, P.; et al. Renal cell neoplasms of oncocytosis have distinct morphologic, immunohistochemical, and cytogenetic profiles. *Am. J. Surg. Pathol.* 2010, *34*, 620–626. [CrossRef]
- 100. Wang, X.M.; Mannan, R.; Zhang, Y.; Chinnaiyan, A.; Rangaswamy, R.; Chugh, S.; Su, F.; Cao, X.; Wang, R.; Skala, S.L.; et al. Hybrid Oncocytic Tumors (HOTs) in Birt-Hogg-Dube Syndrome Patients-A Tale of Two Cities: Sequencing Analysis Reveals Dual Lineage Markers Capturing the 2 Cellular Populations of HOT. *Am. J. Surg. Pathol.* 2024, *48*, 163–173. [CrossRef]
- 101. Trpkov, K.; Hes, O.; Williamson, S.R.; Adeniran, A.J.; Agaimy, A.; Alaghehbandan, R.; Amin, M.B.; Argani, P.; Chen, Y.B.; Cheng, L.; et al. New developments in existing WHO entities and evolving molecular concepts: The Genitourinary Pathology Society (GUPS) update on renal neoplasia. *Mod. Pathol.* 2021, 34, 1392–1424. [CrossRef]
- 102. Ball, M.W.; An, J.Y.; Gomella, P.T.; Gautam, R.; Ricketts, C.J.; Vocke, C.D.; Schmidt, L.S.; Merino, M.J.; Srinivasan, R.; Malayeri, A.A.; et al. Growth Rates of Genetically Defined Renal Tumors: Implications for Active Surveillance and Intervention. *J. Clin. Oncol.* 2020, *38*, 1146–1153. [CrossRef]
- 103. Amin, M.B.; Paner, G.P.; Alvarado-Cabrero, I.; Young, A.N.; Stricker, H.J.; Lyles, R.H.; Moch, H. Chromophobe renal cell carcinoma: Histomorphologic characteristics and evaluation of conventional pathologic prognostic parameters in 145 cases. *Am. J. Surg. Pathol.* 2008, *32*, 1822–1834. [CrossRef]
- Amin, M.B.; Crotty, T.B.; Tickoo, S.K.; Farrow, G.M. Renal oncocytoma: A reappraisal of morphologic features with clinicopathologic findings in 80 cases. Am. J. Surg. Pathol. 1997, 21, 1–12. [CrossRef]
- 105. Perez-Ordonez, B.; Hamed, G.; Campbell, S.; Erlandson, R.A.; Russo, P.; Gaudin, P.B.; Reuter, V.E. Renal oncocytoma: A clinicopathologic study of 70 cases. *Am. J. Surg. Pathol.* **1997**, *21*, 871–883. [CrossRef] [PubMed]
- 106. Cole, A.P.; Garber, J.E.; Baniak, N.; Hirsch, M.S.; Lee Chang, S.; Kibel, A.S. 'Case of the Month' from Brigham and Women's Hospital, Boston, MA, USA: A 70-year-old man with lung cysts and bilateral renal masses. *BJU Int.* 2020, 126, 428–432. [CrossRef] [PubMed]
- 107. Johannesma, P.C.; Reinhard, R.; Kon, Y.; Sriram, J.D.; Smit, H.J.; van Moorselaar, R.J.; Menko, F.H.; Postmus, P.E.; Amsterdam BHD Working Group. Prevalence of Birt-Hogg-Dube syndrome in patients with apparently primary spontaneous pneumothorax. *Eur. Respir. J.* 2015, 45, 1191–1194. [CrossRef]
- Adley, B.P.; Smith, N.D.; Nayar, R.; Yang, X.J. Birt-Hogg-Dube syndrome: Clinicopathologic findings and genetic alterations. *Arch. Pathol. Lab. Med.* 2006, 130, 1865–1870. [CrossRef] [PubMed]
- Furuya, M.; Hong, S.B.; Tanaka, R.; Kuroda, N.; Nagashima, Y.; Nagahama, K.; Suyama, T.; Yao, M.; Nakatani, Y. Distinctive expression patterns of glycoprotein non-metastatic B and folliculin in renal tumors in patients with Birt-Hogg-Dube syndrome. *Cancer Sci.* 2015, 106, 315–323. [CrossRef] [PubMed]
- Li, J.; Liu, F.; Liu, X.; Hu, Y.; Liu, Z.; Shen, Y.; Wan, J. Heterozygous germline FLCN mutation in Birt-Hogg-Dube syndrome with bilateral renal hybrid oncocytic/chromophobe tumor and unilateral renal chromophobe cell carcinoma: A case report. *J. Cancer Res. Clin. Oncol.* 2023, 149, 2319–2325. [CrossRef]
- 111. Durinck, S.; Stawiski, E.W.; Pavia-Jimenez, A.; Modrusan, Z.; Kapur, P.; Jaiswal, B.S.; Zhang, N.; Toffessi-Tcheuyap, V.; Nguyen, T.T.; Pahuja, K.B.; et al. Spectrum of diverse genomic alterations define non-clear cell renal carcinoma subtypes. *Nat. Genet.* 2015, 47, 13–21. [CrossRef]
- 112. Davis, C.F.; Ricketts, C.J.; Wang, M.; Yang, L.; Cherniack, A.D.; Shen, H.; Buhay, C.; Kang, H.; Kim, S.C.; Fahey, C.C.; et al. The somatic genomic landscape of chromophobe renal cell carcinoma. *Cancer Cell* 2014, 26, 319–330. [CrossRef] [PubMed]
- 113. Petersson, F.; Gatalica, Z.; Grossmann, P.; Perez Montiel, M.D.; Alvarado Cabrero, I.; Bulimbasic, S.; Swatek, A.; Straka, L.; Tichy, T.; Hora, M.; et al. Sporadic hybrid oncocytic/chromophobe tumor of the kidney: A clinicopathologic, histomorphologic, immunohistochemical, ultrastructural, and molecular cytogenetic study of 14 cases. *Virchows Arch.* 2010, 456, 355–365. [CrossRef] [PubMed]

- 114. Moch, H.; Cubilla, A.L.; Humphrey, P.A.; Reuter, V.E.; Ulbright, T.M. The 2016 WHO Classification of Tumours of the Urinary System and Male Genital Organs-Part A: Renal, Penile, and Testicular Tumours. *Eur. Urol.* **2016**, *70*, 93–105. [CrossRef]
- 115. Montezuma, D.; Jeronimo, C.; Henrique, R. Is it "hybrid" or "intermediate"?-more than just a semantic issue in oncocytic renal cell tumors. *Ann. Transl. Med.* 2019, 7 (Suppl. 8), S356. [CrossRef]
- 116. Wobker, S.E.; Williamson, S.R. Modern Pathologic Diagnosis of Renal Oncocytoma. J. Kidney Cancer VHL 2017, 4, 1–12. [CrossRef]
- 117. Zhang, Y.; Narayanan, S.P.; Mannan, R.; Raskind, G.; Wang, X.; Vats, P.; Su, F.; Hosseini, N.; Cao, X.; Kumar-Sinha, C.; et al. Single-cell analyses of renal cell cancers reveal insights into tumor microenvironment, cell of origin, and therapy response. *Proc. Natl. Acad. Sci. USA* 2021, 118, e2103240118. [CrossRef]
- 118. Jikuya, R.; Murakami, K.; Nishiyama, A.; Kato, I.; Furuya, M.; Nakabayashi, J.; Ramilowski, J.A.; Hamanoue, H.; Maejima, K.; Fujita, M.; et al. Single-cell transcriptomes underscore genetically distinct tumor characteristics and microenvironment for hereditary kidney cancers. *iScience* 2022, 25, 104463. [CrossRef] [PubMed]
- Skala, S.L.; Wang, X.; Zhang, Y.; Mannan, R.; Wang, L.; Narayanan, S.P.; Vats, P.; Su, F.; Chen, J.; Cao, X.; et al. Next-generation RNA Sequencing-based Biomarker Characterization of Chromophobe Renal Cell Carcinoma and Related Oncocytic Neoplasms. *Eur. Urol.* 2020, 78, 63–74. [CrossRef]
- 120. Metwalli, A.R.; Linehan, W.M. Nephron-sparing surgery for multifocal and hereditary renal tumors. *Curr. Opin. Urol.* 2014, 24, 466–473. [CrossRef]
- 121. Duffey, B.G.; Choyke, P.L.; Glenn, G.; Grubb, R.L.; Venzon, D.; Linehan, W.M.; Walther, M.M. The relationship between renal tumor size and metastases in patients with von Hippel-Lindau disease. *J. Urol.* **2004**, 172, 63–65. [CrossRef]
- 122. Hasumi, Y.; Baba, M.; Ajima, R.; Hasumi, H.; Valera, V.A.; Klein, M.E.; Haines, D.C.; Merino, M.J.; Hong, S.B.; Yamaguchi, T.P.; et al. Homozygous loss of BHD causes early embryonic lethality and kidney tumor development with activation of mTORC1 and mTORC2. *Proc. Natl. Acad. Sci. USA* **2009**, *106*, 18722–18727. [CrossRef] [PubMed]
- 123. Nakamura, M.; Yao, M.; Sano, F.; Sakata, R.; Tatenuma, T.; Makiyama, K.; Nakaigawa, N.; Kubota, Y. A case of metastatic renal cell carcinoma associated with Birt-Hogg-Dube syndrome treated with molecular-targeting agents. *Hinyokika Kiyo* 2013, 59, 503–506. [PubMed]
- 124. Baba, M.; Keller, J.R.; Sun, H.W.; Resch, W.; Kuchen, S.; Suh, H.C.; Hasumi, H.; Hasumi, Y.; Kieffer-Kwon, K.R.; Gonzalez, C.G.; et al. The folliculin-FNIP1 pathway deleted in human Birt-Hogg-Dube syndrome is required for murine B-cell development. *Blood* 2012, 120, 1254–1261. [CrossRef] [PubMed]
- 125. Napolitano, G.; Di Malta, C.; Esposito, A.; de Araujo, M.E.G.; Pece, S.; Bertalot, G.; Matarese, M.; Benedetti, V.; Zampelli, A.; Stasyk, T.; et al. A substrate-specific mTORC1 pathway underlies Birt-Hogg-Dube syndrome. *Nature* 2020, 585, 597–602. [CrossRef]
- 126. Hasumi, H.; Baba, M.; Hasumi, Y.; Lang, M.; Huang, Y.; Oh, H.F.; Matsuo, M.; Merino, M.J.; Yao, M.; Ito, Y.; et al. Folliculininteracting proteins Fnip1 and Fnip2 play critical roles in kidney tumor suppression in cooperation with Flcn. *Proc. Natl. Acad. Sci. USA* 2015, 112, E1624–E1631. [CrossRef]
- 127. Hartman, T.R.; Nicolas, E.; Klein-Szanto, A.; Al-Saleem, T.; Cash, T.P.; Simon, M.C.; Henske, E.P. The role of the Birt-Hogg-Dube protein in mTOR activation and renal tumorigenesis. *Oncogene* **2009**, *28*, 1594–1604. [CrossRef]
- 128. Chen, J.; Huang, D.; Rubera, I.; Futami, K.; Wang, P.; Zickert, P.; Khoo, S.K.; Dykema, K.; Zhao, P.; Petillo, D.; et al. Disruption of tubular Flcn expression as a mouse model for renal tumor induction. *Kidney Int.* **2015**, *88*, 1057–1069. [CrossRef]
- 129. Wu, M.; Si, S.; Li, Y.; Schoen, S.; Xiao, G.Q.; Li, X.; Teh, B.T.; Wu, G.; Chen, J. Flcn-deficient renal cells are tumorigenic and sensitive to mTOR suppression. *Oncotarget* **2015**, *6*, 32761–32773. [CrossRef] [PubMed]
- 130. Chen, J.; Futami, K.; Petillo, D.; Peng, J.; Wang, P.; Knol, J.; Li, Y.; Khoo, S.K.; Huang, D.; Qian, C.N.; et al. Deficiency of FLCN in mouse kidney led to development of polycystic kidneys and renal neoplasia. *PLoS ONE* **2008**, *3*, e3581. [CrossRef]
- 131. Ko, E.J.; Cui, S.; Shin, Y.J.; Lim, S.W.; Lee, K.I.; Lee, J.Y.; Yang, C.W.; Kim, M.; Chung, B.H. Generation of the human induced pluripotent stem cell lines (CMCi009-A) from a patient with Birt-Hogg-Dube syndrome (BHD) with heterozygous frameshift deletion mutation c.1285delC of the FLCN gene. *Stem Cell Res.* **2021**, *51*, 102215. [CrossRef]
- 132. Zhang, Q.; Xu, Y.; Zhang, Z.; Li, J.; Xia, Q.; Chen, Y. Folliculin deficient renal cancer cells exhibit BRCA1 A complex expression impairment and sensitivity to PARP1 inhibitor olaparib. *Gene* **2021**, *769*, 145243. [CrossRef] [PubMed]
- Ruiz-Cordero, R.; Rao, P.; Li, L.; Qi, Y.; Atherton, D.; Peng, B.; Singh, R.R.; Kim, T.B.; Kawakami, F.; Routbort, M.J.; et al. Hybrid oncocytic/chromophobe renal tumors are molecularly distinct from oncocytoma and chromophobe renal cell carcinoma. *Mod. Pathol.* 2019, 32, 1698–1707. [CrossRef] [PubMed]
- Liu, Y.J.; Ussakli, C.; Antic, T.; Liu, Y.; Wu, Y.; True, L.; Tretiakova, M.S. Sporadic oncocytic tumors with features intermediate between oncocytoma and chromophobe renal cell carcinoma: Comprehensive clinicopathological and genomic profiling. *Hum. Pathol.* 2020, 104, 18–29. [CrossRef] [PubMed]
- 135. Mikami, S.; Kuroda, N.; Nagashima, Y.; Ohe, C.; Hayashi, H.; Mizuno, R.; Oya, M.; Kameyama, K. Classification of solid renal tumor with oncocytic/eosinophilic cytoplasm: Is hybrid oncocytic/chromophobe renal tumor a subtype of oncocytoma, chromophobe renal cell carcinoma, or a distinct tumor entity? *Ann. Transl. Med.* 2019, 7 (Suppl. 8), S350. [CrossRef] [PubMed]
- Pote, N.; Vieillefond, A.; Couturier, J.; Arrufat, S.; Metzger, I.; Delongchamps, N.B.; Camparo, P.; Mege-Lechevallier, F.; Molinie, V.; Sibony, M. Hybrid oncocytic/chromophobe renal cell tumours do not display genomic features of chromophobe renal cell carcinomas. *Virchows Arch.* 2013, 462, 633–638. [CrossRef]

- 138. Williams, M.J.; Werner, B.; Barnes, C.P.; Graham, T.A.; Sottoriva, A. Identification of neutral tumor evolution across cancer types. *Nat. Genet.* **2016**, *48*, 238–244. [CrossRef]
- 139. Craig, A.J.; von Felden, J.; Garcia-Lezana, T.; Sarcognato, S.; Villanueva, A. Tumour evolution in hepatocellular carcinoma. *Nat. Rev. Gastroenterol. Hepatol.* **2020**, *17*, 139–152. [CrossRef]

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.